



# FISHERY MANAGEMENT PLANS

FMP STATUS UPDATE MEMO

ESTUARINE STRIPED BASS FMP UPDATE



ROY COOPER  
*Governor*

MICHAEL S. REGAN  
*Secretary*

STEPHEN W. MURPHEY  
*Director*

Oct. 30, 2020

## MEMORANDUM

**TO:** N.C. Marine Fisheries Commission

**FROM:** Corrin Flora, Fishery Management Plan Coordinator  
Fisheries Management Section

**SUBJECT:** Fishery Management Plan Update

---

### Issue

Update the Marine Fisheries Commission (MFC) on the status of ongoing North Carolina fishery management plans (FMPs).

### Action Needed

For informational purposes only, **no action is needed at this time.**

### Overview

This memo provides an overview on the status of the North Carolina FMPs for the November 2020 MFC business meeting.

As noted at the MFC 's August 2019 business meeting, before the initial development of a draft FMP amendment, a scoping period will be held to notice the public that the review of the FMP is underway, inform the public of the stock status (if applicable), solicit input from the public on the list of potential management strategies to be developed, and recruit advisers to serve on the FMP advisory committee. The scoping process is concluding for estuarine striped bass and will be used for all future FMP reviews.

### Southern Flounder FMP

The MFC adopted Amendment 2 to the Southern Flounder FMP at its August 2019 business meeting and actions were taken to address the overfished and overfishing status of the southern flounder stock as determined by the 2019 coast-wide stock assessment. The season closures implemented under the authority of Amendment 2 were deemed critical to address overfishing and the successful rebuilding of the southern flounder stock, while other more comprehensive, long-term management strategies are examined and developed in Amendment 3.

The Southern Flounder FMP Advisory Committee is assisting the division with development of Amendment 3 to continue rebuilding the stock. Lead staff will provide a summary overview of progress at the November 2020 MFC business meeting on the progress of draft Amendment 3.

### **Shrimp FMP**

Staff continue to develop the first draft of the Shrimp FMP Amendment 2. The division is examining management strategies to promote habitat protection, further reductions of non-target species bycatch in the shrimp trawl fishery, and potential changes to existing shrimp management strategies adopted in previous plans. At its February 2020 business meeting, the MFC received a summary of the public comments submitted, received an overview of the potential management strategies and the FMP timeline, and approved the goal and objectives for Amendment 2. The goal adopted by the MFC is to manage the shrimp fishery to provide adequate resource protection, optimize long-term harvest, and minimize ecosystem impacts. Advisory Committee appointment process will begin before the end of the year.

### **Estuarine Striped Bass FMP**

The Estuarine Striped Bass (ESTB) FMP Plan Development Team recently completed the review of Amendment 1 and released the Central Southern Management Area (CSMA) Stock Report and the Albemarle-Roanoke River (A-R) Stock Assessment Report in August 2020. During the review process adaptive management under the current management plan, Amendment 1, was triggered, resulting in a Revision to Amendment 1. At the November 2020 MFC business meeting, lead staff will provide an overview of the Amendment 1 FMP review, including the CSMA and the A-R stock reports, and the recent Revision to Amendment 1.

With the review of Amendment 1 complete, development of Amendment 2 is underway, beginning with the scoping period being held Nov. 2-15, 2020. Results of the scoping period, the draft Goal and Objectives of Amendment 2, and a request for any additional management strategies to be considered, will be brought before the MFC at the Feb. 2021 business meeting.

### **Spotted Seatrout FMP**

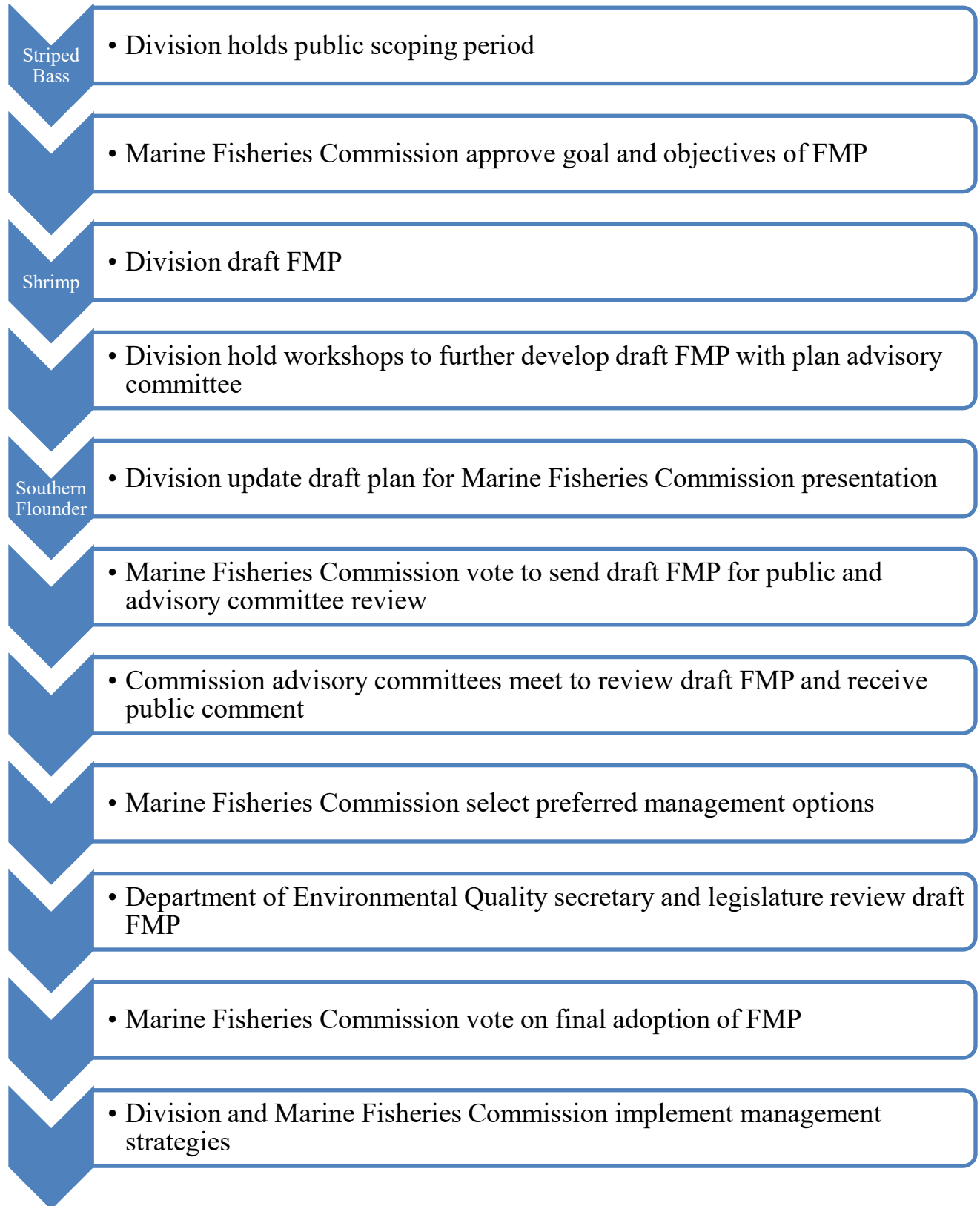
A benchmark stock assessment for spotted seatrout is underway coinciding with the scheduled Spotted Seatrout FMP review. The prior stock assessment from 2014 indicated that the stock is not overfished and is not experiencing overfishing. The Spotted Seatrout FMP Plan Development Team revisited the Data Workshop in October and incorporated data through 2019 to be more reflective of recent fishing activity. The benchmark stock assessment will be completed in 2021.

### **Striped Mullet FMP**

A benchmark stock assessment for striped mullet is underway coinciding with the scheduled Striped Mullet FMP review. The stock assessment update through terminal year 2017 indicated that the stock is not experiencing overfishing. Due to a poor relationship between spawning stock biomass and juvenile abundance, overfished status was unable to be determined. The Striped mullet FMP Plan Development Team will meet in Dec. 2020 for the stock assessment Planning Workshop.

# NORTH CAROLINA FISHERY MANAGEMENT PLANS

Nov. 2020





# ESTUARINE STRIPED BASS FMP

CENTRAL SOUTHERN MANAGEMENT AREA  
STRIPED BASS STOCKS IN NORTH CAROLINA, 2020

ALBEMARLE SOUND-ROANOKE RIVER STOCK  
ASSESSMENT AND ADAPTIVE MANAGEMENT MEMO

ASSESSMENT OF THE ALBEMARLE SOUND-ROANOKE  
RIVER STRIPED BASS IN NORTH CAROLINA, 1991-2017

NOVEMBER 2020 REVISION TO AMENDMENT 1 TO THE  
NORTH CAROLINA ESTUARINE STRIPED BASS FMP  
(ADAPTIVE MANAGEMENT)

SCOPING DOCUMENT: MANAGEMENT  
STRATEGIES FOR AMENDMENT 2



ROY COOPER  
*Governor*

MICHAEL S. REGAN  
*Secretary*

STEPHEN W. MURPHEY  
*Director*

Oct. 23, 2020

## MEMORANDUM

**TO:** N.C. Marine Fisheries Commission

**FROM:** Yan Li, Stock Assessment Scientist  
Todd Mathes, Biologist, Estuarine Striped Bass FMP Co-Lead  
Fisheries Management Section

**SUBJECT:** Central Southern Management Area Estuarine Striped Bass Stocks Report

---

### Issue

During review of the N.C. Estuarine Striped Bass Fishery Management Plan (FMP) begun by DMF and Wildlife Resources Commission (WRC), staff conducted an evaluation of Central Southern Management Area (CSMA) striped bass stocks. The results will inform development of Amendment 2. This memo provides a summary of key findings for the CSMA striped bass stocks that were based on the major analyses conducted by the division.

### Action Needed

For informational purposes only, **no action is needed at this time.**

### Findings

After reviewing available data, life history information, and stock assessment techniques, it was determined traditional stock assessment models are not appropriate for CSMA stocks because of the high hatchery contribution and lack of natural recruitment in these systems. A demographic matrix model was developed to evaluate different stocking and management measures for striped bass in all three CSMA river systems and an additional tagging model was developed to estimate striped bass abundance in the Cape Fear River.

The CSMA Estuarine Striped Bass Stocks report is a collection of (1) all data that have been collected, (2) all management effort, and (3) all major analyses that have been completed for CSMA stocks to serve as an aid in development of Amendment 2. As such:

- **Stock status could not be determined for CSMA striped bass.**
- **No biological reference points were generated for CSMA striped bass.**

### Matrix Model Overview - Tar-Pamlico, Neuse, and Cape Fear rivers

- Matrix model results indicate striped bass populations in the CSMA are depressed to an extent that sustainability is unlikely at any level of fishing mortality, given the current model assumptions.
- Survival and fertility of young fish influenced population growth rate more than older fish.
- Older fish contributed more than younger fish to reproduction due to higher egg production.
- Simulation of stocking and fishing strategies showed the population would likely benefit from stocking more fish.

### Tagging Model Overview - Cape Fear River

- Results showed a consistent decline in striped bass abundance in the Cape Fear River from 2012–2018.
- Abundance in 2018 was reduced to less than 20% of the abundance in 2012, even with a total no-possession provision for Cape Fear River striped bass since 2008.

***For more information, please refer to the full documents included in the Briefing Materials:***

- Central Southern Management Area Striped Bass Stocks in North Carolina, 2020

# **Central Southern Management Area Striped Bass Stocks in North Carolina, 2020**

T. Mathes, Y. Li, T. Tears, and L.M. Lee (editors)

August 2020



This document may be cited as:

Mathes, T., Y. Li, T. Tears, and L.M. Lee (editors). 2020. Central Southern Management Area striped bass stocks in North Carolina, 2020. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2020-02, Morehead City, North Carolina. 161 p. + appendices

## ACKNOWLEDGEMENTS

Members of the North Carolina Division of Marine Fisheries (NCDMF) Striped Bass Plan Development Team and their counterparts at the North Carolina Wildlife Resources Commission (NCWRC) were invaluable in providing assistance for the development of this stock assessment. Plan Development Team members from the NCDMF are Charlton Godwin (co-lead), Todd Mathes (co-lead), Katy West (mentor), Drew Cathey, Sean Darsee, David Dietz, Joe Facendola, Daniel Ipock, Laura Lee, Yan Li, Brian Long, Lee Paramore, Jason Peters, Jason Rock, Scott Smith, Chris Stewart, Thom Tears, Amanda Tong, Curt Weychert, and Chris Wilson. Members from the NCWRC are Jessica Baumann, Courtney Buckley, Kelsey Lincoln, Jeremy McCargo, Katy Potoka, Kyle Rachels, Ben Ricks, Kirk Rundle, Christopher Smith, and Chad Thomas. Thanks also to Kathy Rawls, NCDMF Fisheries Management Section Chief, and Catherine Blum, NCDMF Fishery Management Plan and Rulemaking Coordinator.

## EXECUTIVE SUMMARY

The North Carolina Fisheries Reform Act requires that fishery management plans be developed for the state's commercially and recreationally important species to achieve sustainable levels of harvest. Stock assessments are the primary tools used by managers to assist in determining the status of stocks and developing appropriate management measures to ensure the long-term viability of stocks.

This report represents a joint effort between the North Carolina Division of Marine Fisheries (NCDMF) and the North Carolina Wildlife Resources Commission (NCWRC). A working group of modelers, university researchers, and fishery biologists were brought together to review available data and to develop analyses that would address current management and research interests of Central Southern Management Area (CSMA) striped bass. The CSMA includes three major river systems: the Cape Fear, the Neuse, and the Tar-Pamlico. No stock status determination was performed for CSMA striped bass in this report and biological reference points were not generated due to continuous stocking effort and lack of understanding on the abiotic factors that are hindering the successful natural recruitment given the large number of fish stocked every year. This report is intended to be a collection of (1) all data that have been collected, (2) all management effort, and (3) all major analyses that have been completed for CSMA stocks. This report serves as a record of completed research efforts with implications for fishery management, and as a guide for future research effort based on results and identified data gaps.

A demographic matrix model was developed for striped bass in the three river systems in the CSMA. The matrix model was parameterized by synthesizing existing knowledge and data regarding striped bass, particularly the striped bass in the CSMA, from a literature review, data review and expert opinions. The population growth rate and the relative importance of life history parameters of each age group was estimated and evaluated. The demographic matrix model does not provide population abundance or mortality estimates. Possible stocking and fishery management strategies were evaluated using this matrix model. A tagging model was developed for striped bass in the Cape Fear River using tagging data collected by the NCDMF from 2012 to 2018. The total mortality and annual abundance for age 3–7 striped bass in the Cape Fear River were estimated by the tagging model.

Results from the matrix model indicated that striped bass populations in the CSMA are depressed to an extent that sustainability is unlikely at any level of fishing mortality, especially the assumptions associated with longevity (7 years for Cape Fear River and 11 years for Neuse and Tar-Pamlico Rivers) and age-0 survival (0.000017). Population growth rate was more dependent on survival and fertility of young fish than old fish. Reproductive contribution was most influenced by older age-classes due to higher fertility. Fishing activities typically select larger fish; thus, increases in fishing mortality disproportionately impact the abundance of older fish, constrict the age structure of the population, and limit reproductive contribution. Simulation on stocking and fishing strategies showed that population would likely benefit more from stocking more fish. Among the fishing strategies tested, the 10-year closure was most effective in increasing adult (age 3+) abundance over the entire 15-year simulation time period, and was also most effective in increasing old adult (age 6+) abundance during the first 13 years of simulation. Abundance of older fish (age 6+); however, quickly declined after the 10-year closure ended, and the 10-year closure strategy became less effective than the combo strategy in no stocking scenario, and less

effective than both the 26-inch size limit strategy and the combo strategy in stocking scenarios during the last two years of simulation.

Results of the tagging model showed a consistent decline in abundance estimates for striped bass in the Cape Fear River from 2012–2018. Abundance in 2018 was reduced to less than 20% of the abundance in 2012, even with a total no possession provision for striped bass in place in the Cape Fear River since 2008.

## TABLE OF CONTENTS

ACKNOWLEDGEMENTS .....	iii
EXECUTIVE SUMMARY .....	iv
TABLE OF CONTENTS.....	vi
LIST OF TABLES .....	viii
LIST OF FIGURES .....	xi
1 INTRODUCTION .....	1
1.1 The Resource .....	1
1.2 Life History .....	3
1.3 Habitat .....	8
1.4 Description of Fisheries.....	10
1.5 Fisheries Management.....	10
1.6 Assessment History .....	14
2 DATA .....	15
2.1 Fisheries-Dependent .....	15
2.2 Fisheries-Independent.....	21
3 DEMOGRAPHIC MATRIX MODEL .....	32
3.1 Objectives .....	32
3.2 Methods .....	33
3.3 Discussion.....	38
4 TAGGING MODEL .....	39
4.1 Objectives .....	39
4.2 Methods .....	39
4.3 Results .....	42
4.4 Discussion.....	42
5 GLM ANALYSIS ON COMMERCIAL & RECREATIONAL FISHERIES DATA .....	43
5.1 Objectives .....	43
5.2 Methods .....	44
5.3 Results .....	44
5.1 Discussion.....	45
6 YIELD-PER-RECRUIT .....	45
6.1 Objectives .....	45
6.2 Methods .....	46

6.3 Approach .....	52
6.4 Results .....	52
6.5 Discussion.....	53
7 AGE COMPARISON .....	54
7.1 Introduction .....	54
7.2 Objectives .....	55
7.3 Methods .....	55
7.4 Results .....	57
7.5 Discussion.....	58
7.6 Conclusion.....	59
7.7 Recommendation.....	59
8 RESEARCH RECOMMENDATIONS .....	59
9 LITERATURE CITED .....	61
10 TABLES .....	74
11 FIGURES.....	105
12 APPENDIX A.....	162
13 APPENDIX B.....	170

## LIST OF TABLES

Table 1.1.	Stocking numbers of Phase II (5–7 inches total length) striped bass by system and year for the Tar-Pamlico, Neuse, and Cape Fear rivers, 1980–2018. ....	74
Table 1.1.	<i>(continued)</i> Stocking numbers of Phase II (5–7 inches total length) striped bass by system and year for the Tar-Pamlico, Neuse, and Cape Fear rivers, 1980–2018. ....	75
Table 1.2.	Percent hatchery contribution from striped bass genetic samples collected in the Tar-Pamlico, Neuse, and Cape Fear rivers by NCDMF and NCWRC staff, 2013–2018. (Source: South Carolina Department of Natural Resources).....	76
Table 2.1.	Summary (mean, minimum, maximum and number of samples) striped bass length data (TL in inches) from CSMA commercial harvest, 2000–2018. ....	77
Table 2.2.	Commercial estimates of striped bass discards (standard error in parentheses) in the Tar-Pamlico/Pungo rivers by mesh size, 2013–2018. ....	78
Table 2.3.	Commercial estimates of striped bass discards (standard error in parentheses) in the Neuse/Bay rivers by mesh size, 2013–2018. ....	78
Table 2.4.	Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries. ....	79
Table 2.4.	<i>(continued)</i> Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries. ....	80
Table 2.4.	<i>(continued)</i> Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries. ....	81
Table 2.4.	<i>(continued)</i> Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries. ....	82
Table 2.5.	Annual weighted relative abundance index of striped bass (number of individuals per sample), total number of striped bass collected, and the number of gill net samples (n) in the Tar-Pamlico, Pungo, and Neuse rivers (2004–2018) and the Cape Fear and New rivers (2008–2018). The Percent Standard Error (PSE) represents a measure of precision of the index.....	83
Table 2.6.	NCWRC annual catch summary for the Tar River striped bass electrofishing survey, 1996–2018. ....	84
Table 2.7.	NCWRC annual catch summary for the Neuse River striped bass electrofishing survey, 1994–2018. ....	85
Table 2.8.	NCWRC annual catch summary for the Cape Fear River striped bass electrofishing survey, 2003–2018. ....	86
Table 2.9.	Total number of striped bass PIT tagged by all gears and tagger affiliation in the Cape Fear River, 2011–2018. ....	87
Table 2.10.	Total number of striped bass PIT tagged by gear and tagger affiliation included in the tagging model in the Cape Fear River, 2012–2018. ....	87
Table 2.11.	Mean, standard deviation (SD), minimum, and maximum total length (TL) of striped bass tagged by year, gear, and tagger affiliation included in the tagging model for the Cape Fear River, 2012–2018. ....	88
Table 2.12.	Total number of striped bass PIT tag recaptures by all gears in the Cape Fear River, 2011–2018. ....	88

Table 2.13.	Total number of striped bass PIT tag recaptures, from electrofishing gear, included in the tagging model for the Cape Fear River, 2012–2018. ....	89
Table 2.14.	Distance (miles) between release and recapture sites of striped bass included in the tagging model by days at large in the Cape Fear River, 2012–2018. ....	89
Table 2.15.	Mean, standard deviation (SD), minimum, and maximum number of days at large of striped bass recaptured by year, 2012–2018. ....	90
Table 2.16.	Mean, standard deviation (SD), minimum, and maximum total length (TL) of striped bass recaptured by year in the Cape Fear River, 2012–2018. ....	90
Table 2.17.	Mean, standard deviation (SD), minimum, and maximum growth (mm) of recaptured striped bass by days at large in the Cape Fear River, 2012–2018. ....	91
Table 2.18.	Mean, standard deviation (SD), minimum, and maximum growth (mm) of striped bass recaptured by year in the Cape Fear River, 2012–2018. ....	91
Table 3.1.	Summary of parameter values used to develop the demographic matrix model. ...	92
Table 3.1.	<i>(continued)</i> Summary of parameter values used to develop the demographic matrix model. ....	93
Table 3.2.	Initial year age structure for fishery management strategy evaluation. ....	94
Table 3.3.	Population growth rate estimates from the matrix model. Pr is the probability of population growth rate greater than one. ....	94
Table 4.1.	Cape Fear River tagging model parameters and priors. <i>U</i> denotes the uniform distribution. ....	95
Table 4.2.	Estimated instantaneous total mortality ( $Z$ , yr <sup>-1</sup> ) due to natural causes and fishing, estimated abundance (number) and estimated capture probability ( $\alpha$ ) from the tagging model in the Cape Fear River. Median—posterior median; Lower and Upper—lower and upper 95% credible intervals. ....	95
Table 4.3.	Estimated striped bass effort and catch in the Cape Fear River. (Source: Costal Angling Program (CAP) Central Southern Management Area (CSMA) recreational striped bass creel survey) ....	96
Table 4.3.	<i>(continued)</i> Estimated striped bass effort and catch in the Cape Fear River. (Source: Costal Angling Program (CAP) Central Southern Management Area (CSMA) recreational striped bass creel survey) ....	97
Table 5.1.	Fit of the candidate models. Com = commercial; Rec = recreational; DO = dissolved oxygen (mg/L); K = the number of parameters; AIC <sub>c</sub> = Akaike’s information criterion corrected for small sample size; $\Delta_i$ = Akaike difference; $w_i$ = Akaike weight. The candidate models from Rachels and Ricks (2018) are formatted in <b>bold</b> . ....	98
Table 6.1.	Estimated parameter values of the von Bertalanffy age-length relationship and their associated standard errors (SE) where total length was measured in millimeters (n=166). ....	99
Table 6.2.	Estimated parameter values of the length-weight relationship and their associated standard errors (SE) where total length was measured in millimeters and weight was measured in grams (n=198). ....	99
Table 6.3.	Estimated natural mortality ( $M$ ) at age based on Lorenzen’s (1996) approach. The values given represent instantaneous rates. ....	99



Table 6.4.	Estimated parameter values of the logistic length-maturity relationship and their associated standard errors (SE) where total length was measured in millimeters (n=170).....	99
Table 6.5.	Definitions of symbols used in the per-recruit equations. ....	100
Table 6.6.	Sample frequency at (genetic) age of striped bass collected in the Neuse River by the NCWRC's Spawning Stock Survey in 2017. Catches have been standardized to a collection time of 19 minutes.....	101
Table 6.7.	Estimates of fishing mortality ( $F$ ) at age derived from the catch curve analysis. ....	101
Table 7.1.	Number of scales, otoliths, and genetic (PBT) structures collected by NCDMF available for CSMA striped bass age determination, 1975–2018. Genetic (PBT) structures are only available from 2016–2018.....	102
Table 7.2.	Mean percentage age bias (bias compared to genetic age) for each reader for overall age bias and age bias by method type (standard deviation in parentheses). Cells with no values indicate the reader performed no readings for that method type.....	103
Table 7.3.	Parameter estimates from Bayesian generalized linear mixed effects model for scale ages and otolith ages compared to genetic ages. Estimates are median of posterior distributions with confidence interval in parentheses.....	103
Table 7.4.	Coefficient of variation (%) analyses results for between readers for scale ages. Values in parentheses are percent agreement. Values in <b>bold</b> are significant ( $P < 0.01$ ). Between reader coefficients of variation differ depending on which reader is the reference reader. ....	104

## LIST OF FIGURES

Figure 1.1.	Boundary lines between the Albemarle Sound Management Area, Central Southern Management Area, and the Roanoke River Management Area. ....	105
Figure 1.2.	CSMA striped bass length at age based on otolith and genetic age samples collected by NCDMF, 2004–2018. Blue circles represent the mean size at a given age while the grey squares represent the minimum and maximum observed size for each age. ....	106
Figure 2.1.	Commercial striped bass harvest in numbers and pounds and anchored gill-net trips in the Tar-Pamlico, Pungo, Neuse, and Bay rivers, 2004–2018. ....	107
Figure 2.2.	Commercial striped bass harvest by system, and the TAL in the CSMA, 2004–2018. *There has been a harvest moratorium in the Cape Fear River since 2008. **Landings data for the Pamlico Sound in 2012 are confidential. ....	107
Figure 2.3.	Length frequency of CSMA striped bass landed commercially in the Tar-Pamlico and Pungo rivers, 2004–2018. ....	108
Figure 2.4.	Length frequency of CSMA striped bass landed commercially in the Neuse and Bay rivers, 2004–2018. ....	109
Figure 2.5.	Program 466 CSMA observer trips by the presence or absence of striped bass, 2013–2018. The cross sign is an observer trip that encountered a striped bass (n=284), and the triangle is an observer trip that did not encounter striped bass (n=789). ....	110
Figure 2.6.	Program 466 CSMA observer trips by mesh size, 2013–2018. The square is a small mesh observer trip that encountered striped bass (n=38), and the circle is a large mesh observer trip that encountered striped bass (n=246). Eight large mesh observer trips accounted for 37 striped bass that are not presented on the map due the absence of coordinates. ....	111
Figure 2.7.	Recreational striped bass harvest in numbers and pounds and effort in angler hours for the Tar-Pamlico and Neuse rivers and tributaries, 2004–2018. ....	112
Figure 2.8.	Recreational striped bass harvest in the Tar-Pamlico, Pungo, and Neuse rivers, 2004–2018. ....	112
Figure 2.9.	Annual recreational catch (released and/or harvested) of striped bass in the CSMA, 2004–2018. ....	113
Figure 2.10.	Length frequency of CSMA striped bass recreationally harvested in the Tar-Pamlico and Pungo rivers, 2004–2018. ....	114
Figure 2.11.	Length frequency of CSMA striped bass recreationally harvested in the Neuse River, 2004–2018. ....	115
Figure 2.12.	Location of Central Southern Management Area (CSMA) juvenile striped bass beach seine and trawl sites, Tar-Pamlico and Neuse rivers, NC. ....	116
Figure 2.13.	Location of Cape Fear River juvenile striped bass beach seine and trawl sites, CapeFear River, NC. ....	118
Figure 2.14.	The sample regions and grid system for P915 in Dare (Region 1) and Hyde (Region 2) counties. ....	118
Figure 2.15.	The sample areas and grid system for P915 in the Pamlico Region (Pamlico, Pungo and Neuse rivers) with areas numbered Pamlico/Pungo: 1—Upper, 2—	

	Middle, 3— Lower, 4—Pungo; Neuse: 1—Upper, 2—Upper-middle, 3— Lower-middle, and 4—Lower). .....	119
Figure 2.16.	The sample areas and grid system for P915 in the Central Region with areas numbered (1—West Bay/Upper Core Sound, 2—Lower Core Sound, 3— Newport River/Bogue Sound, and 4—Bogue Sound/White Oak River). Sampling began May 2018. ....	120
Figure 2.17.	The sample areas and grid system for P915 in the Southern Region (New and Cape Fear rivers) with areas numbered (New: 1—Upper, 2—Lower, Cape Fear). ....	121
Figure 2.18.	Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2004–2018 (Tar-Pamlico River, shallow sets, April and October–November). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index. ....	122
Figure 2.19.	Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2004–2018 (Neuse River, shallow sets, April and October–November). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index. ....	122
Figure 2.20.	Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2008–2018 (Cape Fear River, shallow sets). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index. ....	123
Figure 2.21.	Length frequency distribution of CSMA striped bass captured in P915 in the Tar-Pamlico River, 2004–2019 (deep and shallow sets, April and October– November). ....	124
Figure 2.22.	Length frequency distribution of CSMA striped bass captured in P915 the Neuse River, 2004–2019 (deep and shallow sets, April and October– November). ....	125
Figure 2.23.	NCWRC electrofishing survey segments on the Tar-Pamlico River.....	126
Figure 2.24.	NCWRC electrofishing survey area on the Neuse River. The upstream and downstream extent of four sampling strata are by colored markers. ....	127
Figure 2.25.	NCWRC electrofishing sampling sites (indicated by black circles in bold) at Lock and Dams 1, 2, 3, and Buckhorn Dam on the Cape Fear River.....	128
Figure 2.26.	Relative abundance (with associated standard error) of striped bass collected during the NCWRC Tar River electrofishing surveys, 1996–2018. ....	129
Figure 2.27.	Length distributions for striped bass collected during the NCWRC Tar River electrofishing surveys, 1996–2018. Dots indicate individual length measurements.....	130
Figure 2.28.	Relative abundance (with associated standard error) of striped bass collected during the NCWRC Neuse River electrofishing surveys, 1994–2018. ....	131
Figure 2.29.	Striped bass length distributions for the NCWRC Neuse River electrofishing surveys, 1994–2018. Dots indicate individual length measurements. ....	132
Figure 2.30.	Relative abundance (with associated standard error) of striped bass collected at three sample sites in the Cape Fear River, NC, 2003–2018. ....	133

Figure 2.31. Length distributions for striped bass collected during the NCWRC Cape Fear River electrofishing surveys, 2003–2018. Dots indicate individual length measurements.....	134
Figure 2.32. Cape Fear River striped bass tagging and recapture locations, 2012–2018. ....	135
Figure 2.33. Length-frequency distribution of tagged striped bass included in the tagging model by tagger affiliation in the Cape Fear River, 2012–2018.....	136
Figure 2.34. Genetically derived age at length of Cape Fear River striped bass, 2016–2017...	136
Figure 2.35. Length-frequency distribution of recaptured striped bass included in the tagging model by tagger affiliation in the Cape Fear River, 2012–2018.....	137
Figure 3.1. Age-specific natural mortality and fertility used in the matrix model. Black line is median and grey area is 95% confidence interval.....	138
Figure 3.2. Elasticity of population growth rate to survival and fertility and age-specific reproduction contribution. Lines represent various fishing mortality ( $F$ ) values. Lines show the median from 10,000 iterations.....	139
Figure 3.3. Sensitivity of population growth rate to viable egg proportion ( $x$ ), age-0 survival ( $S_0$ ) and the asymptotic length ( $L_\infty$ ). Lines represent various fishing mortality ( $F$ ) values. Lines show the median from 10,000 iterations.....	140
Figure 3.4. Abundance of adults (age 3+) projected under five stocking strategies and six fishing strategies. Stocking 1—no stocking; Stocking 2—stocking 100,000 fish per year with 2-year stocking and 2-year no stocking alternating for 15 years (8 years of stocking in total); Stocking 3—stocking 500,000 fish per year with 2-year stocking and 2-year no stocking alternating for 15 years (8 years of stocking in total); Stocking 4—stocking 100,000 fish per year with 8-year continuous stocking; Stocking 5—stocking 500,000 fish per year with 8-year continuous stocking. Lines show the median from 10,000 iterations.....	141
Figure 3.5. Abundance of old adults (age 6+) projected under five stocking strategies and six fishing strategies. Lines show the median from 10,000 iterations. See Figure 3.4 caption for explanation of the five stocking strategies. ....	142
Figure 4.1. Estimated instantaneous total mortality ( $Z$ , $\text{yr}^{-1}$ ) due to natural causes and fishing, estimated abundance ( $N$ , number) and estimated capture probability ( $\alpha$ ) from the tagging model. Line is posterior median and shaded area is 95% credible interval. ....	143
Figure 4.2. Posterior distributions of annual abundance estimated using a Jolly-Seber model and capture probabilities estimated by the multistate model in the Cape Fear River. The whiskers of the boxplots indicate 95% credible intervals of the estimates; boxes of the boxplots represent 50% credible intervals and the bolded lines of each boxplot represent abundance estimates. (Source: Collier et al. 2013) .....	144
Figure 4.3. NCDMF recreational creel survey estimated striped bass discards (number; dotted line) and recreational fishing effort (hours; solid line) in the Cape Fear River, 2013–2018. In 2013, due to comparatively low recreational striped bass catch, American and hickory shad became the target species. ....	144
Figure 4.4. Dead striped bass at Battleship Park, Wilmington, NC following extensive flooding from Hurricane Florence in September 2018.....	145

Figure 5.1.	Important factors selected in the model when using data from (A) 1994–2015, and (B) data from 2004–2015 without considering recreational information, and (C) when using data from 2004–2015 with recreational information included. These factors are listed in the order of importance from the most important to the least important ones. See the caption of Table 1 for abbreviations of the predictor variables.....	147
Figure 6.1.	Sampling sites in the Neuse River for the NCWRC’s Spawning Stock Survey...	148
Figure 6.2.	Range of sampling times for individual sampling trips from the NCWRC’s Spawning Stock Survey in 2017.....	149
Figure 6.3.	Observed (black circles) and predicted (blue line) values of the von Bertalanffy age-length relationship.....	149
Figure 6.4.	Observed (open black circles) and predicted (blue line) values of the length-weight relationship.....	150
Figure 6.5.	Estimated natural mortality at age based on Lorenzen’s (1996) approach. The values shown represent instantaneous rates.....	151
Figure 6.6.	Observed (grey circles) and predicted (red line) values of the length-maturity relationship. The blue plus signs represent the proportion mature for selected length categories.....	151
Figure 6.7.	Observed (grey circles) and predicted (black line) values of the length-fecundity relationship for non-hatchery origin fish.....	152
Figure 6.8.	Observed (grey circles) and predicted (black line) values of the length-fecundity relationship for hatchery-origin fish.....	152
Figure 6.9.	Selectivity at age assumed in the per-recruit analyses.....	153
Figure 6.10.	Yield per recruit in terms of weight (kilograms) at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.....	153
Figure 6.11.	Yield per recruit in terms of numbers at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.....	154
Figure 6.12.	Spawning potential ratio (%SPR) at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.....	154
Figure 6.13.	Yield per recruit in terms of weight (kilograms) over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.....	155
Figure 6.14.	Yield per recruit in terms of numbers over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.....	155
Figure 6.15.	Spawning potential ratio (%SPR) over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.....	156
Figure 7.1.	Boxplot of percentage age bias by reader ID. The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data.....	157
Figure 7.2.	Boxplot of percentage age bias by ageing method. The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data.....	158

Figure 7.3. Percentage age bias by genetic age (from parental base tagging) with trend line (solid line). The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data. .... 159

Figure 7.4. Posterior distributions for three chains of parameter estimates from Bayesian generalized linear mixed effects model. Alpha's represent reader effects, gamma's represent method effects, mu represents the overall average bias, pct1 represents percentage of error explained by random error, pct2 represents percentage of error explained by reader effects, sigma1 represents standard deviation associated with random error, sigma2 represents standard deviation associated with reader effects, and deviance is a goodness-of-fit estimate. .... 160

Figure 7.5. Contingency table for number of fish in each scale age for each otolith age. Numbers represent number of fish assigned scale age for a given otolith age. .... 161

Figure 7.6. Age-bias plot for average scale age for each otolith age with standard deviation. .... 161

# 1 INTRODUCTION

## 1.1 The Resource

The common and scientific names for the species are striped bass, *Morone saxatilis* (Walbaum; Robins et al. 1991). In North Carolina, it is also known as striper, rockfish, or rock. Striped bass naturally occur in fresh, brackish, and marine waters from Canada to the Gulf of Mexico. Due to their annual spawning migrations into freshwater, striped bass have been the focus of fisheries from North Carolina to New England for several centuries and have played an integral role in the development of numerous coastal communities. Striped bass regulations in the United States date to pre-Colonial times (circa 1640) when striped bass were prohibited from being used as fertilizer. Striped bass populations south of Cape Hatteras, North Carolina are considered to have a primarily endemic riverine life history, having limited or no adult oceanic migration (Setzler et al. 1980; Rulifson et al. 1982a; Callihan 2012).

Various levels of stocking have occurred in the Central Southern Management Area (CSMA; Tar-Pamlico, Neuse and Cape Fear rivers) since the 1940s (Bayless and Smith 1962; Woodroffe 2011), with the North Carolina Division of Marine Fisheries (NCDMF's) formal involvement beginning in 1980 as a result of a cooperative agreement with the United States Fish and Wildlife Service (Table 1.1; NCDMF and NCWRC 2013). The North Carolina Wildlife Resources Commission (NCWRC) was added to the cooperative agreement in 1986 (NCDMF 2013) but has been involved in the CSMA striped bass stocking program since fry stocking began in Neuse River tributaries in 1949 (Bayless and Smith 1965). The practice of cross-stocking (stocking of striped bass from one drainage system to another, e.g., Roanoke River striped bass offspring being stocked throughout the southeastern United States) has introduced non-endemic genetic strains to many striped bass populations. The effects of this long-standing practice remain largely undocumented and unquantified (Rulifson and Laney 1999; Bergey et al. 2003).

A management strategy adopted in the North Carolina Estuarine Striped Bass Fisheries Management Plan (FMP) Amendment 1 continued the annual stocking program in the CSMA rivers. Specific objectives for stocking striped bass included attempts to increase spawning stock abundance while promoting self-sustaining population levels appropriate for various habitats (see Amendment 1, Section 11.2 Striped Bass Stocking in Coastal Rivers, NCDMF 2013). The management strategy from Amendment 1 increased the annual numbers stocked to a goal of 100,000 hatchery reared striped bass in each of the major river systems (Tar-Pamlico, Neuse, and Cape Fear rivers) to aid in recovery of the stocks. From 2004 to 2009, stocking occurred on a rotating basis where only two out of the three systems were stocked annually. Prior to 2004, stocking was focused on the Tar-Pamlico and Neuse rivers with sporadic stocking in the Cape Fear River (Table 1.1).

Prior to 2010, the otoliths of hatchery-reared striped bass were chemically marked with oxytetracycline to determine the percent contribution of hatchery fish to the wild population. Results from the chemical marking methodology suggested hatchery-reared striped bass contributed little to the spawning populations in the CSMA (0 to 31%; Barwick et al. 2008); however, since the adoption of Amendment 1, researchers have realized the chemical mark was not being retained in 100% of fish (73%; Barwick et al. 2008), which led to underestimation of the percent of hatchery reared fish in the striped bass populations in the CSMA (Barwick et al. 2008; NCDMF 2013).

In 2010, the NCWRC implemented parentage-based tagging (PBT) as a more accurate method to determine percent hatchery contribution of the striped bass spawning populations in the CSMA. This method utilizes genetic marking techniques and has proven to be greater than 99% accurate at determining if an individual fish was hatchery produced or not (Denson et al. 2012). In 2016, the NCDMF started collecting striped bass fin clip samples for PBT analysis from the commercial and recreational fisheries and from areas away from the spawning grounds in the lower portions of the rivers to gain additional spatial coverage of samples. Since 2011, PBT analysis of samples collected on the spawning grounds and in internal coastal fishing waters of the Tar-Pamlico, Neuse, and Cape Fear rivers has revealed hatchery-stocked striped bass can comprise up to 90% of the fish sampled in some years (O'Donnell and Farrae 2017); however, PBT results from fish sampled in 2017 revealed a noticeable decrease in contribution of hatchery-stocked fish (Farrae and Darden 2018). In 2017 and 2018, percentages of hatchery fish were much lower for the 2014 and 2015 year classes in NCDMF samples (63% and 41%, respectively) and NCWRC samples (76% and 77%, respectively).

While attempts have been made to use catch curves to assess the stock status of CSMA striped bass (NCDMF 2004, 2013) no peer-reviewed stock assessment has been conducted. The catch-curve analysis conducted in 2003 determined the stock was experiencing overfishing (NCDMF 2004), although it was not used for management; however, a repeat of that analysis in 2010, concluded stock status could not be determined due to uncertainty in the mortality estimates (NCDMF 2013). Therefore, striped bass in the Tar-Pamlico and Neuse rivers have an unknown stock status. The need for continued conservation management efforts has been supported by persistent low overall abundance, minimal natural recruitment, multiple sources of mortality, the absence of older fish on the spawning grounds, non-optimal environmental conditions on the spawning grounds in the spring, potential impacts from stocked juveniles and hybrid striped bass, and the high percentage of stocked fish in the population in most years.

### **1.1.1 Stock Definitions**

There are two geographic management units (northern and southern) and four striped bass stocks inhabiting the estuarine and inland waters of North Carolina. The CSMA is located in the southern geographic management unit and includes all internal coastal, joint, and contiguous inland waters of North Carolina south of the Albemarle Sound Management Area (ASMA) to the South Carolina state line (Figure 1.1). There are spawning stocks in each of the major river systems within the CSMA (Tar-Pamlico River stock, Neuse River stock, and Cape Fear River stock). Spawning grounds are not clearly defined in these systems as access to spawning areas is influenced by river flows and impediments to migration. Management of striped bass within the CSMA is the sole responsibility of the North Carolina Marine Fisheries Commission (NCMFC; coastal and joint fishing waters) and the NCWRC (joint and inland waters) and is not subject to compliance with the ASMFC Interstate FMP for Atlantic Ocean striped bass.

This report focuses on the analyses performed for the striped bass in the CSMA. After reviewing available data, life history information, and stock assessment techniques, it was determined traditional stock assessment models would not be appropriate for CSMA stocks because of the high hatchery contribution and lack of natural recruitment in these systems.



## **1.2 Life History**

### **1.2.1 Movements & Migration**

Striped bass populations in the Tar-Pamlico, Neuse, and the Cape Fear rivers have been considered to have a primarily endemic riverine life history having limited or no adult oceanic migration (Setzler et al. 1980; Rulifson et al. 1982a). Tagging data have indicated there is some movement of striped bass from the Neuse and Pamlico rivers into other systems and the Atlantic Ocean, but this is at low levels (Callihan 2012; Callihan et al. 2014; Rock et al. 2018). Tag-return data from stocked striped bass (Phase II; 5–7 inch total length, TL) suggest that these fish contribute to the commercial and recreational fisheries as well as the spawning stock in the Neuse and Tar rivers but do not commonly migrate to other rivers (Winslow 2007). Acoustic tagging studies within the Cape Fear River Basin demonstrated adult fish making seasonal spawning migrations within the drainage; however, emigration out of the system was minimal (Rock et al. 2018; Prescott 2019). Many striped bass exhibited a pattern of residency in the lower portions of the Tar-Pamlico and Neuse rivers with some detected making multiple seasonal spawning runs with many moving as far upstream as Rocky Mount in the Tar River and Raleigh in the Neuse River (Rock et al. 2018).

### **1.2.2 Age & Size**

Striped bass scales have been collected by the NCDMF since 1975, and otoliths have been collected since 2003. Striped bass otoliths have been documented to provide more accurate and precise age estimates than scales (Humphreys and Kornegay 1985; Boyd 2011; Liao et al. 2013) and that ageing error can bias results of stock assessments. In 2017, the NCDMF compared scale and otolith ages from multiple readers for known age striped bass and found age estimates from scales to be unreliable and commonly underage or overage CSMA striped bass; as a result, only otolith ages are considered in this assessment (see section 7). Additionally, in 2016 and 2017 genetic samples were collected by the NCDMF from striped bass that allowed for age determination of hatchery-produced fish that were used in this analysis. The NCWRC used scales to age Tar-Pamlico River striped bass from 1996–2012 and Neuse River striped bass from 1994–2012. Since the inception of the PBT program in 2010, the NCWRC has determined ages of hatchery-produced fish using PBT analysis and used scales when PBT ages were not available. The NCWRC does not routinely collect striped bass otoliths, and did not provide any otolith ages for this assessment. Based on otolith and PBT age data collected from 2004 to 2017 (Figure 1.2), a maximum age of 11 years has been observed for striped bass in the Tar-Pamlico and Neuse rivers and a maximum age of seven years has been observed for striped bass in the Cape Fear River. Fish older than age eight years are rare in all of the CSMA river systems; however, NCWRC scale-aged fish suggest greater maximum ages in all CSMA rivers (Homan et al. 2010; Fisk and Morgeson 2016). This report found that ageing biases from scale ages resulted in underestimates of population abundance (15%) and female spawning stock biomass (19%), while overestimating fishing mortality in the terminal year (19%) and made strong age-1 recruitment years appear weaker and weak ones stronger.

### **1.2.3 Growth**

As a relatively long-lived species, striped bass (approaching 30 years) can attain a moderately large size. Females grow to a considerably larger size than males; striped bass over 30 pounds are almost exclusively female (Bigelow and Schroeder 1953; NCDMF, unpublished data). Growth

occurs between April and October. During the spawning migration, striped bass stop feeding for a brief period just before and during spawning, however feeding continues during the upriver spawning migration and resumes soon after spawning (Trent and Hassler 1968). From November through March, striped bass growth is thought to be negligible.

Striped bass in the CSMA grow at a faster rate and have a greater total length at age compared to the A-R stock (Knight 2015) and Neuse River striped bass exhibit the fastest growth rate in the CSMA (NCDMF 2020). As an example, in 2017, mean length of age-5 female striped bass in the Roanoke River was 559 mm TL while Neuse River female mean length at age 5 was 634 mm TL (Ricks and Buckley 2018; Smith and Potoka 2018). Fast growth in CSMA rivers has been attributed to a lack of density-dependent forage limitations (Ricks and Buckley 2018). This is possibly attributed to superior growth in the initial year of life for hatchery fish compared to wild fish, abundant food availability, and relatively small population. In addition, a tagging study showed striped bass stocked in the Neuse and Tar-Pamlico rivers had a higher growth rate (growth coefficient of 0.54–0.61 per year) than in their natal habitat (Roanoke River; Callihan et al. 2014).

#### **1.2.4 Reproduction**

Striped bass spawn in freshwater or nearly freshwater portions of North Carolina’s coastal rivers from late March to June depending upon water temperatures (Hill et al. 1989). Spawning behavior is characterized by brief peaks of surface activity when a mature female is surrounded by up to 50 males as eggs are broadcast into the surrounding water and males release sperm, termed “rock fights” by locals (Setzler et al. 1980). Spawning by a given female is probably completed within a few hours (Lewis and Bonner 1966).

Based on data collected on the Tar-Pamlico River in 2004 and 2005, the peak spawning activity was observed in April through mid-May (Smith and Rulifson 2015) and acoustic detection data in the Neuse River shows striped bass were only in the upper portions of the river from March through May (Rock et al. 2018). Despite an apparent spawning migration, and NCWRC surveys that have documented limited numbers of striped bass eggs in various stages of development in the Tar-Pamlico and Neuse rivers (Jones and Collart 1997; Smith and Rulifson 2015), the stocks remain comprised of predominantly hatchery origin fish (Farrae 2019; Table 1.2).

Studies have collected eggs, larvae, juveniles (Winslow et al. 1983; Smith 2009; Smith and Hightower 2012; Morgeson and Fisk 2018), or adult fish (Ashley and Rachels 2006) to show evidence of spawning and/or spawning migrations in the main stem of the Cape Fear River.

##### **1.2.4.1 Eggs**

Mature eggs are 1.0–1.5 mm (0.039 to 0.059 inch) in diameter when spawned and remain viable for about one hour before fertilization (Stevens 1966). Fertilized eggs are spherical, non-adhesive, semi-buoyant, and nearly transparent. Fertilized eggs need to drift downstream with currents to hatch into larvae. If the egg sinks to the bottom, the chances of hatching are reduced because the sediments reduce oxygen exchange between the egg and the surrounding water. After hatching, larvae are carried by the current to the downstream nursery areas.

There is some discrepancy over temperature tolerance for striped bass eggs. Morgan and Rasin (1973) and Rogers et al. (1977) indicated that egg survival gradually declines as temperature drops below 17°C and rapidly declines as water temperature approaches 23°C. In general, lower temperatures lead to longer incubation periods (Hardy 1978). Bain and Bain (1982) documented

hatching at approximately 48 hours after fertilization at a temperature of 18°C, and other studies have shown that hatching time varied from 29 hours at 22°C to 80 hours at 11°C (Mansueti 1958; Hardy 1978). Hassler et al. (1981) found that A-R striped bass eggs hatch in 38 hours. Sampling by the NCWRC in 1965 and 1975 indicated striped bass spawning occurs in the Tar-Pamlico River from mid-April to mid-May with peak egg production occurring from 18 to 21° C (Humphries 1965; Kornegay and Humphries 1975).

Smith and Rulifson (2015) collected striped bass in the Tar-Pamlico River from early March through mid-April in 2004 and 2005. The NCWRC surveyed striped bass eggs in the Tar-Pamlico River in 1996 and collected 1,366 striped bass eggs with 77.3% being identified as viable during sampling from April through May (Jones and Collart 1997). The NCWRC also collected 188 striped bass eggs from Fishing Creek, a tributary of the Tar-Pamlico River, of which 79% were identified as viable.

Numerous studies employing differing methodology have investigated the presence and viability of striped bass eggs in the Neuse River (Baker 1968; Hawkins 1980; Nelson and Little 1991; Burdick and Hightower 2006; Buckley et al. 2019). Eggs have been collected throughout the Neuse River and its tributaries, generally above Kinston, from the end of March through May. Eggs have been collected at all developmental stages with up to 65% viable eggs (Buckley et al. 2019).

A number of studies have examined the presence of striped bass eggs in the Cape Fear River using variable methodology (Smith 2009; Dial Cordy and Associates 2017; Morgeson and Fisk 2018). Eggs were generally collected from April and May despite sampling occurring in March, though there is generally low abundance of eggs in the river and very few eggs are captured above Lock and Dam 3. Most eggs have been collected below Lock and Dam 1, and collected eggs have been identified as being at multiple developmental stages, although Smith and Hightower (2012) found that the river section between Lock and Dam 2 and Lock and Dam 3 had the highest egg collections and highest predicted proportion of the run.

Research suggests the egg buoyancy of certain strains (e.g., Roanoke River and Chesapeake Bay) are adapted to specific flow conditions. Chesapeake Bay strain eggs are lighter and maintain their position in the water column of calmer tidal waters through neutral buoyancy, whereas Roanoke River strain eggs are heavier and use the more turbulent, high energy system of the Roanoke River to maintain their position in the water column (Bergey et al. 2003).

In 2017, North Carolina State University (CRFL# 2017-F-046) initiated research to provide insight into the current striped bass recruitment status by evaluating genetic and environmental influences on egg development. Preliminary results suggest that the heaviest eggs collected in 2018 and 2019 were from striped bass in the Tar-Pamlico and Neuse rivers (Cara Kowalchyk, NCSU, personal communication). It is interesting to note that the heaviest eggs in the study came from the shallowest river systems; the upper Tar-Pamlico River has an average width of 15 m and an average depth of 0.6 m in the upper reaches and an average width of 49 m and average depth of 4.6 m in the lower reaches (NCWRC 2006).

#### 1.2.4.2 Larvae

The larval development of striped bass is dependent upon water temperature and is usually regarded as having three stages: (1) yolk-sac larvae are 5–8 mm (0.20 to 0.31 inch) in total length and depend on yolk material as an energy source for 7 to 14 days; (2) fin-fold larvae (8–12 mm;

0.31-0.47 inch TL) having fully developed mouth parts and persist about 10 to 13 days; and (3) post fin-fold larvae attain lengths up to 30 mm (1.18 inches) in 20 to 30 days (Hill et al. 1989). Researchers of North Carolina stocks of striped bass (primarily the A-R stock) divide larval development into yolk-sac and post yolk-sac larvae.

Over the past several decades, very few striped bass larvae have been collected in CSMA systems. In 2004 and 2005, Smith and Rulifson (2015) first collected striped bass larvae on the Tar-Pamlico River in early March, and collections continued through mid-May with peak spawning periods detected in April through mid-May. In the Neuse River, only one striped bass larva was collected during each sampling conducted in 1978, 1989, and 2017 (Hawkins 1980; Nelson and Little 1991; Buckley et al. 2019). Larvae (n=32) were collected by Burdick and Hightower (2006) between 8 April–28 May in 2003 and 19 April–12 May in 2004 when water temperatures ranged from 14°C to 28°C in the main stem of the Neuse River and its tributaries. In the Cape Fear River, larval striped bass have generally been captured between April and mid-May with water temperatures ranging 18.6°C–22.5°C. In 2006, larval sampling coincided with the egg collections described in section 1.2.4.1. Larvae were collected at sites downstream of LD-1 (n=1), upstream of LD-1 (n=2), and upstream of LD-2 (n=4). No larval striped bass were captured in 2007, however in 2008 larvae were captured at LD-1 (n=3), LD-3 (n=1), and at the Fayetteville site (n=1). Dial Cordy and Associates Inc. (2017) captured one newly hatched larva below LD-2 on 29 March 2017 when the water temperature was 17.6°C, and two larvae were captured at LD-2 in May. An additional survey for larval fish using quatrefoil light traps was completed May–June 2017 in the Cape Fear, Northeast Cape Fear, and Black Rivers (NCWRC, unpublished data). Although 70 trap nights (1 trap night=1 light trap fished overnight) occurred and over 18 species (155 individuals) were collected, no striped bass larvae or juveniles were collected.

#### 1.2.4.3 Juveniles

Most striped bass enter the juvenile stage at about 30 mm (1.18 inches) TL; the fins are then fully formed, and the external morphology of the young is similar to that of the adults. For the A-R stocks, juveniles are often found in schools and associate with clean sandy bottoms (Hill et al. 1989) and there is evidence of density dependent habitat utilization, with juveniles being collected in the Alligator River and Stumpy Point, Pamlico Sound in late June when large year classes are produced by the A-R stock (NCDMF, unpublished data).

Little is known about juvenile striped bass within the CSMA. Historically, very few juveniles have been captured during NCDMF and NCWRC sampling. Seine and trawl surveys conducted by the NCDMF from 1977 to 1983 collected 37 juveniles in Tar-Pamlico River and 14 juveniles in the Neuse River (Hawkins 1980). The Cape Fear River was only sampled from July–December 1977 and June–September 1978; two striped bass were collected in July 1977.

The NCWRC conducted exploratory juvenile sampling in the Neuse River during 2006 and 2007. No juvenile striped bass were collected in 2006, and five juvenile striped bass were collected in 2007. Evaluation of oxytetracycline (OTC) tagging determined that three of these fish were of hatchery origin and the other two had no OTC mark and could have been wild produced fish (Barwick et al. 2008).

In 2017, exploratory juvenile abundance surveys were developed for the Tar-Pamlico, Neuse, and Cape Fear rivers using trawl and seine nets based on historical sampling locations. No striped bass have been collected in the Tar-Pamlico and Neuse rivers; however, a total of 24 juvenile striped

bass were collected in the Northeast Cape Fear River in 2018 and an additional four were collected in 2019 (Program 100 Juvenile Abundance Survey).

Results from these sampling efforts support the hypothesis of very limited natural reproduction occurring in the CSMA for several decades (Hawkins 1979; Judy and Hawkins 1982; NCDMF 2005; Barwick et al. 2008; NCDMF 2013; Darsee et al. 2019). The existence of limited natural reproduction in the CSMA is supported by results of otolith microchemistry work suggesting 53% of striped bass sampled in the Neuse River in 2010 were not of hatchery origin (Rulifson 2014).

#### **1.2.4.4 Maturation & Fecundity**

There is a strong positive correlation between the length, weight, and age of a female striped bass and the number of eggs produced (Monteleone and Houde 1990; Olsen and Rulifson 1992; Boyd 2011; Knight 2015).

In the Tar-Pamlico and Neuse rivers, 50% of female striped bass are mature at 2.7 years and 98% are mature by age-3 (Knight 2015). Length at 50% maturity (L50) in the CSMA was estimated at 467.8 mm TL (18.4 inches TL) and fish were estimated to be 100% mature at 537.3 mm TL (21.1 inches TL). Female striped bass produce large quantities of eggs which are broadcast into riverine spawning areas and fertilized by mature males, typically age-2 and older. In the Tar-Pamlico and Neuse rivers, fecundity ranged from 223,110 eggs for an age-3 female to 3,273,206 eggs for an age 10 female.

#### **1.2.5 Mortality**

A telemetry tagging study on the Neuse River estimated a discrete annual total mortality of 66.3% for phase II stocked juveniles (202–227 mm TL), a discrete annual total mortality of 54.0% for adults (349–923 mm TL), and a discrete natural mortality of 20.1% for adults (Bradley et al. 2018b). A tagging study showed that striped bass stocked in the Neuse and Tar-Pamlico rivers experienced higher mortality (instantaneous total mortality of 0.48–0.51) than in their natal habitat (instantaneous total mortality of 0.33; Callihan et al. 2014).

Instantaneous total mortality of striped bass in the Neuse River varied considerably from 1997–2011, ranging from 0.36 to 1.08 (Rachels and Ricks 2018). Mortality was generally lowest during the period 1997–2007 and highest during the period 2008–2011. Instantaneous fishing mortality ranged from 0.12–0.84 assuming the instantaneous natural mortality rate given by Bradley (2016) remained constant throughout the time series.

#### **1.2.6 Food & Feeding Habits**

Striped bass are opportunistic feeders; specific food types depend upon the size of the fish, habitat, and the season (Rulifson et al. 1982b). Striped bass undergo an ontogenetic shift in diet with larvae feeding primarily on mobile planktonic invertebrates (Doroshev 1970; Markle and Grant 1970; Bason 1971). As they grow, juvenile striped bass diets include larger aquatic invertebrates and small fish (Shapovalov 1936; Ware 1971). Adult striped bass are piscivorous and primarily feed on fish in the Family Clupeidae, including Atlantic menhaden (*Brevoortia tyrannus*), alewife (*Alosa pseudoharengus*), blueback herring (*Alosa aestivalis*), and gizzard shad (*Dorosoma cepedianum*; Manooch 1973).

Binion-Rock (2018) conducted a multispecies food habits study for 25 finfish species in Pamlico Sound, North Carolina and its tributaries and found that Atlantic menhaden, spot (*Leiostomus*

*xanthurus*), and Atlantic croaker (*Micropogonias undulatus*) were the most commonly consumed fish for larger predators, such as striped bass. Atlantic menhaden contributed the most to bluefish (*Pomatomus saltatrix*), longnose gar (*Lepisosteus osseus*), spotted seatrout (*Cynoscion nebulosus*), and striped bass diets. Striped bass diets also consisted of forage species including anchovies, silversides, mollusks, and polychaetes. Striped bass were also reported as prey items consumed by bluefish, longnose gar, and spotted seatrout.

### **1.3 Habitat**

Striped bass use a variety of habitats with variations in preference due to location, season, and ontogenetic stage. Although primarily estuarine, striped bass use habitats throughout the estuaries and the coastal ocean. Striped bass are found in most habitats identified by the North Carolina Coastal Habitat Protection Plan (CHPP) including water column, wetlands, submerged aquatic vegetation (SAV), soft bottom, hard bottom, and shell bottom (NCDEQ 2016).

The loss of habitat has contributed to the decline in anadromous fish stocks throughout the world (Limburg and Waldman 2009). Numerous documents have been devoted entirely to habitat issues and concerns, including the North Carolina Coastal Habitat Improvement Plan (Street et al. 2005) and ASMFC's "Atlantic Coast Diadromous Fish Habitat: A review of Utilization, Threats, Recommendations for Conservation, and Research Needs" (Greene et al. 2009).

#### **1.3.1 Spawning Habitat**

Spawning grounds are not clearly defined in CSMA systems as access to spawning areas is influenced by river flows as well as impediments to migration. In the Tar-Pamlico River, the main spawning habitat for striped bass occurs from the Rocky Mount Mills Dam downstream approximately 72 km to the vicinity of the town of Tarboro (Kornegay and Humphries 1975; Rock et al. 2018). Acoustic tagging data indicate spawning areas in the Tar-Pamlico River likely occur from the area around Dunbar Road downstream to Tarboro (Rock et al. 2018).

During NCWRC striped bass surveys on the Neuse River, spawning aggregates have been observed from Raleigh, North Carolina to Kinston, North Carolina and acoustic detection data indicate striped bass move upriver as far as Raleigh during the spawning season (Rock et al. 2018). During high flows fish are more likely to spawn near Raleigh, North Carolina (approximately river kilometer (rkm) 350), and when flows are lower fish tend to spawn further downstream around Smithfield, North Carolina (approximately rkm 300; Burdick and Hightower 2006). Striped bass spawning has also been observed further downriver near Goldsboro, North Carolina (rkm 240) and was correlated with higher water velocities and larger substrates (Beasley and Hightower 2000).

In a study conducted by the NCWRC in 2016 and 2017, Neuse River water velocities appeared to be sufficient to keep striped bass eggs suspended until hatching (Buckley et al. 2019). Although water velocities were more variable in 2017, mean velocity for all sites in both years was above the minimum water velocity (30 cm/s) recommended by Albrecht (1964). Additionally, Neuse River velocities were comparable to those observed in the Roanoke River (USGS Oak City, NC 02081022, USGS Williamston, NC 02081054, and Barnhill's Landing from Rulifson and Isely, 1995; Buckley et al., 2019). A study in the Roanoke River indicated that river flow during the pre-spawn and post-spawn periods was the most important factor contributing to survival of larval fish (Hassler 1981); however, comparisons between systems may not be appropriate because of differences in river depth.

In the Cape Fear River, historic anadromous fish spawning areas have been identified from below the mouth of Town Creek, North Carolina to upstream as far as Lillington, North Carolina (Sholar 1977). Three locks and dams were constructed on the main stem of the Cape Fear River between Riegelwood, North Carolina and Tar Heel, North Carolina and the lowermost was completed in 1915 and the uppermost in 1935. These impediments to passage limit the ability of striped bass to reach known historic spawning areas near Smiley Falls at the fall line in Lillington, North Carolina (Nichols and Louder 1970). Several studies in the Cape Fear River have tracked adult striped bass to show evidence of spawning and/or spawning migrations in the main stem river to and above the locks and dams (Ashley and Rachels 2006; Smith 2009; Smith and Hightower 2012). Rock et al. (2018) found that striped bass in the Cape Fear River were generally detected at a core region near downtown Wilmington during all seasons and that many striped bass in the Cape Fear system showed fidelity to and made repeated spring migrations each year up the Northeast and Cape Fear rivers, suggesting spawning migrations or behavioral contingents.

In the Northeast Cape Fear River, Winslow et al. (1983) documented striped bass spawning areas to be located from Croombsbridge Road (rkm 130) to Ness Creek (rkm 47) in the lower Northeast Cape Fear River, and stated that peak spawning occurred in the area downstream of Lanes Ferry (rkm 93); however, Rock et al. (2018) determined that during the spawning season, striped bass migrate to at least near Hallsville, North Carolina (rkm 183), and mature fish were captured between White Stocking, North Carolina (rkm 118) and Chinquapin, North Carolina (rkm 168), thus it is likely that the extent of the upriver spawning habitat in the Northeast Cape Fear River has been underestimated.

### **1.3.2 Nursery & Juvenile Habitat**

Neuse River juvenile striped bass captured in 1979 appeared to show no preference for fresh or brackish water areas but were associated with sandy bottom areas near grass beds (Hawkins 1979).

### **1.3.3 Adult Habitat**

In the Tar-Pamlico river, striped bass are able to migrate as far as Rocky Mount, North Carolina, where Rocky Mount Mills Dam prevents further upstream migration. In the Neuse River, Quaker Neck Dam was removed near Goldsboro, North Carolina in 1998 and Milburnie Dam, in Raleigh, North Carolina, was removed in 2017. Currently, striped bass can access habitats from Falls Dam at Raleigh, North Carolina downstream to the Pamlico Sound. Striped bass are primarily found in these upriver locations during the spawning season from March through May. During the summer and fall, striped bass in the Neuse River concentrate in an area from New Bern downstream to Slocum and Hancock Creeks, and in the Tar-Pamlico River striped bass concentrate in an area from Washington to South Creek (Rock et al. 2018). In the Cape Fear River, adult fish distribution is centered in the upper estuary at the confluence of Cape Fear and Northeast Cape Fear rivers (Wilmington, North Carolina; Stewart and Li 2019).

### **1.3.4 Habitat Issues & Concerns**

There are many contaminants known to adversely affect striped bass at various life stages, particularly at the egg and larvae stages (Setzler et al. 1980; see Richards and Rago 1999 for review), but little is known about current contaminants in the CSMA. Adequate river flows during the spawning season are also needed to keep eggs suspended for proper development (Manooch

and Rulifson 1989). Hassler (1981) indicated that river flow during the pre-spawn and post-spawn periods was the most important factor contributing to survival of fish larvae.

Between 1915 and 1935, three locks and dams were constructed on the Cape Fear River. These structures inhibit access to the historical striped bass spawning grounds. A rock arch rapids fishway was constructed at Lock and Dam 1 in 2012 to provide improved volitional passage for anadromous fish; however, Raabe et al. (2019) determined the structure was not effective for striped bass. Consequently, striped bass reproduction is limited because migration to traditional spawning grounds on the Cape Fear River is restricted.

## **1.4 Description of Fisheries**

### **1.4.1 Commercial Fishery**

Commercial landings in the CSMA have been constrained by an annual Total Allowable Landings (TAL) of 25,000 pounds since 1994. Most commercial landings come from the Pamlico and Pungo rivers and the Neuse and Bay rivers and the remainder come from Pamlico Sound. Since 2004, there has only been a spring harvest season, recently opening March 1 each year and closing when the TAL is reached. In 2008 due to continued concerns over low abundance levels, a no-harvest provision was implemented in the Cape Fear River. Due to the no possession measure for the remainder of the CSMA approved in Supplement A to Amendment 1 to the N.C. Estuarine Striped Bass FMP (NCDMF 2019a), the commercial striped bass fishery was closed in 2019 while Amendment 2 to the N.C. Striped Bass FMP is developed (refer to Figure 2.1).

### **1.4.2 Recreational Fishery**

Coastal striped bass populations have continuously provided a popular and economically important recreational fishery in North Carolina. Despite past surveys covering a considerable area, recreational fisheries data were lacking for the CSMA when the stock was listed as overfished in 2003. A comprehensive creel survey was initiated in January 2004 to identify and estimate recreational striped bass effort and catch in the CSMA, particularly the Tar-Pamlico and Neuse river systems. Due to the recreational no possession measure implemented by the NCMFC and the NCWRC in Supplement A to Amendment 1 to the N.C. Estuarine Striped Bass FMP (NCDMF 2019a), the recreational striped bass fishery was closed in 2019 while Amendment 2 to the N.C. Striped Bass FMP is developed (refer to Figure 2.4).

## **1.5 Fisheries Management**

### **1.5.1 Management Authority**

Fisheries management includes all activities associated with maintenance, improvement, and utilization of the fisheries resources of the coastal area including research, development, regulation, enhancement, and enforcement. North Carolina's existing fisheries management system is powerful and flexible and rulemaking (and proclamation) authority is vested in the NCMFC and the NCWRC within their respective jurisdictions.

The North Carolina Department of Environmental Quality (NCDEQ) is the parent agency of the NCMFC and the NCDMF. The NCMFC is responsible for managing, protecting, preserving, and enhancing the marine and estuarine resources under its jurisdiction, which includes all state coastal fishing waters extending to three miles offshore. In support of these responsibilities, the NCDMF



conducts management, enforcement, research, monitoring statistics, and licensing programs to provide information on which to base these decisions. The NCDMF presents information to the NCMFC and NCDEQ in the form of fisheries management and coastal habitat protections plans and proposed rules. The NCDMF also administers and enforces the NCMFC's adopted rules.

The NCWRC is a state government agency authorized by the General Assembly to conserve and sustain the state's fish and wildlife resources through research, scientific management, wise use, and public input. The NCWRC is the regulatory agency responsible for the creation and enforcement of hunting, trapping, and boating laws statewide and fishing laws within its jurisdictional boundaries including all designated inland fishing waters. The NCWRC and NCDMF share authority for regulating recreational fishing activity in joint fishing waters.

### **1.5.2 Management Unit Definition**

There are three geographic management units defined in the Estuarine Striped Bass FMP and the fisheries throughout the coastal systems of North Carolina (NCDMF 2004). The management unit for this evaluation is the CSMA and is defined as:

The CSMA includes all internal coastal, joint and contiguous inland waters of North Carolina south of the ASMA to the South Carolina state line. There are spawning stocks in each of the major river systems within the CSMA; the Tar-Pamlico, the Neuse, and the Cape Fear. These stocks are collectively referred to as the CSMA stocks. Spawning grounds are not clearly defined in these systems as access to spawning areas is influenced by river flows as well as impediments to migration. Management of striped bass within the CSMA is the sole responsibility of the NCMFC and the NCWRC and is not subject to compliance with the ASMFC Interstate FMP for Atlantic Striped Bass (Figure 1.1).

### **1.5.3 Regulatory History**

Estuarine striped bass in North Carolina are managed jointly by the NCMFC and the NCWRC under Amendment 1 (NCDMF 2013), Revision 1 to Amendment 1 (NCDMF 2014), and Supplement A to Amendment 1 (NCDMF 2019a) to the N.C. Estuarine Striped Bass FMP (NCDMF 2004). Amendment 1, adopted in 2013, lays out separate management strategies for the A-R stock in the ASMA and the RRMA and the CSMA stocks in the Tar-Pamlico, Neuse, and Cape Fear rivers. Management measures in Amendment 1 consist of daily possession limits, open and closed harvest seasons, seasonal gill-net attendance and other gill-net requirements, minimum size limits, and slot limits to maintain sustainable harvest and reduce regulatory discard mortality in all sectors. Amendment 1 also maintained the stocking measures in the major CSMA river systems and the harvest moratorium on striped bass in the Cape Fear River and its tributaries, including Snow's Cut (NCDMF 2013).

The following regulations were initially contained in the jointly adopted Amendment 1 to the N.C. Estuarine Striped Bass FMP. Both commercial and recreational fisheries are subject to an 18-inch TL minimum size limit for striped bass within the CSMA. As an additional protective measure in joint and inland CSMA waters, it is unlawful for recreational fishermen to possess striped bass between 22 and 27 inches TL. The recreational harvest season for striped bass within the CSMA is October 1 through April 30. Recreational fishermen are constrained to a two fish per person per day possession limit.

The striped bass commercial fishery in the CSMA is a directed fishery, except in Pamlico Sound where bycatch restrictions are in place and primarily uses anchored large mesh ( $\geq 5$  inches stretched mesh (ISM) gill nets. There is a commercial daily possession limit of 10 fish per person per day with a maximum of two limits per commercial operation issued by proclamation. Daily reporting of the number and pounds of striped bass landed from all licensed striped bass dealers helps ensure the 25,000 pound total allowable landings (TAL) is not exceeded. The commercial harvest season opens by proclamation and may occur between January 1 and April 30 and is closed by proclamation once the annual 25,000 pound TAL is reached or on April 30, whichever occurs first. After closure of the commercial harvest season and continuing through December 31, commercial fishermen are required to use three-foot tie downs in gill nets with a stretch mesh length  $\geq 5$  inches in internal coastal fishing waters west of the 76 28.0000' W longitude line. They must also maintain a minimum distance from shore (DFS) of 50 yards for these nets upstream of the existing DFS line.

In recreational and commercial fisheries, it has been unlawful to possess striped bass taken from the internal coastal and joint waters of the Cape Fear River and its tributaries since 2008 per MFC Rules 15A NCAC 03M .0202 and 03Q .0107, and in the inland fishing waters of the Cape Fear River and its tributaries downstream of Buckhorn Dam per NCWRC rules 15A NCAC 10C .0314 (h).

The following management change was implemented solely under the purview of the NCWRC and was not developed through the NCDMF FMP process. The NCWRC has jurisdiction in the inland waters of the CSMA, and on February 16, 2016, the NCWRC voted to modify the exception to the general statewide size regulation for striped bass in inland waters of the Tar-Pamlico, Pungo, and Neuse rivers and their tributaries by increasing the minimum size limit from 18 inches to 26 inches TL. The no-possession prohibition on fish between 22 and 27 inches TL was removed. The daily creel limit (two fish per person per day) and harvest season (October 1–April 30) were not changed. The new rule was scheduled to go into effect August 1, 2017, but ten letters of objection requesting legislative review of the rule were received in March 2017. No action was taken during the mandatory legislative review period, and the rule 15A NCAC 10C .0314 became effective on June 1, 2018.

Supplement A to Amendment 1 to the N.C. Estuarine Striped Bass FMP was adopted by the NCMFC at their February 2019 business meeting and by the NCWRC in March 2019 (NCDMF 2019a). Supplement actions in the FMP implemented March 29, 2019 consisted of a recreational no possession measure for striped bass (including hybrids) in coastal and inland fishing waters of the CSMA (NCDMF Proclamation FF-6-2019). The NCWRC hook-and-line closure proclamation had the effect of suspending rules 15A NCAC 10C .0107 (l) and 10C .0314 (g). A no-possession requirement has been in place for the Cape Fear River by rule since 2008.

In March 2019, the NCMFC held an emergency meeting that directed the NCDMF to issue a proclamation regarding gill nets, beyond what was contained in Supplement A to Amendment 1 to the N.C. Estuarine Striped Bass FMP. Proclamation (M-6-2019) prohibits the use of all gill nets upstream of the ferry lines from the Bayview Ferry to Aurora Ferry on the Pamlico River and the Minnesott Beach Ferry to Cherry Branch Ferry on the Neuse River. It also maintains tie-down (vertical net height restrictions) and distance from shore restrictions for gill nets with a stretched mesh length 5 inches and greater in the western Pamlico Sound and rivers.

An emergency meeting called under North Carolina General Statute section 113-221.1(d), authorizes the NCMFC to review the desirability of directing the fisheries director to issue a proclamation. Once the NCMFC votes under this provision to direct issuance of a proclamation, the NCDMF fisheries director has no discretion to choose another management option and is bound by law to follow the NCMFC decision. In these cases, under existing law, the decision of the NCMFC to direct the director to issue a proclamation is final and can only be overruled by the courts.

#### **1.5.4 Current Regulations**

Commercial and recreational harvest of striped bass in the CSMA is prohibited. Supplement A to the N.C. Estuarine Striped Bass FMP was adopted by the NCMFC at their February 2019 business meeting (NCDMF Proclamation FF-6-2019) and by the NCWRC in March 2019. The NCWRC hook-and-line closure proclamation had the effect of suspending rules 15A NCAC 10C .0107 (l) and 10C .0314 (g). NCDMF proclamation (M-6-2019) prohibits the use of all gill nets upstream of the ferry lines from the Bayview Ferry to Aurora Ferry on the Pamlico River and the Minnesott Beach Ferry to Cherry Branch Ferry on the Neuse River. It also maintains tie-down (vertical net height restrictions) and distance from shore restrictions for gill nets with a stretched mesh length 5 inches and greater in the western Pamlico Sound and rivers.

As a response to low numbers of documented spawning adults and limited evidence of juvenile recruitment, the NCDMF and NCWRC implemented, by separate rule making, a moratorium on both the commercial and recreational harvest of striped bass in the Cape Fear River in 2008, which is still in effect.

#### **1.5.5 Management Performance**

Stocking appears to have maintained striped bass populations in the Tar-Pamlico and Neuse rivers during recent history, in the absence of stocking, population declines likely would have occurred given the absence of natural recruitment and evidence that populations remain almost entirely composed of hatchery fish. The slot limit imposed on the joint and inland waters portions of the Tar-Pamlico and Neuse rivers does not seem to have protected spawning females to older age classes as intended. In an effort to reduce discards in the commercial fishery, tie-downs and distance from shore measures adopted in the 2004 Estuarine Striped Bass FMP (NCDMF 2004) were implemented in 2008. Rock et al. (2016) investigated the effectiveness of these management measures by collecting effort, catch, and bycatch data for striped bass in the commercial estuarine large mesh gill-net fishery. Due to the persistence of striped bass in nearshore waters and the comparatively low number of discarded striped bass observed in commercial gill nets, it appears as though the distance from shore and tie-down requirements enacted in 2008 have been successful in reducing the number of striped bass discards in the commercial gill-net fishery in the Tar-Pamlico and Neuse rivers. Overall, this study indicated approximately an 82% reduction in striped bass discards from previous levels estimated in Amendment 1 to the Estuarine Striped Bass FMP (NCDMF 2014); however, Rachels and Ricks (2018) observed that gill-net effort (number of nets set annually) had greater impact on Neuse River striped bass mortality rates than commercial harvest and theorized that discard mortality continues to significantly impact the population. The work of Rachels and Ricks (2018) was expanded as part of this assessment to include removals from all sectors (recreational and commercial) that could influence discrete annual mortality. Results from additional analysis showed, along with the relative annual variation in commercial

effort and in commercial harvest, the relative annual variation in recreational effort and in recreational discards were also significant factors contributing to the relative annual variation in total mortality of striped bass in the Neuse River (see section 5).

## **1.6 Assessment History**

No formal peer-reviewed stock assessments have been conducted for the CSMA striped bass.

### **1.6.1 Review of Previous Methods & Results**

No peer-reviewed stock assessments have been conducted for the CSMA striped bass; however, an index-based method of catch curve analysis was used to assess the status of striped bass populations in the CSMA (Appendix 14.7 in NCDMF 2013). The large confidence intervals and lack of precision in the catch curve  $Z$  estimates (total instantaneous mortality rate) made them unsuitable for making a stock status determination (NCDMF 2013).

### **1.6.2 Progress on Research Recommendations**

No peer reviewed stock assessment has been conducted for CSMA striped bass stocks. However, many of the research recommendations from the FMP focused on collection of data and life history information needed for completion of a stock assessment.

- Increase surveys of stocked systems to determine percent contribution of hatchery stocked fish (ongoing through NCWRC and NCDMF genetics survey)
- Conduct egg abundance and egg viability studies

The NCWRC and the NCDMF continue to collect genetic data throughout the range of striped bass to evaluate the percent contribution of hatchery stocked fish. In 2016 and 2017, the NCWRC sampled anadromous ichthyoplankton to investigate striped bass egg and larval abundance and egg viability (Buckley and Ricks 2018). In 2017, North Carolina State University began research designed to investigate striped bass egg yolk composition, egg buoyancy, and recruitment.

- Acquire life history information: maturity, fecundity, size and weight at age, egg and larval survival

Knight (2015) conducted research on striped bass maturation and fecundity in the Neuse and Tar-Pamlico rivers and additional work is ongoing through the NCDMF ageing program. In 2017, to adequately capture all life stages of striped bass, Program 100 was expanded into the CSMA to evaluate juvenile striped bass recruitment.

- Improve tagging program, conduct a mark-recapture study utilizing conventional tags and telemetry approaches to estimate fishing mortality and abundance

Conventional tagging and deployment of acoustic tagged striped bass has continued in the CSMA to improve estimates of fishing mortality and abundance. Rock et al. (2018) assessed critical habitat, movement patterns, and spawning grounds of anadromous fishes in the Tar-Pamlico, Neuse, and Cape Fear rivers using telemetry tagging techniques.

- Develop better estimates of life-history parameters, especially growth and natural mortality

In an effort to improve discard estimates in the commercial gill-net fishery, Rock et al. (2016) evaluated discard estimates through the NCDMF creel survey and an expanded NCDMF observer program. Bradley et al. (2018a and 2018b) conducted research in the Neuse River to estimate mortality rates of juvenile and adult striped bass, determine distribution and migration patterns of adults, and built an age-structured population model to explore the effects of observed mortality rates on the adult population.

## **2 DATA**

### **2.1 Fisheries-Dependent**

#### **2.1.1 Commercial Landings**

Prior to 1978, North Carolina's commercial landings data were collected by the National Marine Fisheries Service (NMFS). Between 1978 and 1993, landings information was gathered through the NMFS/North Carolina Cooperative Statistics program. Reporting was voluntary during this period and North Carolina and NMFS port agents sampled the state's major dealers (Lupton and Phalen 1996).

On January 1, 1994, the NCDMF initiated a Trip Ticket Program (TTP) to obtain more complete and accurate trip-level commercial landings statistics (Lupton and Phalen 1996). Trip ticket forms are used by state-licensed fish dealers to document all transfers of fish sold from coastal waters from the fishermen to the dealer. The data reported on these forms include transaction date, area fished, gear used, and landed species as well as fishermen and dealer information.

The majority of trips reported to the NCDMF TTP only record one gear per trip; however, as many as three gears can be reported on a trip ticket and are entered by the program's data clerks in no particular order. When multiple gears are listed on a trip ticket, the first gear may not be the gear used to catch a specific species if multiple species were listed on the same ticket but caught with different gears. In 2004, electronic reporting of trip tickets became available to commercial dealers and made it possible to associate a specific gear for each species reported. This increased the accuracy of reporting by documenting the correct relationship between gear and species.

##### **2.1.1.1 Sampling Intensity**

North Carolina dealers are required to record the transaction at the time of the transactions and report trip-level data to the NCDMF (see NCDMF 2019).

##### **2.1.1.2 Biological Sampling**

Historically, biological sampling occurred during the spring and fall fishery; however, since 2004 there has only been a spring harvest season. This is a directed fishery (except Pamlico Sound) for striped bass primarily using anchored gill nets. Commercial fish houses are sampled throughout the CSMA, during each open harvest season. Fish are measured to the nearest mm for fork length (FL) and TL and weighed to the nearest 0.01 kg. Striped bass scales and otoliths have been collected sporadically by the NCDMF since 1975, although since 2003 both scales and otoliths have been collected routinely. Scales are removed from the left side of the fish, above the lateral line and between the posterior of the first dorsal fin and the insertion of the second dorsal fin. Scales are cleaned and pressed on acetate sheets using a Carver heated hydraulic press. NCDMF staff read scales using a microfiche reader set on 24x or 33x magnification. Otoliths are collected

from the left and right sides, but only one side (left) is typically sectioned and mounted for ageing. To prepare otoliths for ageing, thin sections of whole otoliths were cut, mounted to a slide, ground down, and covered with a top coat. Starting in 2016, although limited in number, PBT samples were also collected by taking a partial pelvic fin clip and preserving in 95% ethyl alcohol.

#### **2.1.1.3 Potential Biases & Uncertainties**

All fish that are caught are not required to be landed and sold so some fish may be taken for personal consumption and not reported in the landings under this program authority. Hadley (2015) found that 28% of commercial license holders maintained a license for personal consumption or donation of harvest. Another potential bias relates to the reporting of multiple gears on a single trip ticket because the order in which gears are reported is not indicative of the primary method of capture.

#### **2.1.1.4 Development of Estimates**

Commercial landings were summarized by year using the NCDMF TTP data. Length data collected from the commercial fish house sampling program were used to compute annual length-frequency distributions.

#### **2.1.1.5 Estimates of Commercial Landings Statistics**

Commercial landings in the CSMA have been constrained by an annual TAL of 25,000 pounds since 1994. Over the past ten years, landings have closely followed the annual TAL, except for 2008 when less than half of the TAL was landed. Since 2004, striped bass commercial landings in the CSMA have averaged 24,179 pounds and ranged from a low of 10,115 pounds in 2008 to a high of 32,479 pounds in 2004 (Figure 2.1). Most commercial landings come from the Pamlico and Pungo rivers and the Neuse and Bay rivers and the remainder come from the Pamlico Sound (Figure 2.2).

Length data from the commercial harvest shows that on average striped bass in the Neuse and Bay rivers are slightly larger than fish harvested in the Pamlico and Pungo rivers (Table 2.1). Additionally, maximum lengths are generally larger in the Neuse and Bay rivers compared to the Pamlico and Pungo rivers. CSMA commercial length frequencies show that striped bass are routinely harvested up to 30 inches total length and that few fish under the 18 inch total length minimum size limit are harvested (Figures 2.3, 2.4).

### **2.1.2 Commercial Gill-Net Discards**

#### **2.1.2.1 Survey Design & Methods**

NCDMF's Program 466 (Onboard Observer Monitoring) was designed to monitor fisheries for protected species interactions in the gill-net fishery by providing onboard observations. Additionally, this program monitors finfish bycatch and characterizes effort in the fishery. The onboard observer program requires the observer to ride onboard the commercial fishermen's vessel and record detailed gill-net catch, bycatch, and discard information for all species encountered. Observers contact licensed commercial gill-net fishermen holding an Estuarine Gill-Net Permit (EGNP) throughout the state to coordinate observed fishing trips. Observers may also observe fishing trips from NCDMF vessels under Program 467 (Alternative Platform Observer Program), but these data were not used in this analysis due to the lack of biological data collected through the program.

### 2.1.2.2 Sampling Intensity

Commercial fishing trips targeting striped bass are observed during the open season (March–April); however, most observed trips occur outside of that time period when striped bass are discarded as bycatch in other gill-net fisheries.

### 2.1.2.3 Biological Sampling

Data recorded includes species, weight, length, and fate (landed, live discard, or dead discard).

### 2.1.2.4 Potential Biases & Uncertainties

Program 466 began sampling statewide in May 2010. To provide optimal coverage throughout the state, management units were created to maintain proper coverage of the fisheries. Management units were delineated based on four primary factors: (1) similarity of fisheries and management, (2) extent of known protected species interactions in commercial gill-net fisheries, (3) unit size, and (4) the ability of the NCDMF to monitor fishing effort. Total effort for each management unit can vary annually based on fishery closures due to protected species interactions or other regulatory actions. Therefore, the number of trips and effort sampled each year by management unit varies both spatially and temporally.

Program 466 data do not span the entire time series for this analysis (no data are available for 1991–2000) and statewide sampling began in May 2010 decreasing the variability of observed trips with better spatial and temporal sampling beginning in 2012.

Striped bass discard data were not available in sufficient quantities to estimate discards or post-release mortality from other fisheries; however, other gears, like pound nets, are known to have discards of striped bass.

It is also important to note that this survey was designed to target trips that occur in times and areas where protected species interactions are highest; the program does not target striped bass trips. For this reason, a high number of zero-catch trips relative to striped bass occur in the data.

### 2.1.2.5 Development of Estimates

A generalized linear model (GLM) framework was used to estimate striped bass discards in the North Carolina commercial gill-net fishery based on data collected from the mandatory observer program (initiated 2012) during 2013 through 2018. The presence or absence of striped bass from on-board observer trips in the Tar-Pamlico and Neuse rivers (Figure 2.5) was used to more accurately estimate striped bass discards from the commercial gill-net fisheries (Figure 2.6). Only those variables available in all data sources were considered as potential covariates in the model. Available variables were year, season, mesh category (small: <5 inches and large: ≥5 inches), and area (Figure 2.6), which were all treated as categorical variables in the model. Year is based on the calendar year. Season is based on the calendar year such that January through February, and December equates to winter, March through May equates to spring, June through August equates to summer, and September through November equates to fall. Discards were assigned to one of four areas: (1) Albemarle-Roanoke, (2) Neuse, (3) Tar-Pamlico, or (4) Cape Fear. Though estimates for the Albemarle-Roanoke were produced, they are not presented in this report. Due to the overall low gill-net activity and observed striped bass in the Cape Fear River during the 2013 to 2018-time period, commercial discards could not be estimated for this area.

All available covariates were included in the initial model and assessed for significance using likelihood ratio tests (Zuur 2012). Non-significant covariates were removed using backwards selection to find the best-fitting predictive model. An offset term was included in the model to account for differences in fishing effort among observations (Zuur et al. 2009, 2012). Effort was measured as soak time (days) multiplied by net length (yards). Using effort as an offset term in the model assumes the number of striped bass discards is proportional to fishing effort (A. Zuur, Highland Statistics Ltd., personal communication).

Live and dead discards were modeled separately. Examination of the data indicated both the live and dead discard data were zero inflated. There are two types of models commonly used for count data that contain excess zeros. Those models are zero-altered (two-part or hurdle models) and zero-inflated (mixture) models (see Minami et al. 2007 and Zuur et al. 2009 for detailed information regarding the differences of these models). Minami et al. (2007) suggests that zero-inflated models may be more appropriate for catches of rarely encountered species; therefore, zero-inflated models were initially considered though were unable to converge. For this reason, zero-altered models were pursued.

The best-fitting models for live discards and for dead discards were applied to available effort data from the NCDMF TTP to estimate the total number of live discards and dead discards for the North Carolina commercial gill-net fishery. Because not all live discards survive, an estimate of post-release mortality was applied to the predicted number of live discards to estimate the number of live discards that did not survive. Live discards are multiplied by an estimated discard mortality rate for gill nets of 43% (ASMFC 2007). This estimate was added to the number of dead discards to produce an estimate of the total number of dead discards for the North Carolina commercial gill-net fishery.

#### **2.1.2.6 Estimates of Commercial Gill-Net Discard Statistics**

The best-fitting GLM for the commercial gill-net live discards assumed a zero-altered Poisson distribution (dispersion=3.3). The significant covariates for the count part of the model were year and mesh category and the significant covariates for the binomial part of the model were year, season, mesh category, and management area. The best-fitting GLM for the dead discards assumed a zero-altered Poisson distribution as well (dispersion=2.5). The significant covariates for the count part of the model were year, and season, and the significant covariate for the binomial part of the model was season.

In both the Neuse and Tar-Pamlico rivers, dead discards were higher in large mesh ( $\geq 5$  inches) gill nets than in small mesh ( $< 5$  inches) gill nets, though in some years estimates between the two years were similar. Estimates of total dead discards in the Neuse River ranged from a low of 140 striped bass in 2017 to a high of 342 in 2013 (Table 2.2). Estimates of total dead discards in the Tar-Pamlico River were higher than in the Neuse River and ranged from a low of 306 striped bass in 2017 to a high of 709 in 2013 (Table 2.3). Relatively low estimates of dead discards are potentially an indicator that the distance from shore and tie-down requirements enacted in 2008 have been successful in reducing the number of striped bass discards in the commercial gill net fishery in the Tar-Pamlico and Neuse rivers (Rock et al. 2016).



### **2.1.3 Recreational Fishery Monitoring**

A comprehensive angler creel survey was initiated in January 2004 to identify and estimate recreational striped bass effort and catch in the CSMA.

#### **2.1.3.1 Survey Design & Methods**

Survey points in the Neuse River included 45 boat ramps and fishing access points from Milburnie Park in East Raleigh to Lee's Landing on Broad Creek. The river was divided into three segments and all access points in Goldsboro and above classified as the upper zone, sites on Contentnea Creek and downstream from Goldsboro to Core Creek were considered the middle zone, and those downstream from Core Creek, the lower zone. Prior to 2012, the Neuse River was comprised of only two zones and all sites above Contentnea Creek considered the upper.

Access points surveyed on the Tar-Pamlico River include 19 boat ramps and access sites from Battle Park in Rocky Mount to the Quarterdeck Marina in Bath, North Carolina. This system was divided into upper and lower zones and sites upstream of Greenville, North Carolina are considered the upper zone. The Pungo River was surveyed at the Leechville ramp (NC-264 bridge), the Belhaven NCWRC ramp, Wrights Creek (NCWRC) ramp, and Cee Bee Marina on Pungo Creek.

#### **2.1.3.2 Sampling Intensity**

Recreational fishing statistics from the CSMA are calculated through a non-uniform stratified access-point creel survey (Pollock et al. 1994) on the Neuse, Pamlico, and Pungo rivers from January–December. Site probabilities were set in proportion to the likely use of the site according to time of day, day of the week, and season. Probabilities for this survey were assigned based on observed effort from past years and direct observation by creel clerks. Morning and afternoon periods were assigned unequal probabilities of conducting interviews and each period represents half a fishing day. A fishing day was defined as the period from one hour after sunrise until one hour after sunset. This is slightly different than in years prior to 2012 when the fishing day was defined as beginning 1.5 hours after sunrise. Monthly sampling periods for each river and zone were stratified accordingly, and all weekend and holiday dates along with two randomly selected weekdays were chosen from each week for sampling.

Tar-Pamlico River anglers in the upper zone were interviewed throughout the spring months (January–May), while anglers in the lower zone were interviewed year round based on the evidence of a year-round fishery and no seasonal closures. Two creel clerks were assigned to this river, with one surveying the upper zone January through May and one clerk surveying the lower zone from January through December. The three zones within the Neuse River were covered with one creel clerk per zone. The lower zone was surveyed from January to December while middle zone surveys were conducted January–May and the upper zone surveys from February–May. The Pungo River was surveyed throughout the year with one creel clerk.

Returning fishing parties are interviewed by a creel clerk at the selected access point to obtain information regarding party size, effort, total number of fish harvested and/or released, primary fishing method, and location.

Creel clerks also obtained socioeconomic information from the angler, including age, state and county of residence, sex, ethnic background, marital status, number of individuals within household, and trip information and expenditures.

### 2.1.3.3 Biological Sampling

Harvested fish are identified, counted, measured to the nearest mm fork length (converted to centerline length and total length for appropriate species), and weighed to the nearest 0.1 kg, while information on discarded fish was obtained from the angler to acquire the number and status of discarded individuals. Scale collections were taken from available fish to determine age of catch. Since 2015, additional biological sampling has included the collection of striped bass fin clips for genetic analysis.

### 2.1.3.4 Potential Biases & Uncertainties

The current dockside sampling methodology only intercepts those individuals accessing inland, joint, and coastal waters via public boating access sites thereby excluding those individuals using private access such as residences, marinas, and community boat ramps. Given the substantial human footprint within the CSMA, it is certain that estimates of effort and catch currently being produced by the NCDMF are under-representative of the actual fishing pressure and associated catch occurring in these systems.

### 2.1.3.5 Development of Estimates

#### Effort and Catch Estimations

Only striped bass effort and catch data were used to produce estimates. Results were stratified by river, access point, and time of day. Catch was defined as the sum of harvested fish and discarded fish. Discarded fish equaled the sum of fish caught in excess of creel limits (over-creel), legal-sized fish caught and released, and sub-legal fish returned to the water. Daily effort and catch for each river were calculated by expanding observed numbers by the sample unit probability (time of day probability multiplied by access area probability). Total catch estimates for the CSMA and catch estimates for each zone and type of day were calculated based on the Horvitz-Thompson estimator for non-uniform probability sampling (NCDMF 2019b; Pollock et al. 1994). Total effort, in number of trips, over the CSMA and each individual zone and type of day were estimated in the same fashion, as were other extrapolated data. Targeted trips refer to trips where the angler explicitly identified their target species during the sampling interview. If multiple species were targeted, then a primary target species was designated (1st target; see Appendix IV.1 in NCDMF 2017). Approximate standard errors (SE) of the catch and effort estimates within zone and type of day were calculated based on the variance of the observations, the number of days sampled, and the number of days of that type available for sampling (Pollock et al. 1994). Percent standard errors (PSE) for the year are presented by river system and zone. Monthly PSEs within river system and zone are available upon request. Estimated catch-per-unit-effort (CPUE) values were obtained by dividing estimated catch by estimated striped bass trips as well as angler hours (angler-h) in order to identify trends in fishing pressure and angler success. Size structure of striped bass in harvests was described for each zone using length-frequency distributions of observed samples. Fishing party characteristics and methods used during striped bass trips reported by anglers were documented by river and day type. Beginning in 2012, the NCWRC Portal Access to Wildlife Systems (PAWS) was used to house these data and estimate effort and catch. NCDMF and NCWRC staff have been verifying calculations to ensure consistency with the previous work. Please note that estimates of catch and effort are expanded averages presented as whole numbers. Any inconsistency in the total catch and/or effort due to adding across rows or columns presented in this chapter is due to rounding.

### **2.1.3.6 Estimates of Recreational Fishery Statistics**

In 2018, recreational landings were 10,844 pounds; however, recreational landings have fluctuated since 2004 and have ranged from lows in 2008 and 2009 to a high of 26,973 pounds most recently in 2017 (Table 2.4). In recent years, both the number of trips and the hours spent targeting striped bass within the CSMA have increased, although recreational harvest dropped sharply by more than half of the 2016 and 2017 values in 2018 (Table 2.4; Figure 2.7). Harvest on the Pungo River has remained consistent at a relatively low level compared to fluctuations in the Tar-Pamlico and Neuse rivers. Since 2011, harvest in the Tar-Pamlico and Neuse rivers has been similar, ranging from 4,000 pounds to 9,000 pounds; however, in 2016 and 2017 there was a sharp increase in recreational harvest (25,260 and 26,973 pounds, respectively; Figure 2.8).

Legal-sized striped bass discards have increased over the past six years, more than doubling in 2017 but returning to more normal levels in 2018 (12,232 legal sized discarded fish; Table 2.4). Fish released that were within the slot limit, have fluctuated since 2004 and have ranged from a low in 2004, 2006, and 2007 of zero fish to a high of 6,779 fish in 2016. In 2018, there were approximately 1,890 discarded striped bass that were within the slot limit. In 2017, mainly due to the large number of undersized striped bass available, there was more than a fivefold increase in the number of discards occurring in the fishery since 2015; however, in 2018 there was a sizeable decline back to more normal levels (34,128 under sized discarded fish; Table 2.4; Figure 2.9). Within the CSMA, there is a significant catch-and-release fishery during the summer in the middle reaches of the Tar-Pamlico and Neuse rivers. Releases during the last ten years have averaged 43,255 fish per year (Table 2.4). CSMA recreational length frequencies show that striped bass are routinely harvested up to 25 inches TL and that few fish under the 18 inch total length minimum size limit are harvested (Figures 2.10, 2.11).

## **2.2 Fisheries-Independent**

### **2.2.1 Juvenile Abundance Survey (Program 100)**

#### **2.2.1.1 Survey Design & Methods**

In 2017, exploratory juvenile abundance sampling was initiated in the Tar-Pamlico, Neuse, and Cape Fear rivers using trawl and seine nets replicating methods used in the ASMA. The fixed station survey uses an 18-foot semi-balloon trawl with a body mesh size of 0.75-inch and a 0.25-inch mesh tail bag with ten-minute tow times. Beach seines are 60-ft long by 6-ft tall, with a 6 ft by 6 ft by 6 ft bag constructed of 0.25-inch stretch mesh (ISM) in the body and 0.125 ISM in the bag. Seine nets are stretched parallel to shore approximately 30 feet from shore and pulled directly to the beach. NCDMF staff continue to develop and refine these abundance surveys in order to standardize sampling methods and locations. In the Tar-Pamlico River, sampling occurs from Washington, North Carolina to South Creek and in the Neuse River sampling occurs from New Bern to Slocum Creek (Figure 2.12). In the Cape Fear River, sampling occurs in the mainstem as well as in the Northeast Cape Fear, and between Lock and Dams 1 and 2 (Figure 2.13).

#### **2.2.1.2 Sampling Intensity**

Sampling in the Tar-Pamlico and Neuse rivers occurs during early June and continues through late October. Beach seines are conducted weekly at six locations in the Tar-Pamlico River and at six locations in the Neuse River. Sampling using seines starts the first week of June and continues weekly until the second week of July, for a total of six rounds of sampling and 72 total combined

samples. Bi-weekly trawl samples are conducted at six locations in the Tar-Pamlico and at six locations in the Neuse River with ten-minute tow times. Sampling occurs from the third week of July through late October, for a total of eight rounds of sampling and 96 total combined samples.

Sampling in the Cape Fear River occurs during early June and continues through late October. Beach seines are conducted weekly from June through mid-July. Due to the hydrological features of the Cape Fear and tidal nature of the system, distance from shore is at maximum 30 feet. Several sites are unable to be sampled occasionally due to environmental conditions (i.e., high flow/flood conditions, low tide line at the drop-off to 60 feet).

Bi-weekly trawl samples are conducted at fixed locations in the Cape Fear River with ten-minute tow times starting the third week of July and ending in late October.

#### **2.2.1.3 Biological Sampling**

All striped bass captured are counted and a subsample (maximum of 30) is measured (mm; FL and TL). Genetic samples (fin clips) are collected from all juvenile striped bass captured in the CSMA surveys.

Surface and bottom water temperature (°C), dissolved oxygen (mg/L), secchi depth (cm), and salinity (ppt) are recorded at each station. Submerged aquatic vegetation (SAV) is identified to species and/or genus level.

#### **2.2.1.4 Potential Biases & Uncertainties**

The Juvenile Abundance Survey employs a fixed-station survey design that is currently exploratory in nature. A fixed-station survey can run the risk of bias if the sites selected do not adequately represent the sampling frame. Additionally, even if the sites adequately cover the sampling frame, the increased variation that would come about from sampling randomly is not accounted for and is therefore at risk of being neglected.

Indices derived from fixed-station surveys such as P100 may not accurately reflect changes in population abundance (Warren 1994, 1995); however, Blanchard et al. (2008) found that fixed-stratified survey design provided the greatest power to identify abundance trends in depleted stocks compared to random or random stratified. The accuracy of the estimates is tied to the degree of spatial persistence in catch data of the species (Lee and Rock 2018). The persistence of the P100 data in the CSMA has not been evaluated.

#### **2.2.1.5 Development of Estimates**

Because of the exploratory nature of the survey and the short time series and low catches, estimates of juvenile striped bass abundance cannot be developed at this time.

#### **2.2.1.6 Estimates of Survey Statistics**

In three years of sampling, no juvenile striped bass have been captured in the Tar-Pamlico or Neuse rivers. In the Cape Fear River (Northeast Cape Fear River), a total of 24 young-of-year (YOY) striped bass were captured in 2018 and four were captured in 2019. The YOY striped bass surveys in the CSMA were implemented to have sampling programs in place to monitor natural recruitment in these systems and measure the success of management strategies developed in Amendment 2 to the Striped Bass Fishery Management Plan. If natural recruitment does occur in the CSMA river systems, data from this survey will be valuable for estimating year-class strength and as an index of juvenile abundance in stock assessment models.

## **2.2.2 Independent Gill-Net Survey (Program 915)**

The Fisheries-Independent Gill-Net Survey, also known as Program 915 (P915), employs a random survey design stratified by area and depth that has sampled in Hyde and Dare counties (Pamlico Sound) since 2001 and in the Tar-Pamlico, Pungo, and Neuse rivers since 2003. Sampling in the Cape Fear and New rivers was added in 2008, and sampling in the Central Region (Bogue Sound, Core Sound, White Oak River, etc.) has occurred since 2018.

The goal of the survey is to maintain long-term fisheries-independent surveys that will provide data on catch composition, relative abundance, size, and age for key species taken in the survey. The survey occurs over much of the habitat commonly utilized by striped bass and is used to calculate annual indices of abundance in major North Carolina estuaries for key estuarine species including striped bass.

### **2.2.2.1 Survey Design & Methods**

The Independent Gill-Net Survey employs a stratified-random sampling design based on area and water depth for each region. Sampling in the Pamlico Sound is divided into two regions: Region 1 includes areas of eastern Pamlico Sound adjacent to the Outer Banks from southern Roanoke Island to the northern end of Portsmouth Island; Region 2 includes Hyde County bays from Stumpy Point Bay to Abel's Bay and adjacent areas of western Pamlico Sound (Figure 2.14). After grid delineation, each region is further segregated into four similar sized areas to ensure that samples are evenly distributed throughout each region.

Sampling in CSMA rivers is divided into three regions: the Pamlico Region includes areas of the Pamlico River from Washington, North Carolina to the mouth of the Pamlico River (south of Wade Point) and the Pungo River from Haystack Point and west to Belhaven and south to Jordan Creek; and the Neuse River from New Bern to Oriental, North Carolina (from Old House Point south to Sandy Point; Figure 2.15). The Central Region includes coastal waters from West Bay to the White Oak River, including parts of Core and Bogue Sounds (Figure 2.16). The area that includes the North River, Back Sound, southern Core Sound, lower portions of Jarrett Bay, and Barden Inlet (estuarine gill net management unit D-1) were removed from the study area to mitigate concerns over interactions with endangered sea turtles; and the Southern Region includes the New and Cape Fear rivers (Figure 2.17).

Each region is overlaid with a one-minute by one-minute grid system (equivalent to one square nautical mile) and delineated into shallow (<6 ft) and deep (>6 ft) strata using bathymetric data from NOAA navigational charts and field observations. NCDMF staff also considered factors such as obstructions to fishing, safety, and accessibility when evaluating each grid for inclusion in the sampling universe. After grid delineation, the Pamlico Sound and Pamlico/Pungo and Neuse rivers (Pamlico Region) are each segregated into four similar size areas to ensure samples are evenly distributed throughout each region. In the Pamlico/Pungo rivers, areas are assigned as follows: upper Pamlico (Washington, North Carolina to Ragged Point), middle Pamlico (Ragged Point to Gum Point), lower Pamlico (Gum Point to Wades Point), and Pungo (Haystack Point south to Sandy Point). In the Neuse River, areas are assigned as follows: upper Neuse (New Bern to Bay Point), upper-middle Neuse (Bay Point to Kennel Beach), lower-middle Neuse (Kennel Beach to Wilkinson Point), and lower Neuse (Wilkinson Point to Gum Thicket Shoal; Figure 2.15).

The Central region is divided into four areas of roughly equal geographic size (Figure 2.16). Area 1 includes West, Long, Cedar Island, and West Thorofare bays, as well as the northernmost part of Core Sound. Area 2 includes Core Sound and all adjoining waters south of area 1 to a line running west from the Clubhouse in Core Sound through the northernmost section of Jarrett Bay. This is the same line that separates the D-1 and B estuarine gill net management areas. Area 3 includes Newport River and adjoining waters, and eastern Bogue Sound to its midpoint. Area 4 includes western Bogue Sound and the White Oak River. The Central Region utilizes only shallow water sets due to depth limitations.

In the Southern region areas are assigned as follows: upper New (from Wilson Bay to Hines Point line extending eastward to French's Creek), lower New (Hines Point to the intersection of the New River and the Intracoastal Waterway), and the Cape Fear River is considered one area (the northern end of US Army Corps of Engineer's Island 13 south to the mouth of the river; Figure 2.17).

SAS/STAT® software procedure PLAN is used to select random sampling grids within each area (SAS Institute 2004). Sampling gear for the Pamlico, Central, and Southern regions consists of an array of gill nets (30-yard segments of 3, 3½, 4, 4½, 5, 5½, 6, and 6½-ISM webbing, 240 yards of gill net per sample). Catches from this array of gill nets comprised a single sample, while two samples (one shallow, one deep), totaling 480 yards of gill net fished, are completed in a sampling trip. In the Cape Fear River and Central Region, only shallow water samples are completed. If adverse weather conditions or other factors prevented the primary grid in an area from being sampled, alternative grids for that area are randomly selected to increase flexibility and ensure completion of sampling requirements each month.

Nets are deployed parallel or perpendicular to shore based on the strata and common fishing techniques for each area. Gear is deployed within an hour of sunset from February 15 to April 30 and September 1 to December 30 and within an hour and a half of sunset from May 1 to August 31. Gear is fished the following morning to keep soak times at a standard 12 hours. In the Southern Region, soak times are reduced to four hours from April 1 through September 30 and deployed within two hours of sunset and fished in the dark (sampling was modified in July 2008). This action was taken to minimize interactions with endangered and threatened sea turtles. Twine size is based on the twine size most frequently used by local commercial fishermen in the corresponding region (Pamlico, Central, and Southern: #177 or 0.47mm). All gill nets are constructed with a hanging ratio of 2:1. Nets constructed for shallow strata have a vertical height between six and seven feet. All deep water nets are constructed with a vertical height between ten and eleven feet. With this configuration, all gill nets fished the entire water column.

Physical and environmental conditions including surface and bottom water temperature (°C), salinity (ppt), dissolved oxygen (mg/L), bottom composition, and a qualitative assessment of sediment size are recorded upon retrieval of the nets on each sampling trip. Reported water temperature, salinity, and dissolved oxygen values are the mean of surface and bottom values at deployment and retrieval of nets. All attached submerged aquatic vegetation (SAV) in the immediate sample area was identified to species and density of coverage is estimated visually when possible. Additional habitat data recorded include distance from shore, presence/absence of sea grass or shell, and substrate type.

Each sampling area within each region is sampled twice a month. For the Pamlico/Pungo and Neuse rivers, a total of 32 samples are completed (eight areas x twice a month x two samples; shallow and deep) each full month. For the Southern Region, a total of 12 samples are completed (New River: two areas x twice a month x two samples; Cape Fear River: one area x four times a month x one shallow sample) each month. Samples are collected from February 15 through December 15 each year. The period of December 16 through February 14 is not sampled due to low catch rates and safety concerns associated with fewer daylight hours and cold water and air temperatures during this period.

#### **2.2.2.3 Biological Sampling**

Each collection of fish (30-yard net) is sorted into individual species groups. All species groups are enumerated and an aggregate weight (nearest 0.01 kilogram, kg) is obtained for most species. Individuals are measured to the nearest millimeter FL or TL according to morphology of the species. Selected species, such as striped bass, are retained and taken to the lab where data on weight, lengths (FL and TL), age structures (otoliths, scales, and/or fin clips), sex, and maturity stage are collected.

#### **2.2.2.4 Potential Biases & Uncertainties**

Although this program was not designed to specifically target striped bass, striped bass occur in large enough numbers to make this survey a valuable data source to help manage this species. Though this survey does not sample the many shallow creeks and tributaries off the main river stems, habitats frequently used by striped bass, the stratified random design of the survey, and the broad area of habitats sampled in the main estuarine system should be sufficient to detect trends in striped bass relative abundance. The range of gill-net mesh sizes used in this survey would exclude the availability of the smallest and largest individuals to the sample gear.

Many factors affect gill-net catch efficiency including net visibility and turbidity (Berst 1961; Hansson and Rudstam 1995), though setting nets overnight may offset some concerns of net visibility. Efficiency can also decrease if nets become tangled or fouled with debris. In Program 915, performance of individual net panels is evaluated and recorded and catch is evaluated at the sample level (catch from a gang of nets is a sample), so performance of individual net panels may not have a large impact on catch from a sample.

#### **2.2.2.5 Development of Estimates**

The relative index is defined as the number of striped bass captured per sample (240 yards of gill net). P915 index precision appears to be good for most strata, months, and years, with some exceptions (Southern Region). The deep strata do not track well with the shallow strata after 2011 (Pamlico Region) and prior to 2005 (Pamlico Sound). Overall, the percent frequency of occurrence is lower and PSE values are typically higher in the deep stratum; thus, the deep stratum was dropped from index calculations. The months of April and October to November are used in index calculation because striped bass are most available to the survey during these months. The Pamlico Sound data were not used due to low catch numbers and concerns about stock assignment. Pungo River data were also excluded due to mixed stock concerns. Central Region data were not used due to the very short time series. In the Southern Region, although striped bass catch rates were

very low, data from the Cape Fear River data were used to calculate an index. New River data were not used in index calculations because striped bass were seldom captured there.

#### **2.2.2.6 Estimates of Survey Statistics**

Samples collected from P915 on the Pamlico, Pungo, and Neuse rivers show most striped bass were captured in the upper and middle portions of the rivers. Over the past twelve years, striped bass indices show relative abundance has been higher in the Pamlico/Pungo and Neuse rivers when compared to the Cape Fear River (Table 2.5; Figure 2.18–2.20). Since 2004, striped bass relative abundance in the Pamlico/Pungo and Neuse rivers ranged from 0.84 to 2.66 fish per sample, whereas relative abundance in the Cape Fear River ranged from 0 to 0.14 fish per sample (Table 2.5). Length frequencies from P915 are represented in Figures 2.21 and 2.22. Length frequency distributions generally follow a normal bell-shaped patterns; however, in 2016 and 2017 in the Pamlico/Pungo and 2015–2017 in the Neuse rivers, there was a higher percentage of small fish that could represent the two year classes of striped bass thought to be the result of successful natural reproduction in 2014 and 2015. Due to a commercial and recreational no possession measure implemented in March 2019, fishery-independent programs like P915 will be the only source CSMA striped bass data while Amendment 2 to the N.C. Estuarine Striped Bass Fishery Management Plan is being developed and adopted.

### **2.2.3 Electrofishing Surveys**

#### **2.2.3.1 Survey Design & Methods**

The objectives of the NCWRC spawning ground surveys are to monitor and quantify population metrics of striped bass migrating to the spawning grounds during spring of each year. Sampling in all rivers normally begins in March and continues into May when water temperatures consistently exceed optimal temperatures for spawning (18–22°C) and striped bass spawning appears complete. The NCWRC uses a boat mounted electrofishing unit (Smith-Root 7.5 GPP; 5000–7000 W; 120 Hz) and either one or two dip netters to collect striped bass as they are observed. To minimize size selection during sampling, striped bass are netted as they are encountered regardless of size. Electrofishing time (seconds) is recorded for each sample site, and relative abundance of striped bass for each sample is indexed by the number of fish caught per hour (fish/h). Water temperature (°C) and other water quality measurements are recorded at each sample site.

In the Tar-Pamlico River, the electrofishing on the spawning grounds began in 1996. The survey uses a stratified random design, although the sampling design was less rigid in early years of the time series. The sample area extends from Battle Park in Rocky Mount to Tarboro, North Carolina and is divided into three approximately 20 km strata (Tar 1 Battle Park = Battle Park to Dunbar; Tar 2 Dunbar = Dunbar to Bell's Bridge; Tar 3 Bell's Bridge = Bell's Bridge to Tarboro town ramp; Figure 2.23). Weekly sampling events consist of boat electrofishing for approximately 1,800 seconds followed by maneuvering downstream several kilometers and sampling again for another 1,800 seconds within a stratum. The starting location of each sample site is randomly chosen within a stratum on a sample day. Sampling within each stratum is attempted each week, but low flow conditions can prohibit sampling in the upper stratum and flood conditions can prevent sampling all strata.

In the Neuse River, striped bass electrofishing surveys began in 1994. Sampling design has varied throughout the time series, but the survey has typically employed a stratified random design.



During some years, opportunistic sample sites were added if catches were low at random sites. Four strata were developed based on observation of striped bass spawning activity near Kinston, Goldsboro, Smithfield, and Raleigh, North Carolina (Figure 2.24). Only the Kinston and Goldsboro strata were sampled from 1994–1997, but Smithfield and Raleigh strata were added after removal of Quaker Neck Dam in 1998. Additionally, the Kinston stratum was only sampled after 1998 during drought conditions. The two primary sampling strata are located near Goldsboro, North Carolina, which is attempted weekly, but ability to sample the Smithfield, North Carolina and Raleigh, North Carolina strata is highly dependent upon accessibility due to low streamflow. Sample sites approximately 1 km in length are randomly selected within strata in most years; however, longer sites were sampled once per week in 2005, 2008, and 2014.

The Cape Fear River striped bass electrofishing survey was initiated in 2003. The survey is a fixed station design with four fixed sites: Buckhorn Dam (rkm 316) near Moncure, Lock and Dam 3 (rkm 186) near Tar Heel, Lock and Dam 2 (rkm 149) near Elizabethtown and Lock and Dam 1 (rkm 97) near Riegelwood (Figure 2.25). Fixed sites are sampled once weekly for 30 minutes of electrofishing time at each site. Sampling occurs immediately downstream of each dam with lock chambers sampled opportunistically during 2014–2016. Lock chamber sampling contributed little to striped bass catches. The number of sampling events per year ranged from eight to 43. Striped bass abundance during March sampling for American shad is typically low; therefore, March samples were excluded from analysis. The Buckhorn Dam site, added in 2014, was also excluded from final analyses because boating access is limited by low flows, the short time series is inconsistent with other sites, and catch rates are typically low at the site.

#### **2.2.3.2 Sampling Intensity**

In the Tar-Pamlico River, NCWRC personnel normally begin striped bass sampling in March and continue into May when water temperatures consistently exceed optimal temperatures for spawning (18–22°C) and striped bass spawning appears complete.

NCWRC sampling on the Neuse River is conducted a minimum of once at each stratum per week during spawning season (dependent on adequate streamflow) and generally occurs April–May.

NCWRC personnel collect striped bass on the Cape Fear River weekly in April and May at each of three sample sites (Lock and Dam 1, Lock and Dam 2, Lock and Dam 3). Sampling continues through May until water temperatures exceed 22°C, or until a decline in CPUE signifies spawning completion. In 2009, sampling effort was standardized to approximately 30 minutes at each sample site. Sampling is typically not conducted when streamflow exceeds 20,000 cubic feet per second (cfs), which creates dangerous sampling conditions.

#### **2.2.3.3 Biological Sampling**

Individual striped bass are measured for TL (mm) and weighed (g). Sex is determined by applying directional pressure to the abdomen toward the vent and observing the presence of milt (male) or eggs (female). Typically, scales are removed from a subsample of fish in the field (target maximum of 15 fish for each sex and 25-mm size-class) on the left side of the fish between the lateral line and the dorsal fin. Before release, untagged striped bass are tagged with an individually numbered internal anchor tag as a cooperative effort with the NCWRC as part of the ongoing NCDMF Multi-Species Tagging Program. A partial pelvic fin clip is collected (approximately 200) and preserved in 95% ethyl alcohol to estimate contribution of hatchery fish to the spawning stock using

parentage-based tagging. Striped bass scales are examined at 24X and 36X magnification using a microfiche reader, and annuli are counted to estimate age in accordance with standard protocols (NCWRC and NCDMF 2011). A subsample of 15 scales per 25-mm size-class per sex (as available) was aged by one reader, and a 20% verification subsample by size class was aged by a second reader. Differences between readers were resolved to establish 100% reader agreement. Subsample ages of the primary reader are compared to the secondary reader to determine ageing precision, and the entire sample is re-aged if bias patterns are detected.

#### **2.2.3.4 Potential Biases & Uncertainties**

Sample stations are often not accessible due to low river levels. This could bias the abundance estimates either by concentrating striped bass in the accessible areas or allowing striped bass to go undetected because of boating obstacles. Biases can also occur due to variation in river discharge; catch rates can be greatly influenced during high and low flows years by making fish less available. Additionally, it is possible that fish may be missed by the dip netter, or that using different numbers of dip netters could impact index calculations. If striped bass are not universally available to the dip netter at all population densities (full range of sizes and ages) during the spawning run, it could bias abundance estimates.

In the Tar-Pamlico River, an attempt is made to distribute sampling evenly among each of the sampling strata; yet, due to low river levels on some sampling days, the lower segment (closer to Tarboro) often receives a slightly greater proportion of the sampling effort. Spring streamflow and associated navigability significantly affect our ability to access spawning areas and may inflate or underestimate striped bass abundance within and among seasons. Analyses of relative abundance indices are further deterred by the lack of well-defined, concentrated spawning grounds such as those found on the Roanoke River.

In the Neuse River, striped bass catch rates can be influenced by streamflow conditions and obstructions to upstream migration. Quaker Neck Dam was removed in 1998, and sample sites further upstream were added thereafter. Upstream strata in Raleigh, North Carolina and Smithfield, North Carolina were added because striped bass had access to the upstream habitats after dam removal. In some years (e.g., 2005, 2008, 2014), entire strata were sampled rather than randomly selecting sites within the strata. Sampling upstream strata is highly dependent upon accessibility due to streamflow, with low flow conditions causing sampling to only occur in lower river strata. In these instances, striped bass potentially utilizing upper river habitats would not be sampled; however, striped bass access to upper river habitats is also limited during low water levels.

In the Cape Fear River, striped bass catch rates are influenced by abundance, habitat below each dam structure, and upstream passage rates through each lock and dam. Since the 1960s, the U.S. Army Corps of Engineers has operated the lock structures each spring for anadromous fish passage. In 2012, a rock arch rapids fishway was completed at Lock and Dam 1 and anadromous fish locking operations ceased at that location. It is likely that this operational change has influenced striped bass catch at each lock and dam due to habitat modification at Lock and Dam 1 and altered passage rates. The number of dip netters has varied (1 or 2) among and within years; however, the number of striped bass encountered on the Cape Fear River never approaches gear saturation with one dip netter; therefore, it is unlikely that catch rates are influenced by a second dip netter.

Other biases could be due to the gear itself. Striped bass of abnormal size may not be as vulnerable to the stunning effects of the electrofishing gear and could escape capture. Electrofishing tends to select for larger fish as they are more visible to the dip netters and have a lower immobilization threshold (Sullivan 1956; Reynolds 1996; Dolan and Miranda 2003; Ruetz et al. 2007). For this reason, the relative abundance of smaller fish is likely biased too low (Reynolds 1996). Collection of fish by netting may be associated with bias. Daugherty and Sutton (2005) demonstrated that capture efficiency was affected by moderate flow rates due to movement of fish out of range of the netters. Schoenebeck and Hansen (2005) indicated how gear saturation caused electrofishing catch rate to be non-linearly related to abundance. Some fish may be less likely to be immobilized by electrofishing gear. Dolan and Miranda (2003) demonstrated how immobilization thresholds were inversely proportional to body size. Conductivity, water temperature, water transparency, dissolved oxygen, depth, flow, and electric current are some of the factors that can impact the efficiency of electrofishing gear (Reynolds 1996; McNerny and Cross 2000; Speas et al. 2004; Buckmeier and Schlechte 2009).

#### **2.2.3.5 Development of Estimates**

Relative abundance of striped bass for each sample was computed as the number of striped bass collected per hour of pedal time of electrofishing (fish/h). For the Tar and Neuse rivers, relative abundance indices and associated standard errors were calculated for all samples each year. For the Cape Fear River, relative abundance and associated standard errors were calculated for each of the three sample sites and for all sites combined. Annual length-frequency distributions were graphically examined using density ridgeline plots (R packages *ggplot2* and *ggridges*; Wickham 2009; Wilke 2019). Fish age and the proportion of non-hatchery fish were determined using PBT when possible. Ages derived using PBT were used in the matrix model.

#### **2.2.3.6 Estimates of Survey Statistics**

##### Tar River

Electrofishing surveys in the Tar River yielded 10,933 individual striped bass from 1996–2018. Total catch ranged from 180 fish in 2017 to 1,429 fish in 2005, and relative abundance ranged between 18.2 and 99.8 fish/h (Table 2.6). Other than peaks in 2005 and 2010, relative abundance was consistently between 25 and 50 fish/h throughout the time series, and an obvious temporal trend was not apparent (Figure 2.26). However, abundance declined during the 2016 to 2018-time period, with the lowest mean CPUE of the survey (18.2 fish/h) occurring in 2018. Striped bass ranged in size from 155–1,190 mm. Length distribution of the Tar River striped bass population was typically unimodal, and the modes progressed in size for several years, suggesting persistence of periodic, strong year classes (Figure 2.27). A high percentage of hatchery fish (83–93%) contributed to Tar River striped bass samples between 2013 and 2016, but the proportion of non-hatchery fish increased in 2017 (30%) and 2018 (59%; Table 1.2)

##### Neuse River

A total of 4,866 striped bass were collected in the Neuse River electrofishing survey from 1994–2018. Total catch ranged from 58 fish in 2006 to 401 in 2003, and the relative abundance index ranged between 4.4 and 20.4 fish/h (Table 2.7). No trend in relative abundance was apparent since 1994, despite the removal of Quaker Neck dam in 1998 and implementation of conservative harvest limits in 2008 (Figure 2.28). Striped bass have ranged in length from 185–1,140 mm. Length distributions of Neuse River electrofishing samples were typically unimodal and the peak

of the distributions occurred around 500 mm or 600 mm (Figure 2.29). Analysis of hatchery contribution indicated the Neuse River striped bass population is mostly composed of stocked fish (Table 1.2). The fish of unknown origin in most years were all large enough to be fish stocked prior to 2010 and therefore not eligible for identification by PBT. In 2018, however, 17% of fish less than 550 mm were non-hatchery.

### Cape Fear River

Total catch of striped bass ranged from a low of five fish in 2006 to a high of 202 fish in 2016 (Table 2.8). Striped bass ranged in length from 158–891 mm. The oldest PBT-aged fish was an age-8 male collected in 2018 and age 8 was the maximum possible PBT age in that survey year. There was little trend in relative abundance for all sites combined throughout the time series; however, the relative abundance index increased at Lock and Dam 1 after the construction of the rock arch rapids in 2012 but has been followed by a declining trend since 2016 (Figure 2.30). Relative abundance has remained low at lock and dams 2 and 3, indicating few fish are migrating above Lock and Dam 1. Length distribution increased between 2007 and 2012 as the stock expanded following the initiation of annual stocking in 1998; however, length distributions are truncated throughout the time series, with few larger (e.g.,  $\geq 700$  mm) fish occurring in the survey (Figure 2.31). Additionally, PBT analysis indicates the stock is overwhelmingly hatchery-origin fish (e.g., 93% in 2018; Table 1.2).

## **2.2.4 Cape Fear Tagging Program**

### **2.2.4.1 Survey Design & Methods**

In 2011, the NCDMF and NCWRC initiated a fishery-independent mark-recapture study to estimate the total mortality and population size of Cape Fear River striped bass using a tag return model. All healthy striped bass were tagged using internal anchor tag and passive integrated transponder (PIT) tags; only data from PIT tagged fish were used for the model. A combination of electrofishing and hook-and-line gears are used to capture fish throughout the Cape Fear River and its tributaries (Figure 2.32). A boat-mounted electrofishing unit (Smith-Root 7.5 GPP) is the primary gear used (2 dip netters) to catch and tag striped bass. A combination of continuous and ambush (intermittent) electrofishing was used during daylight hours. Continuous shocking assures that all habitat types are sampled and particular habitat types are not preferentially selected. To minimize size selection during sampling, striped bass were netted as they were encountered regardless of size.

Striped bass were also tagged using hook-and-line gear during the Cape Fear River Watch Striped Bass Tournament and by a volunteer recreational fisherman trained by NCDMF staff. Additional fish were tagged using run-around gill nets by NCDMF staff to supplement tagging when environmental conditions were not conducive for electrofishing and as part of targeted sampling for the NCDMF Multispecies Tagging Program. Striped bass captured in the Cape Fear River in NCDMF Independent Gill-Net Surveys (Program 915) were also tagged and released if in good condition.

### **2.2.4.2 Sampling Intensity**

Sampling within the Cape Fear River and its tributaries (Brunswick, Black, and Northeast Cape Fear rivers) was conducted by the NCDMF from January to April 2011–2018 (Figure 2.32). Sampling on the Cape Fear River spawning grounds was conducted by the NCWRC at the base of

the three lock and dams from April–June; however, additional samples were collected as part of NCDMF P366 (Multi-Species Tagging Program) throughout the year.

#### **2.2.4.3 Biological Sampling**

All striped bass were scanned for existing PIT tags by NCDMF and NCWRC staff prior to being tagged with an internal anchor tag and an PIT tag. Tagged fish were measured to the nearest millimeter for FL and TL and weighed to the nearest 0.01 kilogram (kg).

#### **2.2.4.4 Potential Biases & Uncertainties**

PIT tag retention was assumed to be 100% and the tag reporting rate was assumed to be 100% because the tag can only be returned by the NCDMF and NCWRC staff through fishery-independent surveys. No angling fishing effort was involved, so the tagging data cannot inform fishing mortality and cannot separate fishing and natural mortalities. Striped bass with estimated ages of 3–7 were tagged in the study, so the estimates only apply to age 3–7 striped bass.

#### **2.2.4.5 Development of Estimates**

Prior to October 1, 2014 all data on striped bass tagged and recaptured as part the Cape Fear River Striped Bass Mark Recapture Study were entered in the NCDMF Biological Database (BDB) according to the Program 311 documentation. As of October 1, 2014, all data are entered into the BDB under the Program 366 documentation (Multi-Species Tagging Program). Following the transition period between tagging programs, data collected in Program 311 was reformatted to match the Program 366 documentation to allow recaptured fish to be linked back to the original tagging event in Program 311 and to be accounted for in the new multi-species tagging program (P366) upon re-release.

Data were extracted from the NCDMF Biological Database (BDB) and transformed into a PIT tag matrix. Only fish that were PIT tagged using electro-fishing and hook-and-line gears within the selected time period of 2012–2018 were included as releases. To minimize bias associated with higher post-release mortality, fish tagged using gill nets were excluded from the analysis. Data from the 2011 field season were excluded from the analysis due to low sample size ( $n=265$ ) and to limit the chance of selection bias. Only tagged fish that were recaptured after seven days at large were included as recapture events. In addition, only the fishery-independent PIT tags recaptures by NCDMF and NCWRC staff were included in the analysis. Recreational anglers were not provided PIT tag readers. Multiple recapture events of the same individual were also removed from the analysis. Also, for ease of analysis, all tagging and recapture events were merged into a single recapture category for the matrix. Missing FL and TL were estimated using:  $FL = (TL * 0.945673822) - 5.277089838$  or  $TL = 6.206909513 + (1.055954699 * FL)$ ; see Appendix 1).

#### **2.2.4.6 Estimates of Survey Statistics**

A total of 3,760 striped bass were tagged and released with PIT tags using all gears from 2011 to 2018 (Table 2.9); however, only 3,450 striped bass were included in the tagging model from 2012 to 2018 (Table 2.10). The majority (88%) of the striped bass included in the model were captured using electrofishing gear. Of the fish included in the model, NCDMF tagged 2,507 striped bass in the mainstem of the Cape Fear River and its tributaries (Figure 2.32). The NCWRC tagged 585 striped bass included in the model on the spawning grounds at lock and dams 1, 2, and 3. Volunteer anglers tagged 358 of the striped bass included in the model using hook-and-line gear at various locations in the Cape Fear River and its tributaries as well as the Northeast Cape Fear River.

Mean length of striped bass that were included in the model ranged from 508.5 mm TL in 2015 to a high of 569.0 mm TL in 2018 (Table 2.11). Minimum TL of tagged striped bass ranged from 192 to 337 mm. Maximum TL of tagged striped bass ranged from 800 to 891 mm. The length-frequency distribution of fish included in the model had bimodal peaks at 375 and 500 mm TL length classes (Figure 2.33). Volunteer anglers using hook-and-line gear primarily tagged larger striped bass, while NCDMF and NCWRC staff tagged fish over a wider range of sizes (Table 2.11; Figure 2.33). Using the results of the 2016 and 2017 genotyping and parentage analyses of Cape Fear River striped bass (Figure 2.34), the length-frequency distribution of striped bass included in the tagging model are thought to represent age three to seven striped bass.

A total of 259 striped bass were recaptured from all gears from 2011 to 2018 (Table 2.12). Two-hundred and twenty-one tag returns (6.4% return rate) from electrofishing gear were included in the tagging model from 2012 to 2018 (Table 2.13). Annual return rates ranged from 1.3% (2018) to 11.8% (2013). Striped bass were recaptured in all sampling areas (Brunswick River, Cape Fear River, Northeast Cape Fear River, and at lock and dams 1, 2, and 3); however, most of the recaptures occurred near downtown Wilmington, North Carolina (Figure 2.35). Distance between release and recapture sites ranged from 0 to 65.0 miles with an average of 6.5 miles and a median distance traveled of 1 mile (Table 2.14). Time at large ranged from 8 to 2,232 days with a mean time at large of 457.5 days (Table 2.15).

Mean length of recaptured striped bass included in the model ranged from 481.2 mm TL in 2018 to 611.6 mm TL in 2012 (Table 2.16). Minimum TL of recaptured striped bass ranged from 359 to 469 mm. Maximum TL of recaptured striped bass ranged from 534 to 845 mm. The length-frequency distribution of recaptured striped bass included in the model had had bimodal peaks at 550 and 650 TL size classes and had a similar distribution as those tagged (Figures 2.33, 2.35). Growth varied by time at large, ranging from 0 to 367 mm (Table 2.17). Twenty negative growth values were removed from the growth estimates and were the result of measuring errors. The mean annual growth rate for all recaptured fish included in the tagging model was 0.190 mm/day and ranged from 0.162 to 0.243 mm/day (Table 2.18).

### **3 DEMOGRAPHIC MATRIX MODEL**

#### **3.1 Objectives**

Objectives of this analysis were to (1) estimate the growth of striped bass in each of the three CSMA rivers (Tar-Pamlico, Neuse, and Cape Fear rivers) using von Bertalanffy growth (VB) model; (2) estimate age-specific natural mortality using the Lorenzen method and growth parameters; (3) develop a demographic matrix model for each system; (4) conduct sensitivity and elasticity analyses to identify critical age classes and demographic parameters for sustaining population growth; (5) evaluate efficacy of hypothetical restoration strategies to aid in management of striped bass and to prioritize recovery efforts in these three rivers. Objectives (1) and (2) provide information on demographic parameters used in the matrix model. The demographic matrix model does not provide population abundance or mortality estimates.

## 3.2 Methods

### 3.2.1 Demographic Matrix Model

An age-structured demographic matrix model was developed to forward project population dynamics for striped bass (Quinn and Deriso 1999; Caswell 2001). In the matrix model, the population vector of abundance  $N_y$  in year  $y$  is multiplied by the projection matrix  $A$  to obtain the population vector of abundance in year  $y+1$ . The top row of the projection matrix specifies the fertility for each age group, which serves as the renewal part of the model. The sub-diagonal of the projection matrix specifies the survival for each age group. The matrix model takes the form as below:

$$N_{y+1} = AN_y,$$

where  $N_y = [N_{y,1}, N_{y,2}, \dots, N_{y,T}]$  is a vector of age-specific population size (i.e., number of individuals) from age one to age  $T$  in year  $y$ , where  $T$  is the maximum observed age for striped bass. In this study,  $T=7$  for Cape Fear River and  $T=11$  for Neuse and Pamlico Rivers based on survey data (previously described programs, with the exception of Program 100) with striped bass otolith and PBT ages (. Notation  $A$  represents the projection matrix with a size of  $T \times T$ :

$$A = \begin{bmatrix} f_1 & f_2 & \dots & f_{T-1} & f_T \\ S_1 & 0 & \dots & 0 & 0 \\ 0 & S_2 & \dots & 0 & 0 \\ \vdots & \vdots & \ddots & \vdots & \vdots \\ 0 & 0 & \dots & S_{T-1} & 0 \end{bmatrix},$$

where  $f_t$ ,  $t = 1, 2, \dots, T$  is the fertility (i.e., number of actual recruitment produced per individual) for age  $t$ , and  $S_t$  is the probability of individuals surviving from age  $t$  to the next age class (or from one year to the next year) and can be calculated as:

$$S_t = \exp(-M_t) \text{ for non-harvested population, and}$$

$$S_t = \exp(-M_t - F_t) \text{ for harvested population,}$$

where  $M$  and  $F$  are instantaneous natural and fishing mortality (per year). Values for  $f$  and  $S$  are non-negative by definition. In this study, the age-specific natural mortality  $M_t$  was estimated using growth information (Section 3.2.2).

The age-specific population abundance can be forward projected by using the projection matrix, which will eventually become stationary. The dominant eigenvalue of this projection matrix ( $A$ ) equals the population growth rate ( $\lambda$ ); the right and left eigenvectors of the projection matrix give the stable size structure of the population and the reproductive contribution of each age class, respectively (Caswell 2001).

### 3.2.2 Growth

In this study, the von Bertalanffy growth (VB) model was used to describe individual growth (von Bertalanffy 1938):

$$L_{i,j} = \left( L_{\infty,j} \left( 1 - \exp\left(-K_j(t_{i,j} - t_{0,j})\right) \right) \right) \exp(\varepsilon_{L,i,j}),$$

$$\varepsilon_{L,i,j} \sim N(\mathbf{0}, \sigma_{L,j}^2),$$

where  $j$  indexes the  $j$ th population,  $L_{\infty}$  is the asymptotic length (mm),  $K$  is the Brody growth coefficient ( $\text{yr}^{-1}$ ),  $t_0$  is the age at which fish has a length of zero,  $L_i$  and  $t_i$  are the length and age of each individual  $i$ , respectively. The observed individual length  $L_{i,j}$  was assumed to follow a lognormal distribution.

In this study, a Bayesian hierarchical approach was used to estimate parameters, in which priors for the growth parameters ( $L_{\infty,j}$ ,  $K_j$ , and  $t_{0,j}$ ) in the VB model were hierarchically structured. The growth parameters were assumed to vary across populations but were constant over time. Specifically, logarithm of population-specific parameters  $L_{\infty,j}$ , and  $K_j$  were assumed to follow a multivariate normal distribution ( $MVN$ ), and  $t_{0,j}$  to follow a normal distribution which were further governed by population-average parameters:

$$\begin{bmatrix} \ln L_{\infty,j} \\ \ln K_j \end{bmatrix} \sim MVN \left( \begin{bmatrix} \ln \bar{L}_{\infty} \\ \ln \bar{K} \end{bmatrix}, \Sigma \right),$$

$$t_{0,j} \sim N(\bar{t}_0, \sigma_{t_0}^2),$$

where  $\bar{L}_{\infty}$ ,  $\bar{K}$  and  $\bar{t}_0$  are population-average parameters describing the growth across populations, and they further follow a uniform distribution. The standard deviation  $\sigma_{t_0}$  was also uniformly distributed. The  $\Sigma$  denotes the variance-covariance matrix that was modeled with an inverse-Wishart distribution (Gelman and Hill 2007):

$$\Sigma = \begin{bmatrix} \sigma_{L_{\infty}}^2 & \boldsymbol{\varphi} \\ \boldsymbol{\varphi} & \sigma_K^2 \end{bmatrix},$$

where  $\sigma_{L_{\infty}}$  and  $\sigma_K$  are standard deviations of  $\ln L_{\infty}$  and  $\ln K$  across populations, and represent spatial variability in growth;  $\boldsymbol{\varphi}$  is the covariance of  $\ln L_{\infty}$  and  $\ln K$  across populations. To improve model convergence given highly negatively correlated  $L_{\infty}$  and  $K$  in VB model, these two parameters were jointly modeled with a negative correlation (Kimura 2008; Midway et al. 2015).

The posterior distribution was obtained through the Metropolis-Hasting algorithm using Markov Chain Monte Carlo (MCMC) simulation (Hilborn et al. 1994; Hoff 2009). Three concurrent chains were run with a total of 100,000 iterations for each chain. The first 70,000 iterations were discarded as burn-in and every 10th of the remaining samples from each chain were saved for analysis. The JAGS (version 4.0.1) was used to run the Bayesian analysis. The data collected from the above fishery-independent surveys during 2004 to 2017 were used to fit the VB growth model.



### 3.2.3 Mortality

The Lorenzen method (Lorenzen 2000; Lorenzen 2005) was used to estimate age-specific natural mortality ( $M_t$ ) for striped bass, which assumes  $M_t$  is inversely proportional to the length at age  $t$  ( $L_t$ ):

$$M_t = M_0 L_t^d,$$

where  $M_0 > 0$  and  $d < 0$  are constants. The constant  $M_0$  can be determined by setting the integral of  $M_t$  equal to the integral of a constant natural mortality  $M_c$ :

$$\int_{t_{\min}}^{t_{\max}} M_t dt = M_c (t_{\max} - t_{\min}),$$

where  $t_{\max}$  and  $t_{\min}$  are the maximum and minimum ages for calculating  $M_t$ . In this study,  $t_{\max} = 7$  for Cape Fear River,  $t_{\max} = 11$  for Neuse and Tar-Pamlico Rivers, and  $t_{\min} = 1$  was fixed for all three rivers. Let  $d = -1$  and  $L_t$  following the VB growth model, i.e.,  $L_t = L_{\infty} (1 - \exp(-K(t - t_0)))$ , then  $M_0$  can be solved as:

$$M_0 = \frac{M_c (t_{\max} - t_{\min}) L_{\infty} K}{K(t_{\max} - t_{\min}) + \ln \left( \frac{1 - \exp(-K(t_{\max} - t_0))}{1 - \exp(-K(t_{\min} - t_0))} \right)}.$$

In this study,  $L_{\infty}$  and  $K$  were set to be the posterior medians estimated from the above growth analysis (Section 3.2.2; Table 3.1). Because natural mortality is one of the most uncertain and difficult-to-estimate parameters in stock assessments (Vetter 1988; Clark 1999),  $M_c$  was modeled using a hierarchical structure in this study. Compared with a non-hierarchical model where the projection matrix is further governed by parameters, in a hierarchical model, both parameters and hyper-parameters determine values in the projection matrix (Caswell 2001; Jiao et al. 2009; Li and Jiao 2015). In the hierarchical model, the  $M_c$  followed a normal distribution  $N(\bar{M}, \sigma_M^2)$  with a mean natural mortality  $\bar{M}$  and a standard deviation  $\sigma_M$ , and the mean was further governed by hyper-parameters  $m_1$  and  $m_2$  in a uniform distribution  $U(m_1, m_2)$ :

$$M_c \sim N(\bar{M}, \sigma_M^2),$$

$$\bar{M} \sim U(m_1, m_2).$$

In this study, the standard deviation of natural mortality ( $\sigma_M$ ) was calculated as:

$$\sigma_m = CV \times \bar{M},$$

where  $CV$  is the coefficient of variation and was randomly assigned a value between 20 and 40%, which has been used as a reasonable uncertainty level in fisheries data analyses (Jiao et al. 2009; Li and Jiao 2015). In this study,  $m_1 = 0.6$  and  $m_2 = 1$  for age 1–3, and  $m_1 = 0.1$  and  $m_2 = 0.5$  was fixed for age 4+. Such parameter values resulted in a natural mortality pattern (Figure 3.1) that approximates previous estimates for striped bass in Neuse River (Bradley et al. 2018b).

The fishing mortality ( $F$ ) can be scaled by the age-specific fishery selectivity ( $g_t$ ) to obtain the age-specific fishing mortality ( $F_t$ ):

$$F_t = Fg_t.$$

In North Carolina, no striped bass harvest is allowed in the Cape Fear River whereas both commercial and recreational harvest occurred in Neuse and Tar-Pamlico rivers prior to spring 2019. In this study, fishery selectivity was estimated (Table 3.1) using 2017 fishery-dependent data for Neuse and Tar-Pamlico rivers and using 2017 fishery-independent data for the Cape Fear River (see section 6). The previous estimates for fishing mortality in the Neuse River ranges from 0.53 to 0.71 (Rachels and Ricks 2015; Bradley et al. 2018b). Therefore, in this study, the matrix model was tested at six fishing intensities (i.e.,  $F = 0, 0.2, 0.4, 0.6, 0.8, 1$ ) to represent possible fishing intensities in these three systems.

### 3.2.4 Reproduction

In this study, a pre-breeding population was assumed and thus, the age-specific fertility ( $f_t$ ) is a product of the age-specific fecundity ( $E_t$ , the number of eggs produced per mature female), the proportion of viable eggs ( $x$ ), the survival of offspring from birth to next census ( $S_0$ , i.e., the survival of offspring through the first year), and the age-specific maturity ( $w_t$ ):

$$f_t = E_t \times x \times S_0 \times w_t \times 0.5,$$

where the value of 0.5 was multiplied because a 1:1 sex ratio was assumed. In this study,  $x = 0.64$  based on a study for the Neuse River (Buckley et al., 2019), and  $S_0 = 0.000017$  based on a single field study that measured the survival of eggs, yolk-sac larvae (from hatching to complete absorption of yolk-sac), and postlarvae (from yolk-sac absorption to demersal or fully developed juvenile) for striped bass (Table 1 in Dahlberg 1979).

In this study, the age-specific fecundity ( $E_t$ ) was derived from the survey data collected from the Neuse River and Tar-Pamlico River during 2013–2014 (Knight 2015):

$$\text{Cape Fear: } \ln(E_t) = 12.484 + 0.205t,$$

$$\text{Neuse: } \ln(E_t) = 12.52 + 0.214t,$$

$$\text{Tar-Pamlico: } \ln(E_t) = 12.429 + 0.203t,$$

where  $t$  is age, and the relationship for Cape Fear River was developed by pooling all data from the Neuse River and Tar-Pamlico River because no fecundity data are available for the Cape Fear River.

In the survey data for Neuse and Tar-Pamlico rivers, striped bass older than three years old are 100% mature (Knight 2015); however, striped bass in the Roanoke River may reach 100% maturity at age five or six (Olsen and Rulifson 1992; Boyd 2011). Therefore, in this study,  $w_i = 0$  for age  $\leq 2$  and  $w_i = 1$  for age  $\geq 5$ . Due to uncertainty in maturity estimates for ages 3–4, similar to natural mortality, a hierarchical structure was developed to describe the maturity for these two ages:

$$w_i \sim N(\bar{w}, \sigma_w^2),$$

$$\bar{w} \sim U(w_1, w_2),$$

$$\sigma_w = CV \times \bar{w},$$

where  $CV = 20\text{-}40\%$ ,  $w_1 = 0.29$  and  $w_2 = 1$  for age 3, and  $w_1 = 0.94$  and  $w_2 = 1$  for age 4.

### 3.2.5 Elasticity and Sensitivity

In a demographic matrix model, elasticity analysis can help compare the relative influence of different age classes to the population growth rate ( $\lambda$ ), and therefore identify critical age classes to focus on in management. The elasticity is defined as the proportional change in population growth rate in response to the proportional change in matrix parameters  $\theta$  (Caswell 2001). The definition of sensitivity is similar to elasticity except that sensitivity is defined using the absolute change in growth rate and in matrix parameters. In this study, elasticity was calculated through Monte Carlo simulation (Jiao et al. 2009; Li and Jiao 2015):

$$\text{Elasticity } \frac{\theta}{\lambda} \frac{d\lambda}{d\theta} = \frac{\theta}{\lambda} \frac{\Delta\lambda}{\Delta\theta},$$

$$\text{Sensitivity } \frac{d\lambda}{d\theta} = \frac{\Delta\lambda}{\Delta\theta},$$

where  $\theta$  can be survival or fertility for each age class.

### 3.2.6 Evaluation of Fishery Management Strategies

In this study, five stocking strategies were evaluated and under each stocking strategy, six fishing strategies were tested. Simulations were run for the Neuse River only because the conclusions would be consistent across rivers given the similar life history characteristics among populations. In the stocking scenarios stocked fish were assumed to be age 1; Phase II fish are hatched in the spring and released in the winter near the end of their first year of life. The five stocking strategies included: (1) no stocking; (2) stocking 100,000 fish per year with 2-years stocking and 2-years no stocking alternating for 15 years (8 years of stocking in total); (3) stocking 500,000 fish per year with 2-years stocking and 2-years no stocking alternating for 15 years (8 years of stocking in total); (4) stocking 100,000 fish per year with 8-years continuous stocking; (5) stocking 500,000 fish per year with eight-years continuous stocking.

The six fishing strategies included: (1) baseline scenario in which the fishing mortality was set at  $F = 0.53$  based on the estimates for Neuse River (Bradley et al., 2018b); the fishery selectivity in Table 3.1 was used; the fishery selectivity in Table 3.1 was used; (2) 26-inch (approximately five years old) minimum size limit scenario in which fishery selectivity  $g_t = 1$  for fish of five years and older and  $g_t = 0$  for fish younger than five years; (3) 2-year closure scenario in which  $F = 0$  for the first two years; (4) 5-year closure scenario in which  $F = 0$  for the first five years; (5) 10-year closure scenario in which  $F = 0$  for the first 10 years; (6) a scenario with 5-year closure, followed by 26-inch minimum size limit.

A 15-year time period was used according to Morris and Doak (2004) who suggests a minimum number of ten years to examine the population trend in the population viability analysis. Additionally, a 15-year time period is relatively sufficient given the observed maximum age of striped bass (11 years) in our study systems. The initial population size was set at 5,000 fish. The initial size structure was constructed based on information from the most recent surveys on the Neuse River (North Carolina Wildlife Resources Commission, personal communication; Table

3.2). The adult abundance (age  $\geq 3$  year) and old adult (age  $\geq 6$  year) abundance over time was tracked for each scenario. Results were obtained from 10,000 Monte Carlo simulation runs. For each run, parameter values in the projection matrix were randomly drawn from corresponding statistical distributions. Extreme values (i.e., within 2.5% of the lower and upper bounds of the distribution) were discarded to avoid unrealistic combinations of parameter values.

### 3.2.7 Results

The Neuse River had the largest population growth rate estimates with medians ranging from 0.87 to 1.13, followed by Tar-Pamlico (medians ranging from 0.86 to 1.1) and Cape Fear rivers (medians ranging from 0.75–1.01; Table 3.3). Estimated population growth rates and the probability of population increasing (i.e.,  $\Pr(\lambda > 1)$ ) declined with increased fishing intensity. Even without fishing allowed, the striped bass in these three rivers would barely sustain, with the medians of population growth rate being slightly above one, and the probability of population growing ranging from 0.52 to 0.8. At fishing mortality rates  $\geq 0.4$ , median population growth rates for all three populations dropped below one, and there was less than 50% probability that the population would grow.

Elasticity of population growth rate to survival and fertility and age-specific reproductive contribution showed similar patterns across three rivers (Figure 3.2). Regardless of fishing intensity, survival and fertility of younger fish influenced population growth rate more than older-age fish, whereas older fish contributed more than younger fish to reproduction due to higher fecundity. As fishing mortality increased, the influence of older fish survival and fertility on the population growth rate decreased, while the influence of younger fish increased. Population growth rate was sensitive to the proportion of viable egg, and age-0 survival but not to the asymptotic length parameter in the growth model (Figure 3.3). As the viable egg proportion and age-0 survival increased, population growth rate estimates increased.

Stocking scenarios produced greater abundance than the scenario with no stocking, and stocking more fish resulted in greater abundance than stocking fewer fish (Figures 3.4 and 3.5). Regardless of stocking strategy, the fishing strategy with 10-year closure was most effective to increase abundance for adults (age 3+), followed by the strategy with 5-year closure combined with 26-inch size limit after closure. The 26-inch size limit strategy was competitive with the closure-size limit combo strategy to increase adult and older adult abundance in stocking scenarios. However, in the no stocking scenario, it was far less effective than the combo strategy. Although 10-year closure was the most effective for age-3+ adults during the first 13 years of simulation, its effectiveness to increase old adult abundance was reduced dramatically once the closure ended and fishing selectivity reverted to the 18-inch size limit, and it became less effective than the combo strategy in no stocking scenario, and less effective than both the 26-inch size limit strategy and the combo strategy in stocking scenarios during the last two years of simulation (Figure 3.5).

### 3.3 Discussion

Fishing activities driven by fishery selectivity that targets older and larger fish not only reduce fish abundance but also alter age structure of the population. As fishing intensity increased, the influence of older fish survival and fertility to population growth rate decreased in response to massive reduction in their abundance due to fishing. The influence of younger fish survival and fertility increased as fishing mortality increased due to their increased proportion in population

abundance. Reproductive contribution by each age group depends on both abundance and fertility of the age group. Although fishing reduces abundance of older fish faster than younger fish, older fish have far greater fertility than younger fish. Thus, the fertility of young fish is too low to offset the reduction in its abundance, and its reproductive contribution became smaller as fishing mortality increased. As the reproductive contribution of younger fish declined with increased fishing mortality, the contribution of older fish rose. This is congruent with Secor (2000), who found that older striped bass contribute far more to reproduction than young age classes, even in populations experiencing fishing mortality. Given that fishing mortality typically impacts older age-classes more than younger age-classes, it is apparent that even relatively moderate levels of fishing mortality can substantially reduce the reproductive potential of a population.

Sensitivity analysis suggests the demographic matrix model is very sensitive to several assumed values. Choice of age-0 survival ( $S_0$ ) has the greatest influence on model results, with relatively small parameter changes resulting in dramatic changes to the population growth rate (see Table 3.3). In fact, the modeled variability in  $S_0$  was likely minimal compared to realized  $S_0$  in systems with natural striped bass recruitment, which regularly experience order-of-magnitude changes (e.g., Cowan et al. 1993; Martino and Houde 2010). Regardless, the choice of any  $S_0$  is arbitrary as PBT analyses and NCDMF juvenile surveys have demonstrated little age-0 survival in all three systems. This prevents interpretation of the estimated level of the population growth rates; rather, the demographic matrix model best serves as a comparison of the relative efficacy of the several management scenarios investigated.

Finally, the maximum age in each river system was based on the oldest observed fish aged using either PBT or otoliths. NCWRC scale-aged fish have demonstrated longevity greater than the modeled maximum age in each system, and striped bass are known to reach ages in excess of 20 years throughout their range including stocks in other regions. Research has found, however, that scales underestimated ages of older fish when compared to otoliths (Welch et al. 1993; Secor et al. 1995; Liao et al. 2013). Additionally, results of the Age Comparison study (see section 7) show that otoliths provide a more precise and accurate age estimate for CSMA striped bass when compared to scales. Nevertheless, the demographic matrix model results as evaluated with a maximum age of 7 years in the Cape Fear River and 11 years in the Neuse and Tar-Pamlico rivers are likely conservative; models that allow survival to older age classes will likely result in greater divergence in model results among the fishing mortalities investigated.

## **4 TAGGING MODEL**

### **4.1 Objectives**

Objectives of this analysis were to (1) estimate total mortality of striped bass (*Morone saxatilis*) in Cape Fear River using a tagging model; (2) estimate abundance of striped bass in Cape Fear River based on Jolly-Seber method.

### **4.2 Methods**

#### **4.2.1 Tagging Data**

PIT tagging data from 2012 to 2018 were used in this analysis. No recreational fishing effort was involved in this study; PIT tags could only be returned by NCDMF or NCWRC staff through fishery-independent surveys. The tagging data cannot inform fishing mortality and cannot separate

fishing and natural mortalities, and thus only total mortality was estimated in this study. Striped bass ages were estimated to range from 3–7 years old based off of length frequency data, so the estimated total mortality only applies to striped bass ages 3–7.

#### 4.2.2 Tagging Model

In the tagging model (e.g., Jiang et al. 2007; Bacheler et al. 2009; Ellis et al. 2018), the observed number of tags returned from fish tagged and released in period  $i$  and captured in period  $j$  ( $j \geq i$ ),  $X_{ij}$ , follows a multinomial distribution with parameters  $R_i$  and  $P_{ij}$ . The parameter  $R_i$  is the total number of tags from fish tagged and released in period  $i$ ,  $P_{ij}$  is the probability of a tag returned from a fish tagged and released in period  $i$  being captured in period  $j$ . In the model,  $\sum_j X_{ij} = R_i - X_{i, unknown}$  and  $\sum_j P_{ij} = 1 - P_{i, unknown}$  where  $X_{i, unknown}$  and  $P_{i, unknown}$  are the total number of tags and the probability of a tag returned from fish tagged and released in period  $i$  with unknown destiny (i.e., have never been captured) by the end of the study period. The parameter  $P_{ij}$  can be estimated as follows:

$$P_{ij} = \phi \lambda \rho S_c (1 - S_{ij}) \frac{U_j}{U_j + Z_j + \Omega},$$

$$S_{ij} = \begin{cases} \exp(-U_j - Z_j - \Omega) & \text{when survey occurs in period } j \\ \exp(-Z_j - \Omega) & \text{otherwise} \end{cases},$$

where  $S_{ij}$  is the survival of tags in period  $j$  from fish tagged and released in period  $i$ ,  $\phi$  is survival from tagging procedure,  $\rho$  is immediate tag retention probability,  $\lambda$  is tag reporting rate, and  $\Omega$  is tag loss. In this study, PIT tags were used and only NCDMF and NCWRC staff can return the tags through a fishery-independent survey, and thus it was assumed that  $\phi = 1$ ,  $\rho = 1$ ,  $\lambda = 1$  and  $\Omega = 0$ . The parameter  $S_c$  is cumulative survival of tags from fish tagged and released in period  $i$  before being captured in period  $j$  and can be calculated as:

$$S_c = \begin{cases} 1 & \text{when } j = i \\ \prod_{v=i}^{j-1} S_v & \text{when } j > i \end{cases}.$$

Major assumptions for the tagging model in this study include: (1) tagged individuals mix completely with untagged population given that there were 7 days allowed for mixing before starting to recapture fish; (2) all tagged individuals have the same survival and recapture probabilities; (3) tagged individuals have independent fates; (4) a monthly time-step is assumed (i.e.,  $j$  represents  $j$ th month); however, the total mortality was estimated on a yearly basis and was assumed constant over months within the year (i.e.,  $Z_j = Z_y / 12$ ), where  $y$  is the year that month  $j$  corresponds to; (5) tags from the fish that were caught and released with tag intact were treated as though tags were cut off; the new subsequent captures of those fish were ignored (Bacheler et al. 2009).

### 4.2.3 Bayesian Estimator

In this study, the Bayesian approach was used to estimate parameters. The posterior probability of a parameter set ( $\theta$ ) given the observed data ( $X$ ),  $p(\theta|X)$  can be calculated as follows:

$$p(\theta|X) = \frac{f(X|\theta)\pi(\theta)}{\int_{\theta} f(X|\theta)\pi(\theta)d\theta},$$

where  $f(X|\theta)$  is the probability density function of the observed data  $X$  given the parameter set  $\theta$ , and  $\pi(\theta)$  is the prior probability, i.e., the probability density function of  $\theta$ . In the tagging model, the observed data  $X$  include the number of tags returned from each time period ( $X_{ij}$ ), and the parameter set  $\theta$  includes the total number of tags from fish tagged and released ( $R_i$ ) and the probability of a tag returned ( $P_{ij}$ ). With multinomial distribution, the density function  $f(X|\theta)$  is:

$$f(X_{11}, \dots, X_{IJ} | P_{11}, \dots, P_{IJ}, R_1, \dots, R_I) = \prod_i \left( \frac{R_i!}{X_{i1}! \dots X_{iI}! X_{i, unknown}!} P_{i1}^{X_{i1}} \dots P_{iI}^{X_{iI}} P_{i, unknown}^{X_{i, unknown}} \right),$$

where  $J$  is the end return time period and  $I$  is the end release time period. The posterior distribution was obtained through the Metropolis-Hasting algorithm using Markov Chain Monte Carlo (MCMC) simulation (Hilborn et al. 1994; Hoff 2009). Three concurrent chains were run with a total of 50,000 iterations for each chain. The first 20,000 iterations were discarded as burn-in and every 10th of the remaining samples from each chain were saved for analysis. The software JAGS (version 4.0.1) was used to run the Bayesian analysis.

### 4.2.4 Model Priors

Non-informative priors (i.e., uniform priors) were used for parameters in the tagging model, except for total mortality  $Z_y$  (Table 4.1). In this study, a hierarchical prior was used for  $Z_y$  where  $Z_y$  follows an unknown lognormal distribution centering around  $\bar{Z}$  that is further governed by a uniform distribution bounded by  $z_1$  and  $z_2$ :

$$Z = \bar{Z} \exp(\varepsilon_z)$$

$$\bar{Z} \sim \text{Uniform}(z_1, z_2),$$

where  $\varepsilon_z \sim \text{Normal}(0, \sigma_z^2)$  is a random error representing the variation in total mortality. Based on previous studies,  $z_1 = 0.1$  and  $z_2 = 1.5$  (Bradley et al. 2018b).

### 4.2.5 Abundance Estimate

The Jolly-Seber method (Seber 1982) was used to estimate abundance of age 3–7 striped bass in the Cape Fear River:

$$N_y = \frac{R_y}{\alpha_y},$$

where  $N$  is abundance,  $y$  indexes year,  $R$  is the total number of tags from fish tagged and released, and  $\alpha$  is the capture probability, i.e., the probability that a tagged fish is captured. The tag recovery probability can be calculated as:

$$\alpha_y = \left(1 - \exp(-U_y - Z_y - \Omega)\right) \frac{U_y}{U_y + Z_y + \Omega}.$$

### 4.3 Results

Median estimates of instantaneous total mortality ( $Z$ ) for age 3 to 7 striped bass ranged from 0.53 (2017) to 1.13 (2014; Table 4.2; Figure 4.1). Total mortality estimates were high in 2012 (median = 0.96; 95% credible interval (CI) = 0.53–1.43) and 2014 (median = 1.13; 95% credible interval (CI) = 0.71–1.47). In 2013, total mortality was low (median = 0.58; 95% credible interval (CI) = 0.21–1.00), and declined in 2015, until another increasing in 2018. Early years (2012–2014) were associated with less uncertainty than the later in the time period (2015–2018).

Abundance estimates ranged from 1,578 (2017) to 10,983 (2012) (Table 4.2; Figure 4.1). Abundance estimates consistently declined over the study period (2012–2018). Abundance in 2018 (median = 1,914; 95% CI = 1,415–,765), was reduced to less than 20% of the abundance in 2012 (median = 10,893; 95% CI = 5,418–23,479). Abundance estimates had greater uncertainty in earlier years of the study period. Median capture probability estimates ranged from 0.04 (2012) to 0.22 (2017; Table 4.2).

### 4.4 Discussion

Previous estimates of total mortality for adult striped bass in Neuse River, Tar-Pamlico River, and Albemarle Sound-Roanoke River ranged from 0.33 to 1.52 on average (Callihan et al. 2014; Harris and Hightower 2015; Rachels and Ricks 2015; Bradley et al. 2018b). These systems are more intensively subject to fishing than Cape Fear River, which would result in higher total mortality in these systems if assuming similar natural mortality. Total mortality estimates for the Cape Fear River fell within the range from previous studies on North Carolina striped bass.

Collier et al. (2013) estimated total mortality and abundance for adult striped bass in the Cape Fear River using tagging data from 2011 to 2013. The study estimated an average total mortality of 0.24 per year (95% CI = 0.02–0.59), a median annual abundance of 15,209 with a 95% CI between 5,000 and 25,000 (Figure 4.1). The authors reported a capture probability ranging from 0.01 to 0.03. Compared to estimates from Collier et al. (2013), total mortality estimates for 2012 and 2013 had a median of 0.96 and 0.58 respectively, which is three times and 1.4 times greater than their estimates. Estimates of abundance for 2012 was not significantly different from the Collier et al. (2013), and estimates of capture probability for 2012 (median = 0.04; 95% CI = 0.02–0.07) were close to the range reported by Collier et al. (2013); however, abundance estimates for 2013 (median = 4,535; 95% CI = 3,024–6,921) were 70% lower, and capture probability estimates were 3 to 4 times higher than the Collier et al. (2013) estimates (Figure 4.2). Collier et al. (2013) accounted for fish movement between four locations within the Cape Fear River, emigration and immigration, which may have contributed to their lower total mortality estimates, lower capture probability estimates and higher abundance estimates compared to this study. While striped bass in the Cape Fear River are thought to remain in the river year around, Raabe et al. (2019) detected a fish leaving



the telemetry array at the river mouth and in 2017 a fish tagged with an anchor tag at Lock and Dam 1 was recaptured by a recreational angler on the Roanoke River.

Estimates of striped bass recreational fishing effort and discards reported by the NCDMF recreational creel survey were substantially higher in 2014 compared to other years (2013, 2015–2018) surveyed (Table 4.3; Figure 4.3), although survey probabilities may be imprecise because they are not set up for striped bass and the estimates have high PSEs. The estimated recreational fishing effort (number of hours fished) was approximately 1.5 times the effort reported in 2016 and three times those in 2013 and 2015; the estimated discards in 2014 were 3 to 64 times higher than other years in the survey. Thus, the high total mortality estimates in 2014 in this study may be caused by high fishing and discard mortalities. In September of 2018, Hurricane Florence made landfall at Wrightsville Beach, North Carolina, causing extensive damage and extreme flooding along the Cape Fear River and its tributaries. Heavy flooding after the storm led to large fish kills due extended periods of hypoxic conditions along the Cape Fear River, likely contributing to the increased mortality estimates observed in 2018. NCDMF staff observed 574 dead striped bass at Battleship Park over the course of two days following the storm (Figure 4.4). Twenty-three anchor tags were recovered from fish tagged with both anchor and PIT tags. NCDMF staff could not access the Wilmington Regional Office due to the hurricane, thus these fish were not scanned to determine if PIT tags were present. If these fish were included in the model, the 2018 total mortality estimates would likely be much higher than those reported for just PIT tag returns alone. The small number of tag returns during 2015–2018 may have also contributed to the high uncertainty of total mortality estimates in this time period. An average of 46 tags were returned per year during 2012–2014, whereas an average of 21 tags were returned during 2015–2018. The low uncertainty in capture probability estimates in early years numerically led to the large variation in abundance estimates given the total number of fish released annually was a known constant.

The use of PIT tags has proven to be an effective means to collect biological data for a variety of species (Gibbons and Andrews 2004; Marvin 2012). While the cost of PIT tags exceeds that of traditional anchor tags, their high retention rate, low mortality associated with tagging, and their ability to retain a fish's identity after multiple recapture events makes them ideal in systems such as the Cape Fear River. In 2019, additional money was secured through the NCDMF's Multi-Species Tagging Program (P366) to continue PIT tagging striped bass in the Cape Fear River. Models used to estimate parameters such as mortality and abundance often have the highest amount of uncertainty for the terminal year. Thus, adding additional years of data to the model should lower the variation in abundance and uncertainty of the total mortality estimates observed during 2015–2018. This additional data should also give managers a better understanding of the true impact of Hurricane Florence on striped bass in the Cape Fear River.

## **5 GLM ANALYSIS ON COMMERCIAL & RECREATIONAL FISHERIES DATA**

### **5.1 Objectives**

The linear regression analysis was extended in Rachels and Ricks (2018) by adding recreational data for the striped bass population in the Neuse River. The goal of this analysis was to identify important factors that influence the response variable (i.e., the relative annual variation in spawning stock mortality). The details of Rachels and Ricks (2018) analysis can be found in Appendix 2.

## 5.2 Methods

The time period of the analysis was confined to 2004–2015 because recreational data are only available since 2004. Along with the four predictor variables that were used in Rachels and Ricks (2018), namely commercial gill-net effort (number of trips), commercial harvest (kg), summer temperature (°C) and summer dissolved oxygen (DO, mg/L), five predictor variables were added to represent recreational fishing activities in this analysis. These five recreational variables included recreational effort (number of trips), recreational harvest (kg), recreational discard (number), recreational total catch (number) and recreational total removal (number, catch + dead discard). The same exact assumptions and procedures were followed as in Rachels and Ricks (2018). These assumptions included: (1) a simple linear regression was applied; (2) original data (both response and predictor variables) were transformed by taking the difference between every two years, i.e., the variation relative to previous year (relative annual variation) and the transformed data were then used in the regression; (3) a one-year delay was applied to all predictor variables except commercial harvest. The one-year delay for commercial gill-net and environmental factors were based on Rachels and Ricks (2018), and the one-year delay for recreational variables was based on the same rationale that the recreational fishing occurs in fall, after the survey sampling season for the current year.

The sensitivity of model outcomes to a series of scenarios was explored further (Figure 5.1). These scenarios included a combination of (1) how long time series of data to use, i.e., a time period of 1994–2015 as in Rachels and Ricks (2018) or a shorter time period of 2004–2015 when recreational data are available; (2) whether or not to apply one-year delay to the variable commercial gillnet effort; fishing effort and fishery harvest are generally considered to occur simultaneously and to associate together; (3) whether or not to transform data, i.e., using relative annual variation or using original data; (4) what error distribution to assume when using original data, i.e., normal error as in Rachels and Ricks (2018) or lognormal error that can describe the possible nonlinear relationship between response and predictor variables; lognormal error cannot be applied when using transformed data due to negative values in response variable that are generated during transformation, and thus only normal error was applied. A stepwise variable selection procedure was used to select the most important factors based on Akaike information criterion, AIC (e.g., Li et al. 2016). This procedure starts with a model only including an intercept. At each step, the variable that reduces the AIC value most or shows the most significant effects (i.e., the smallest P-value) on the response variable will be selected into the model. This step is repeated until including an additional variable will not lead to substantial improvement to model goodness-of-fit.

## 5.3 Results

A total of 31 candidate models were tested, of which eight models had  $\Delta AIC_c$  values less than two (Table 5.1). In this analysis, the eight candidate models with a  $\Delta AIC_c$  value less than two are considered equally plausible in terms of goodness-of-fit and parsimony. The variables contained in these eight models included commercial effort, commercial harvest, recreational effort, recreational discard, recreational total catch and recreational harvest. The model with commercial effort had the highest weight ( $w_i=0.15$ ), followed by the model with commercial effort and commercial harvest ( $w_i=0.086$ ), the model with recreational effort ( $w_i=0.081$ ), and the model with recreational discard ( $w_i=0.071$ ). This result suggested the relative annual variation in both

commercial and recreational fisheries related factors such as fishing effort and removal (including harvest and discard) could play an important role in driving the relative annual variation in total mortality of striped bass in Neuse River.

Sensitivity results showed that model outcomes could be very sensitive to the assumptions that were tested (Figures 5.1 A–C). First, commercial gill–net effort being one-year lagged had a great impact on the outcomes, especially when using data from 2004–2015. For example, using transformed data from 2004–2015, when commercial gill–net effort was not one-year lagged, none of the predictor variables were significant, regardless of including recreational information; however, when commercial gill–net effort was one-year lagged, the variables commercial gill–net effort, commercial harvest, and DO were significant (Figures 5.1 B and C). Second, use of transformed data versus non-transformed data greatly influenced the model outcomes, especially for data from 2004–2015. For example, using non-transformed data from 2004–2015 with recreational information considered and no one-year lag being applied to commercial gill–net effort, the variable commercial gill–net effort was selected as the most significant factor, followed by recreational effort and summer temperature; by contrast, none of the variables showed significant impacts when using transformed data (Figure 5.1 C). Third, whether to include recreational information was critical to determine the model outcomes. For example, using non-transformed data from 2004–2015 with commercial gill-net effort being lagged by one year, none of the variables were selected (Figure 5.1 B) whereas the variables recreational effort and summer temperature were significant when adding recreational information, regardless of the model error distribution (Figure 5.1 C). Model error distribution showed little impacts on model outcomes.

## **5.1 Discussion**

Although using different time series of data due to the availability of recreational data, both this analysis and Rachels and Ricks (2018) documented commercial effort as an important predictor of striped bass mortality in the Neuse River. Model averaging analysis by Rachels and Ricks (2018) indicated commercial gill-net effort was far more influential than the other parameters that were examined. Although Rachels and Ricks (2018) did not include recreational effort or harvest due to benefits of the longer available time series for commercial data, the study also acknowledged the potential importance of recreational angling on total mortality of Neuse River striped bass. Results from this analysis indicated recreational effort and recreational discards may indeed be as influential on annual striped bass mortality as commercial effort and commercial harvest.

## **6 YIELD-PER-RECRUIT**

### **6.1 Objectives**

Yield-per-recruit analysis can be used to evaluate the impacts of fishing mortality and selectivity on fishery yield. The analysis can be extended to estimate the spawning potential in a stock under different conditions. The results of these analyses can be used to balance management and biological objectives for the population of interest.

In this report, several per-recruit analyses are applied to data characterizing striped bass collected from the Neuse River during 2017. Yield-per-recruit analysis is used to examine the impacts of various minimum length limits and fishing mortality rates on fishery yield in terms of numbers

and weight. Spawning stock biomass- and eggs-per-recruit models were also applied to estimate the spawning potential ratio based on conditions in 2017 and to evaluate how the spawning potential ratio varied under different management scenarios.

Traditional per-recruit analyses have been modified here to allow for age-varying natural mortality and logistic selectivity and to account for both non-hatchery and hatchery-origin fish. Due to low spawning stock sizes and limited recruitment, an annual stocking program has occurred in the Neuse River since 1981 (Table 1.1).

## **6.2 Methods**

### **6.2.1 Data**

#### **6.2.1.1 Description**

The primary source of data characterizing striped bass in the Neuse River comes from the North Carolina Wildlife Resource Commission's (NCWRC) Spawning Stock Survey (Figure 6.1). The goal is to monitor striped bass migrating to the spawning grounds. The survey occurs in the spring and is conducted using boat-mounted electrofishing gear. Sampling is contingent on adequate streamflow to allow boat access to sites. Effort on any one individual sampling event varied from 11 to 58 minutes during 2017 (Figure 6.2). The median sampling time on an individual trip was 19 minutes. The survey began in 1994. Scales were collected for ageing from 1994 through 2015. Beginning in 2015, genetic ages have been taken. Only genetic ages were used in the analyses in this report.

### **6.2.2 Initialization**

#### **6.2.2.1 Initial Number of Recruits**

The analyses applied here (see section 6.2.2) track the development of a fixed number of recruits over time. That initial number is simply used for scaling and all final calculations are computed on a per-recruit basis. Here, the initial number of recruits was set at 1,000 individuals.

#### **6.2.2.2 Age Range**

The minimum age was set at 1 and the maximum age was set at 11. A plus group was set at age 7. The maximum age of 11 was selected based on the maximum (scale) age observed in the NCWRC Spawning Stock Survey since it started in 1994. The plus group was selected based on the maximum age observed in 2017.

#### **6.2.2.3 Sex Ratio**

In the absence of compelling evidence to the contrary, a sex ratio of 50:50 was assumed in the analyses.

### **6.2.3 Hatchery Fish**

#### **6.2.3.1 Proportion of Hatchery Fish in Population**

Data on origin (hatchery versus non-hatchery) were collected from 266 striped bass in the Neuse River during 2016 and 2017. Of those individuals, a total of 207 (78%) were of hatchery origin. The per-recruit models assumed that 78% of the population was hatchery-origin fish.

### 6.2.3.2 Initial Number of Recruits

The length of stocked hatchery fish ranges from 152 millimeters (6 inches) to 203 millimeters (8 inches). The length of stocked hatchery fish assumed in the analyses was 178 millimeters (7 inches). Because the assumed length of stocked hatchery fish is less than the length at age 1 (252 millimeters or 10 inches; section 6.2.4.1), the minimum age used in the analyses, changing this value will not have an impact on any of the results presented.

## 6.2.4 Growth

Biological data collected from the NCWRC Spawning Stock Survey during 2017 were used in the estimation of growth parameters described below.

### 6.2.4.1 Age-Length

The relationship of age to length was modeled using the von Bertalanffy function:

$$L_t = L_\infty(1 - e^{-K(t-t_0)})$$

where  $L_t$  is total length in millimeters at age  $t$ ,  $L_\infty$  is the theoretical asymptotic average length (if  $K > 0$ ),  $K$  is growth rate at which the asymptote is approached, and  $t_0$  is the hypothetical age at which length is zero.

It was necessary to fit the age-length model using inverse weighting (based on sample size at age) to ensure reasonable parameter estimates due to the low sample sizes at the youngest and oldest ages. The estimated parameters of the von Bertalanffy age-length function are given in Table 6.1 and a graph of the observed and predicted values is shown in Figure 6.3.

### 6.2.4.2 Length-Weight

The relation of length to weight as modeled using:

$$W = aL^b$$

where  $W$  is weight in grams,  $L$  is total length in millimeters, and  $a$  and  $b$  are the parameters that are estimated.

The estimated length-weight parameters are given in Table 6.2 and a graph of the observed and predicted values is shown in Figure 6.4.

## 6.2.5 Mortality

### 6.2.5.1 Fully-Recruited Fishing Mortality

The value assumed for fully-recruited fishing mortality was 0.33 and was derived from a catch curve analysis, which is described in section 6.2.2.1 of this report.

### 6.2.5.2 Discard Mortality

Bradley et al. (2018b) used telemetry and tag reporting data collected from December 2013 through September 2015 to estimate mortality rates of striped bass in the Neuse River. Their estimate of discard mortality was 0.0% so discard mortality was assumed negligible in the per-recruit analyses.

### 6.2.5.3 First Length at Capture

The length at first capture was assumed equal to the current minimum length limit, 457 millimeters (18 inches).

### 6.2.5.4 Pre-Spawning Mortality

Peak spawning in the Neuse River is assumed to occur the second week of April. If natural mortality is assumed to occur equally throughout the year, the proportion of natural mortality that occurs before spawning is 0.35 (4.25/12).

The proportion of fishing mortality that occurs before spawning was estimated by calculating the amount of total catch (commercial plus recreational) that occurs before April. Estimates of commercial landings, commercial discards, recreational harvest, and recreational discards were available by season for 2017. The total catch was computed by season and then the proportion of the total was calculated for each season. The proportion of the total catch occurring in the winter season (January through March) was 0.28. This value (0.28) was assumed for the proportion of fishing mortality that occurs before spawning.

### 6.2.5.5 Natural Mortality

The idea that natural mortality of fishery resources changes with body weight or length is supported by both ecological theory and empirical evidence. For a given species, the youngest life stages tend to experience higher natural mortality than older life stages.

Lorenzen's (1996) approach was used to estimate age-specific natural mortality for striped bass in the Neuse River. This approach requires parameter estimates from the von Bertalanffy age-length growth model (to translate age to length), parameter estimates from the length-weight function (to translate length to weight), and the range of ages for which natural mortality will be estimated.

The growth parameter values reported in section 6.2.1.4 of this report were used to compute natural mortality at ages 1 through 7+, using Lorenzen's (1996) equation. The estimates of natural mortality at age used in the per-recruit modeling are given in Table 2.3 and the relationship is shown in Figure 6.5.

## 6.2.6 Spawning

### 6.2.6.1 Maturity

A logistic model was used to describe the relationship between total length and maturity based on data collected from the Tar and Neuse rivers in 2013 and 2014 (Knight 2015). Data from the Tar and Neuse rivers were combined because too few immature fish were observed in the Neuse River alone to support modeling. The logistic model used was:

$$mat = \frac{e^{a+bL}}{1 + e^{a+bL}}$$

where *mat* is the proportion mature, *L* is total length in millimeters, and *a* and *b* are the parameters that are estimated.

The estimated length-maturity parameter values are given in Table 6.4 and a graph of the observed and predicted values is shown in Figure 6.6. The estimated length at 50% maturity is 471 millimeters total length.

Because no immature fish of non-hatchery origin were observed, it was not possible to consider separate models for non-hatchery and hatchery-origin fish.

#### 6.2.6.2 Fecundity

As with the maturity data, fecundity data collected from the Tar and Neuse rivers in the Knight (2015) study were combined to ensure adequate sample sizes for modeling. There were sufficient numbers to model fecundity relationships separately for non-hatchery and hatchery-origin fish. Linear models were used to describe the relationship between total length and fecundity for fish of each origin type. The relationship for non-hatchery origin fish (Figure 6.7) was estimated as:

$$Fecundity = -3,222,798 + 6,365.4622L$$

where Fecundity is the number of eggs produced per female and L is total length in millimeters.

The linear relationship for hatchery-origin fish (Figure 2.8) was estimated as:

$$Fecundity = -1,875,954 + 4,429.5759L$$

An analysis of covariance (ANCOVA) was used to compare the two linear regressions (Zar 1999). The ANCOVA can test whether the slopes and/or intercepts are significantly different from each other. Here, the ANCOVA found the slopes and intercepts to be significantly different, suggesting it was appropriate to use different fecundity models for fish of different origin.

### 6.2.2 Analyses

A table of symbols, their definitions, and measurement units used in the equations in this section is given in Table 6.5.

#### 6.2.2.1 Catch Curve & Selectivity

A catch curve approach was used to estimate total mortality and selectivity for striped bass. The method developed by Thorson and Prager (2011) estimates logistic selectivity (to avoid the need to choose an age at full selection) in addition to estimating total mortality and incorporates age-varying natural mortality. Traditional per-recruit analyses assume knife-edge selection in which selectivity transitions from 0 at the length (or age) before length (or age) at full recruitment to 1 at the length (or age) at full recruitment. In the analyses here, this assumption was modified to allow for a logistic-shaped selection curve. Selectivity at lengths smaller than the minimum length limit (section 6.2.5.3) was assumed equal to 0 and selectivity at lengths greater than or equal to the minimum length limit was equal to the selectivity predicted by the logistic model (Figure 6.9).

Because the sampling time varied among sampling events (section 6.2.1), the frequency at age was standardized to 19 minutes (Table 6.6). The values assumed for natural mortality at age were those values estimated in section 6.2.5.5 of this report.

Natural mortality at age was subtracted from the estimated total mortality at age for each year to produce annual estimates of fishing mortality at age (Table 6.7). The apical fishing mortality from this vector ( $F_{2017} = 0.33$ ) was assumed for the fully-recruited fishing mortality (section 6.2.5.1) in the per-recruit analyses.

#### 6.2.7 Yield-per-Recruit

Yield-per-recruit models follow a fixed number of recruits and track their growth and mortality over time and evaluate the impacts of various factors on fishery yield. The methods of Thompson

and Bell (1934) and Ricker (1975) have been modified to allow for more realistic conditions in that the modifications allow for age-varying natural mortality and logistic selectivity (in contrast to knife-edge selectivity). The modified approach also allows for contributions to the stock from both non-hatchery and hatchery-origin fish. Note that the model assumes no migration.

### 6.2.7.1 Fishing & Total Mortality

Fishing mortality,  $F$ , at age  $t$  was computed as:

$$F_t = F_{full}S_t$$

where  $F_{full}$  is the assumed value for the fully-recruited fishing mortality (section 6.2.5.1) and  $S_t$  is the vector of selectivity at age (section 6.2.2.1).

Total mortality,  $Z$ , at age  $t$  was calculated as the sum of natural mortality at age,  $M_t$ , and fishing mortality at age:

$$Z_t = M_t + F_t$$

### 6.2.7.2 Population Size

The total population size (in numbers) at the minimum age used in the analyses, age 1, was set equal to 1,000 individuals (see section 6.2.3.2):

$$N_1 = 1,000$$

Total population size at ages older than age 1 (in numbers) was calculated using:

$$N_t = N_{t-1}e^{-Z_{t-1}}$$

The weight (kilograms) of the total population at age,  $B_t$ , was calculated as:

$$B_t = N_t \left( \frac{w_t}{1,000} \right)$$

where  $w_t$  is the individual weight at age in grams.

The number of individuals in the population at age of non-hatchery origin,  $U_t$ , was computed as:

$$U_t = N_t(1 - h)$$

where  $h$  is the assumed proportion of hatchery fish in the population (section 6.2.3.1).

The number of individuals in the population at age that are of hatchery origin,  $H_t$ , was calculated as:

$$H_t = N_t h$$

The weights of non-hatchery and hatchery individuals at age were calculated the same way as the total population weight at age.

### 6.2.7.3 Catch

The total number of individuals in the catch at age,  $C_t$ , was computed as:

$$C_t = N_t \frac{F_t}{Z_t} (1 - e^{-Z_t})$$

The yield per recruit for the entire population, YPR, in numbers was calculated as:



$$\text{YPR} = \frac{\sum_t C_t}{N_1}$$

The weight (kilograms) of the total catch at age,  $W_t$ , was calculated as:

$$W_t = C_t \left( \frac{w_t}{1,000} \right)$$

The yield per recruit in weight (kilograms) for the entire population, WPR, was calculated as:

$$\text{WPR} = \frac{\sum_t W_t}{N_1}$$

### 6.2.8 Spawning Stock Biomass-per-Recruit

The yield-per-recruit analysis can be extended to evaluate the effects of fishing mortality and minimum length limit on spawning potential. The method of Gabriel et al. (1989) has been modified to incorporate age-varying natural mortality.

SSB at age for non-hatchery female fish,  $\text{SSU}_t$ , in weight (kilograms) was calculated as:

$$\text{SSU}_t = pU_t \left( \frac{w_t}{1,000} \right) \text{mat}_t e^{-(fF_t+mM_t)}$$

where  $p$  is the proportion of individuals in the population that are female (section 6.2.2.3),  $U_t$  is the number of individuals in the population that are non-hatchery origin,  $\text{mat}_t$  is maturity at age  $t$  (section 6.2.6.1),  $f$  is proportion of fishing mortality that occurs before spawning (section 6.2.5.4), and  $m$  is the proportion of natural mortality that occurs before spawning (section 6.2.5.4).

SSB per recruit for the non-hatchery female fish,  $\text{SSU/R}$ , in weight (kilograms) was calculated as:

$$\text{SSU/R} = \frac{\sum_t \text{SSU}_t}{N_1}$$

SSB at age for hatchery-origin female fish,  $\text{SSH}_t$ , in weight (kilograms) was calculated as:

$$\text{SSH}_t = pH_t \left( \frac{w_t}{1,000} \right) \text{mat}_t e^{-(fF_t+mM_t)}$$

where  $H_t$  is the number of individuals in the population that are of hatchery origin.

SSB per recruit for hatchery-origin female fish,  $\text{SSH/R}$ , in weight (kilograms) was calculated as:

$$\text{SSH/R} = \frac{\sum_t \text{SSH}_t}{N_1}$$

SSB per recruit for the entire population,  $\text{SSB/R}$ , was computed as:

$$\text{SSB/R} = \text{SSU/R} + \text{SSH/R}$$

### 6.2.9 Eggs-per-Recruit

Eggs-per-recruit models estimate the number of eggs, on average, that a single female produces in a lifetime. By comparing the current estimate of eggs per recruit to an estimate computed assuming no fishing, one can calculate the spawning potential ratio, which is a measure of the reproductive

health of the stock (see below). Goodyear's (1993) approach has been modified to allow for different assumed fecundity relationships for non-hatchery and hatchery-origin fish.

The total number of eggs at age for the non-hatchery female fish,  $EU_t$ , was computed as:

$$EU_t = [pU_t mat_t e^{-(fF_t + mM_t)}] [-3,222,798 + 6,365.4622L_t]$$

Eggs per recruit for the non-hatchery female fish,  $EU/R$ , in numbers of eggs was calculated as:

$$EU/R = \frac{\sum_t EU_t}{N_1}$$

The total number of eggs at age for hatchery-origin female fish,  $EH_t$ , was computed as:

$$EH_t = [pH_t mat_t e^{-(fF_t + mM_t)}] [-1,875,954 + 4,429.5759L_t]$$

Eggs per recruit for hatchery-origin female fish,  $EH/R$ , in numbers of eggs was calculated as:

$$EH/R = \frac{\sum_t EH_t}{N_1}$$

Eggs per recruit for the entire population,  $E/R$ , was computed as:

$$E/R = EU/R + EH/R$$

The spawning potential ratio (SPR) is a measure of the reproductive health of the stock based on fecundity that is calculated relative to the virgin stock condition (i.e., unfished stock; Goodyear 1993). SPR was computed as:

$$\%SPR = \frac{E/R_{F=F_{full}}}{E/R_{F=0}}$$

### 6.3 Approach

The per-recruit analyses were used to estimate SPR based on conditions in 2017 using the values indicated in the descriptions above. Additionally, yield per recruit in both numbers and weight as well as SPR were calculated for combinations of minimum length limits and fully-recruited fishing mortality values. The minimum length limits evaluated ranged from 406 millimeters (16 inches) to 673 millimeters (26.5 inches) at increments of 13 millimeters (0.5 inches). The range of fully-recruited fishing mortality values evaluated was 0.0 to 2.0 at increments of 0.1.

### 6.4 Results

The per-recruit analyses indicated that SPR based on conditions in 2017 was 44%.

In terms of weight, yield per recruit is maximum at minimum length limits ranging from 508 millimeters (20 inches) to 559 millimeters (22 inches) when fishing mortality rates are at the highest levels evaluated ( $F > 1.6$ ; Figure 6.10). Yield per recruit in terms of numbers is maximized at smaller minimum length limits ( $< 500$  millimeters or 20 inches) and fishing mortality rates greater than 0.60 (Figure 6.11). SPR is maximum when fully-recruited fishing mortality is equal to 0.0 (Figure 6.12), which is expected. In the presence of fishing mortality, SPR increases with decreasing fishing mortality and increasing minimum size.

Over the range of fishing mortality rates evaluated, there is not much difference in terms of yield per recruit in weight among minimum size limits less than 610 millimeters (24 inches; Figure 6.13). A different pattern emerges when evaluating yield per recruit in terms of numbers. Regardless of fishing mortality, yield in numbers generally decreases as the minimum size limit increases (Figure 6.14). SPR generally increases as the minimum length increases (Figure 6.15).

At the current size limit, yield per recruit in terms of both weight and numbers is maximized when fully-recruited fishing mortality is 2.0, possibly higher as this was the largest value evaluated (Figures 6.13, 6.14). At a fully-recruited fishing mortality rate equal to 2.0, SPR would be reduced to 10% (Figure 6.15).

## 6.5 Discussion

Balancing management objectives against biological objectives is often challenging. Increasing the harvest rate (i.e., fully-recruited fishing mortality rate) will result in increased yield per recruit (in weight and numbers) but the spawning potential of the stock will be reduced. Increasing the minimum size limit could increase SPR but would result in increased discards, though the mortality of these discards is currently assumed negligible (section 6.2.5.2; Bradley et al. 2018b).

There are a number of uncertainties in the analyses that affect the interpretation of the results. One important issue is that the estimate of fully-recruited fishing mortality assumed in the analyses is likely inaccurate. The estimate was derived from catch curves based on data collected from spawning fish, which are likely not representative of fish in the catch. Bradley et al. (2018b) estimated mortality rates of striped bass in the Neuse River using telemetry and tag reporting data collected from December 2013 through September 2015. Their estimate of harvest mortality of adult striped bass was 0.131. Assuming this value in the per-recruit analyses results in a SPR value of 69%.

Bradley et al. (2018a, 2018b) estimated a fishing mortality of 0.53 and suggested their reported mortality levels were lower than those outside the study area because fishing practices differed between the study area and the entire area used by the population. Bradley et al. (2018b) also estimated an adult natural mortality rate of 0.24. Bradley et al.'s (2018b) estimates of both fishing and natural mortality were not sex- or age-specific and applied to a range of ages (ages 3 to 9 based on length). The average Lorenzen estimate of  $M$  over ages 3 to 9 used in this study is 0.31, which is only slightly higher than 0.24.

Rachels and Ricks (2015) conducted a yield-per-recruit analysis for Neuse River striped bass assuming a fishing mortality rate equal to 0.69 and a natural mortality rate equal to 0.16. They estimated SPR equal to 3% assuming the same minimum size limit as modeled in this analysis (457 mm). The disparity in SPR (3% vs. 44%) and length limits producing maximum yield per recruit between Rachels and Ricks (2015) and this analysis are due to different underlying assumptions regarding Neuse River striped bass growth, longevity, natural mortality, fishing mortality, selectivity, and the contribution of non-hatchery versus hatchery-origin fish.

Rachels and Ricks (2015) assumed a maximum age of 30 years, which has not been observed in the Neuse River stock and so is not reflective of current conditions. Although selection of the maximum age considered can be arbitrary (Ricker 1975), the maximum age used in a YPR analysis, whether 11 years as used here or 30 years as in Rachels and Ricks (2015), can alter model results and should be realistic for the modeled species and system at the time of the analysis.

Assuming an older maximum age in the yield-per-recruit analysis will result in a lower estimate of SPR.

In the yield-per-recruit analysis performed here, selectivity was assumed to follow a logistic curve (i.e., changing with age) as opposed to knife-edge selection assumed in the Rachels and Ricks (2015) analysis, which assumes selectivity equivalent to zero until a pre-defined age at which selectivity is equal to one for that age and all older ages.

Per-recruit analyses do have the advantage of considering both growth overfishing and recruitment overfishing; however, another source of error and a disadvantage to using per-recruit approaches is that they do not account for differences in recruitment at varying stock abundance.

## **7 AGE COMPARISON**

### **7.1 Introduction**

Accurate age determination of fish is one of the most important elements to consider when conducting age structured stock assessments and is crucial information in estimating population parameters including recruitment, natural mortality, and growth.

Striped bass (*Morone saxatilis*) scales and otoliths have been collected sporadically by the North Carolina Division of Marine Fisheries (NCDMF) since 1975, although since 2003 both scales and otoliths have been collected routinely (Table 7.1). Since 1975, a total of 8,949 scale samples have been collected (primary ageing structure for striped bass), with roughly 8,518 collected between 2002 and 2018 (Table 7.1). Very few striped bass otoliths were collected before 2003, however since 2003, 2,122 otoliths have been collected by NCDMF (Table 7.1).

Beginning in 2010, a new genetics technique, termed parental based tagging (PBT), was implemented by the North Carolina Wildlife Resources Commission (NCWRC) to more accurately determine the percent hatchery contribution to striped bass populations in the Central Southern Management Area (CSMA). This method has proven to be greater than 99% accurate in determining if a fish was hatchery produced (Denson et al. 2012). In addition to determining hatchery contribution, PBT samples from hatchery produced fish identify the cohort or year class the striped bass was produced and consequently its age as each parent group is only used once. In 2016, the NCDMF started collecting striped bass fin clip samples for PBT analysis to determine percent hatchery contribution, and age of hatchery reared striped bass collected in the lower portions of CSMA rivers.

Though scale samples were collected by NCDMF from 1975 to 2001, very few striped bass were aged, and no striped bass were aged using scales from 2002–2017. To address the backlog of scale samples in anticipation of the 2017 stock assessment, all striped bass scales from 2002–2017 were processed to be aged.

In 2016, NCDMF began ageing the striped bass scales collected from 2002 to 2017, however concerns were quickly raised about the difficulty in interpreting CSMA striped bass scale annuli and disagreement between readers was high. Additionally, beginning in 2016 exact ages of stocked striped bass through PBT analysis became available.

## **7.2 Objectives**

The objectives of this study were to: 1) determine and compare the accuracy and precision of scale ageing versus otolith ageing for CSMA striped bass, assuming genetic ages are true ages, and 2) to determine the difference in ageing-bias at each age and determine the precision among readers for each method using ages from scales and otoliths.

## **7.3 Methods**

### **7.3.1 Preparation**

#### **7.3.1.1 Scale Preparation**

To prepare scales for ageing, scale impressions were made on acetate sheets with a Carver© heated hydraulic laboratory press and annuli were counted by examination at 24x and 33x magnification on a microfiche reader. For a more detailed explanation of North Carolina Estuarine Striped Bass scale preparation and ageing protocol see the cooperative scale ageing document developed by NCWRC and NCDMF staff (NCWRC and NCDMF 2011).

#### **7.3.1.2 Otolith Preparation**

To prepare otoliths for ageing, a thin sectioning machine was used to section whole otoliths. The water-cooled, thin sectioning machine is equipped with two individual tools; a diamond blade cut-off saw and a precision diamond grinder. The precision grinder is fitted with a dial indicator gauge to control thickness and allows for varied section thicknesses. Both have guide arms for feeding slides to the blades.

Although left and right otoliths are collected, only one side is typically sectioned for ageing. Alternating between left and right otoliths for a species could lead to inconsistencies in the ageing process. The Ageing Lab at the NC Division of Marine Fisheries typically uses the left otolith for sectioning unless the left otolith was not collected or is of lower quality (e.g., crystalized, broken) than the right.

Otoliths are hand held and ground on the transverse plane adjacent to the focus. The purpose of sectioning is to remove both ends of the otolith leaving the transverse section containing the focus. The otolith is then mounted cut side down with the sulcal groove upward onto a frosted microscope slide using an ultra-violet (UV) cure adhesive, Loctite AA 349. After curing, the slides are placed on the guide-arm of the cut-off saw and guided past the saw to remove the bulk of the otolith. Slides are then placed onto the guide arm of the precision grinder and ground down by turning the guide arm adjuster gradually, starting at 1.0 mm thick and stopping at 0.5 mm thick for striped bass, and passing the sample on the guide-arm across the precision grinder.

Once the slides have been ground down, striped bass otolith sections are covered with a top coat. The top coat fills in the rough ground surface of the otolith section providing a clearer view of annuli. In a fume hood, a disposable pipette is used to apply enough Flo-Texx to entirely cover the sample. Adding this cover eliminates the need for polishing most samples.

#### **7.3.1.3 Genetic Sample Collection and Preparation**

A small piece of the pelvic fin was clipped from an individual striped bass and preserved in 95% ethyl alcohol for use in PBT analysis. The South Carolina Department of Natural Resources

(SCDNR) Population Genetics lab conducted microsatellite genotyping for individual fin clips, using a suite of 12 microsatellite markers for striped bass.

### 7.3.2 Age Determination

#### 7.3.2.1 Scale & Otolith Age Determination

Scale and otolith annuli were counted to estimate age and assign a year-class. A minimum of two independent reads were required to age a fish and determine estimates of precision and accuracy. If both readers agreed on an age, that age was assigned to the fish. Discrepancies were resolved by readers sitting together and re-ageing the fish to assign a final age. If an agreement could not be reached, the sample was excluded from further analysis and not used in calculating the age agreement rate with known PBT ages.

#### 7.3.2.2 PBT Age Determination

Since 2010, all broodstock used at the hatcheries to produce the stocked striped bass each year are genotyped (makeup of specific genes as passed on from ancestors). This is done each year, so a genetic record now exists of all the broodstock fish since 2010 used to produce striped bass that are stocked in CSMA rivers each year. This technique can only be applied to striped bass produced in the hatcheries since 2010. Therefore, year-classes produced before 2010 are of unknown origin via PBT. As of 2018, hatchery origin can be determined for all fish that are eight years of age and younger.

### 7.3.3 Comparison Analysis

#### 7.3.3.1 Objective 1

In this analysis, a generalized linear mixed model (GLMM) was used to compare scale ageing versus the otolith ageing. In the model, ageing method was set as a fixed effect. The data included ages from scales (n=445; 2016–2017), otoliths (n=126; 2016–2018), and genetics (PBT ages; n=513; 2016–2018) for the years 2016 through 2018 from 513 total striped bass. PBT ages ranged from one to seven. A total of five readers participated in ageing, among which all five readers read scale ages whereas only two readers read otolith ages. The response variable ( $Y$ ) was the percentage ageing-bias relative to the genetic age (%):

$$Y = (\text{observed age} - \text{genetic age}) / \text{genetic age} * 100,$$

where observed age is either scale age or otolith age. During the ageing process, the same reader aged multiple fish. Thus, in the model, the reader was set as a random effect that assumed the percentage ageing-bias from the same reader was dependent while those from different readers were independent. This random effect represents the variability in percentage ageing-bias among readers, and thus it contributes to explaining the part of the variation that cannot be explained by the fixed effects. The GLMM was developed as follows:

$$Y_{gij} \sim \text{Normal}(\mu + \gamma_j + \alpha_i + \gamma_j * \alpha_i, \sigma_e^2)$$

where  $Y_{gij}$  is the percentage ageing-bias from fish  $g = 1, \dots, n$ , reader  $i = 1, \dots, m$ , and ageing method  $j = \{\text{otolith ageing, scale ageing}\}$ ;  $\gamma_j$  is the ageing method fixed effect,  $\alpha_i$  is the reader random effect. An interaction term between reader and ageing method was included because a reader may be more proficient at one ageing method than the other. The fixed and random effects were modeled as:

$$\alpha_i \sim Normal(0, \sigma_r^2)$$

with priors;

$$\begin{aligned} \mu &\sim Normal(0, 100), & \gamma_j &\sim Normal(0, 100), \\ \sigma_r^2 &\sim Gamma(0.1, 0.1), & \sigma_e^2 &\sim Gamma(0.1, 0.1) \end{aligned}$$

The total variance was  $Var(Y_{ij}) = \sigma_e^2 + \sigma_r^2$ . The relative contribution of reader and error variance are given as the posterior summaries for random error and reader random effects is as follows:

$$p_e = \frac{\sigma_e^2}{\sigma_e^2 + \sigma_r^2} \quad p_r = \frac{\sigma_r^2}{\sigma_e^2 + \sigma_r^2}$$

The analysis was modeled in a Bayesian framework using JAGS version 4.30 (Plummer 2003) in RStudio version 3.6.0 (R Core Team 2013) with 200,000 iterations, three separate chains, a 20,000 iteration burn-in, and thinning set to ten. Convergence was verified by visual inspection of chain trace plots and Rhat values of 1.03 or less.

### 7.3.3.2 Objective 2

Statistical analyses were done using a symmetric test (Hoenig et al. 1995) and coefficient of variation (CV) analysis to determine bias and precision. The data included ages from scales (n=3,611; 2002–2017) and otoliths (n=1,890; 2003–2018) for the years 2002 through 2018 for 4,604 striped bass. Ages ranged from one through 16. Ages were read by seven readers in total and scales were read by all seven readers whereas otoliths were read by only two of the readers. Tests were done for the following comparisons: (1) between otolith and scale ages; (2) between scale readers; and (3) between otolith readers.

## 7.4 Results

### 7.4.1 Objective 1

Preliminary investigation to understand the underlying relationships that explain the variability in the data indicated that the reader identification (ID) might be an important variable (Figure 7.1, Table 7.2) as well as method type (Figure 7.2). The summary statistics from the raw data (Table 7.2) indicated that readers 1 and 2 demonstrated differences in accuracy and precision dependent on the method type. Reader 1 had similar accuracy for both method types but with higher variability for scale ages with mean age bias for otolith ages of -0.899% (sd=8.45) and for scale ages of -0.0935% (sd=15.2); however, reader 2 had very different levels of accuracy and precision between methods with a mean age bias for otolith ages of -1.81% (sd=7.25) and for scale ages of 23.1% (sd=22.3). This demonstrates a need to account for the interaction between reader and method type.

Some ages may be more likely to be underestimated or overestimated than other ages thus, the genetic age of the fish was also initially considered as a possible variable that may affect ageing accuracy and precision (Figure 7.3). However, since the genetic ages were used in the calculation

of the response variable, there was an inherent correlation between these values and thus, genetic age was not used in the analysis.

The results demonstrated differences in accuracy and precision due to reader ID (Table 7.3, Figure 7.4) with readers 1, 3, and 4 tending to slightly underestimate age with low variability indicated by the posterior medians of -0.858% (sd=0.274), -0.931% (sd=0.291), and -1.00% (sd=0.302), respectively. Readers 2 and 5 tended to overestimate with higher variability as shown by the posterior medians of 2.47% (sd=2.48) and 0.280% (sd=0.916), respectively. The posterior median of standard deviation from reader effects was 1.67 (sd=1.46). Reader effects explained 1.7% (sd=4.0) of the total variability in data. The ageing method results (Table 7.3, Figure 7.4) showed that ages from scales tended to overestimate age with a posterior median of 7.90% (sd=3.90, 95% conf. interval=1.37 to 16.6) and ages from otoliths were unbiased with much higher precision than scales demonstrated by the posterior median of -1.19% (sd=0.82, 95% conf. interval=-2.88 to 0.51). The random error standard deviation posterior median was 15.5 (sd=0.261) and accounted for the remaining 98.3% (sd=4.0) of the total variability.

#### **7.4.2 Objective 2**

The comparison between scales and otoliths (Figure 7.5) indicate agreement of 50.7% with a CV of 5.4% ( $\chi^2=1373.2$ ,  $df=64$ ,  $P<0.01$ ). The percentage age-bias plot (Figure 7.6) demonstrates that scales compared to otoliths tended to overestimate ages less than 5 and underestimate ages greater than 5. There was no difference between otolith readers with a CV of 2.2% ( $\chi^2=35.6$ ,  $df=34$ ,  $P=0.392$ ); however, the results from the between reader comparisons for scale ages (Table 7.4) indicate that 11 out of the 13 unique reader combinations were significantly different with CVs ranging from 2.5% to 7.0% and percent agreements ranging from 19% to 88%.

### **7.5 Discussion**

#### **7.5.1 Objective 1**

This analysis demonstrates the importance of understanding and accounting for differences among readers and the interaction between the method type and reader ID to accurately assess the potential bias and level of precision in ageing striped bass. The results from this analysis indicate that scale ageing was biased with comparatively low precision whereas, otolith ageing was unbiased with a higher level of precision. These results are in agreement with previous research where ages from otoliths were significantly different than ages from scales (Secor et al. 1995; Liao et al. 2009). Furthermore, they agree with results from Liao et al. (2013) where known ages were compared with ages from both otoliths and scales and otoliths were found to have much smaller error than scales.

#### **7.5.2 Objective 2**

The results from this analysis agree with the Bayesian GLMM analysis demonstrating significant differences in ages from otoliths compared to ages from scales. In both analyses, the precision between otolith readers was much higher than for scale readers indicating scale ages have higher uncertainty associated with them. Moreover, the results from this analysis also agree with prior research where scale ages from older fish tend to be underestimated and younger fish tend to be overestimated (Secor et al. 1995; Liao et al. 2009; Liao et al. 2013). The reason for the overestimation of younger ages is likely due to false annuli being mistaken for true annuli and



older ages are underestimated due to the slenderness of tightly packed annuli on the periphery of the scales (Secor et al. 1995; Liao et al. 2013).

## **7.6 Conclusion**

Estimating striped bass age with scales is a common practice and the preferred method for anadromous striped bass on the Atlantic Coast (ASMFC 2003). Scales are relatively easy to collect in the field and striped bass may be released alive after structure collection. In addition, scales may be collected with negligible effect on striped bass intended for market. However, Liao et al. (2013) found that scales overestimated ages of young fish and underestimated ages of old fish. Studies by Welch et al. (1993) and Secor et al. (1995) also indicate scales tend to underestimate the actual age of older fish beginning at age 10 when compared to otoliths. Biases in age estimates impact catch-at-age data and estimates of recruitment, natural mortality and growth.

Unlike most studies, due to PBT analysis, known-age samples from striped bass are now available and allow for validation of scale and otolith ages. A comparison of scale and otolith ages collected from striped bass in the Chesapeake Bay found otoliths provided more accurate and precise estimates of ages than scales when compared to known age fish (Liao et al. 2013). The current study similarly shows otoliths provide a more precise and accurate age estimate for CSMA striped bass when compared to scales.

This research has important implications regarding the use of ages from scales and otoliths for the management of striped bass populations. Age bias and imprecision can have significant effects on estimates of growth parameters used in stock assessment modeling (either estimated outside of the assessment model or within an integrated assessment model). Previous research has demonstrated that population dynamics estimates and biological reference points used for management are sensitive to the misspecification (bias) in growth parameter estimates (Zhu et al. 2016). Liao et al. (2013) demonstrated that age bias can adversely affect catch-at-age models by reducing the ability to track the progression of year classes caused by incorrectly assigning fish to appropriate cohorts resulting in strong recruitment events appearing weaker thus resulting in a subsequent reduction in recruitment variability. Liao et al. (2013) suggest the inability to track recruitment signals would prolong recovery of a depleted stock and result in an unnecessarily restricted harvest after recovery had occurred.

## **7.7 Recommendation**

The NCDMF recommends that otoliths should be used by both agencies to age CSMA striped bass if PBT ages are not available, and a power analysis should be conducted to determine sample sizes needed for determining the representative age structure. Another recommendation is conduct a similar study across NCDMF and NCWRC biologists to determine and compare the accuracy and precision of scale ageing versus otolith ageing for the Albemarle Sound and Roanoke River striped bass management areas.

## **8 RESEARCH RECOMMENDATIONS**

The research recommendations listed below (in no particular order) are intended to improve future assessments of the CSMA striped bass stocks. The bulleted items outline the specific issue and are organized by priority ranking.

### High

- Acquire life history information: maturity, fecundity, size and weight at age, egg and larval survival (ongoing through CRFL funded projects and NCDMF P930 data collection; see Knight, 2015, for recent work on maturation and fecundity in the Neuse and Tar-Pamlico rivers)
- Conduct delayed mortality studies for recreational and commercial gear during all seasons factoring in relationships between salinity, dissolved oxygen, and water temperature
- Develop better estimates of life-history parameters, especially growth and factors influencing rates of natural mortality for all striped bass life stages (growth is ongoing through NCDMF P930 data collection; for natural mortality, see recent publications Bradley 2016 and Bradley et al. 2018b)

### Medium

- Determine factors impacting survivability of stocked fish in each system (Bradley et al. 2018b)
- Implement a random component to NCDMF program 100 juvenile sampling in the CSMA
- Conduct a power analysis to determine minimum sample sizes needed for determining the representative age structure

### Low

- Determine if contaminants are present in striped bass habitats and identify those that are potentially detrimental to various life history stages (ongoing through N.C. Division of Water Quality but could be expanded; in 2017, NCSU was awarded a CRFL grant to conduct research on striped bass eggs, including evaluating for Gen X)
- Identify minimum flow requirements in the Tar-Pamlico, Neuse, and Cape Fear rivers necessary for successful spawning, egg development, and larval transport to nursery grounds
- Evaluate factors influencing catchability of striped bass, particularly larger striped bass, in electrofishing surveys conducted on the spawning grounds
- Obtain improved commercial discard estimates from the estuarine gill-net fisheries (i.e., anchored, runaround, and strike gill nets) in the CSMA systems to better characterize harvest and discards
- Investigate factors influencing mixing rates between A-R and CSMA striped bass stocks
- Identify water quality parameters that impact spawning, hatching, and survival of striped bass in CSMA systems
- Develop a consistent ageing approach across agency sampling programs
- Continue PIT tagging striped bass in the Cape Fear River and expand PIT tagging to the Tar-Pamlico and Neuse rivers to estimates of spawning population size
- Investigate factors influencing rates of natural mortality for all striped bass life stages in the CSMA systems

## 9 LITERATURE CITED

- Albrecht, A.B. 1964. Some observations on factors associated with survival of striped bass eggs and larvae. *California Fish and Game* 50(2):100–113.
- Ashley, K.W., and R.T. Rachels. 2006. Cape Fear River striped bass spawning stock survey, 2006. Project F-22, Federal Aid in Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Atlantic States Marine Fisheries Commission (ASMFC). 2003. Amendment # 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 41, Washington, D.C.
- ASMFC. 2007. Addendum 1 to Amendment 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 16, Washington, D.C.
- Bacheler, N.M., J.A. Buckel, J.E. Hightower, L.M. Paramore, and K.H. Pollock. 2009. A combined telemetry–tag return approach to estimate fishing and natural mortality rates of an estuarine fish. *Canadian Journal of Fisheries and Aquatic Sciences* 66:1230-1244.
- Baker, W. 1968. A reconnaissance of anadromous fish runs into the inland fishing water of North Carolina. Final Report, Project AFS-3, Federal Aid in Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Bain, M.B., and J.L. Bain. 1982. Habitat suitability index models: coastal stocks of striped bass. U.S. Fish and Wildlife Service, Office of Biological Services, Report No. FWS/OBS-82/10.1, Washington, D.C.
- Barwick R.D., K. Rundle, J. Homan, W. Collart, and C. Thomas. 2008. An assessment of striped bass stocks and contribution by stocked juveniles in two North Carolina rivers (1998–2008). Final Report, Project F-22, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Bason, W.H. 1971. Ecology and early life history of striped bass, *Morone saxatilis*, in the Delaware Estuary. *Ichthyology Association Bulletin* 4. 122 p.
- Bayless, J.D., and W.B. Smith. 1962. Survey and classification of the Neuse River and tributaries, North Carolina. Final Report, Federal Aid in Fish Restoration Job IA, Project F-14-R. North Carolina Wildlife Resources Commission, Raleigh.
- Beasley, C.A., and J.E. Hightower. 2000. Effects of a low-head dam on the distribution and characteristics of spawning habitat used by striped bass and American shad. *Transactions of the American Fisheries Society* 129:1316–1330.
- Bergey, L.L., R.A. Rulifson, M.L. Gallagher, and A.S. Overton. 2003. Variability of Atlantic coast striped bass egg characteristics. *North American Journal of Fisheries Management* 23:558–572.
- Berst, A.H. 1961. Selectivity and efficiency of experimental gill nets in South Bay and Georgian Bay of Lake Huron. *Transactions of the American Fisheries Society* 90:423–418.
- Bigelow, H.B., and W.C. Schroeder. 1953. Fishes of the Gulf of Maine. U.S. Fish and Wildlife Service Fisheries Bulletin 53.

- Binion-Rock, S.M. 2018. Trophic dynamics and ecosystem modeling of finfishes in Pamlico Sound, North Carolina. Doctoral Dissertation. North Carolina State University, Raleigh. 301 p.
- Blanchard, J.L., D.L. Maxwell, and S. Jennings. 2008. Power of monitoring surveys to detect abundance trends in depleted fish populations: the effects of density-dependent habitat use, patchiness, and climate change. *ICES Journal of Marine Science* 65:111–120.
- Boyd, J.B. 2011. Maturation, fecundity, and spawning frequency of the Albemarle/Roanoke striped bass stock. Master's thesis. East Carolina University, Greenville, North Carolina. 132 p.
- Bradley, C. 2016. Evaluation of juvenile and adult striped bass mortality and distribution, with implications for recovery efforts in the Neuse River, North Carolina. Master's thesis. North Carolina State University, Raleigh.
- Bradley, C.E., J.A. Rice, and D.D. Aday. 2018a. Modeling the effects of vital rate manipulation and management scenarios to predict the population impact of restoration programs on an unrecovered coastal population of striped bass. *North American Journal of Fisheries Management* 38:639–649.
- Bradley, C.E., J.A. Rice, D.D. Aday, J.E. Hightower, J. Rock, and K.L. Lincoln. 2018b. Juvenile and adult striped bass mortality and distribution in an unrecovered coastal population. *North American Journal of Fisheries Management* 38:104–119.
- Buckley, C., K. Lincoln, K. Rachels, and B. Ricks. 2019. Striped bass ichthyoplankton surveys in the Neuse River, 2016–2017. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Buckmeier, D.L., and J.W. Schlechte. 2009. Capture efficiency and size selectivity of channel catfish and blue catfish sampling gears. *North American Journal of Fisheries Management* 29(2):404–416.
- Burdick, M., and J.E. Hightower. 2006. Distribution of spawning activity by anadromous fishes in an Atlantic slope drainage after removal of a low-head dam. *Transactions of the American Fisheries Society* 135:1290–1300.
- Burnham, K., and D. Anderson. 2002. Model selection and multimodel inference: a practical information-theoretic approach. Springer, New York.
- Callihan, J. 2012. Summary maps of North Carolina tagging programs. NCDMF Marine Fisheries Fellowship, 2011–2012. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Callihan, J.L., C.H. Godwin, and J.A. Buckel. 2014. Effect of demography on spatial distribution: movement patterns of Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*) in relation to their stock recovery. *Fisheries Bulletin* 112(2-3):131–143.
- Caswell, H., 2001. Matrix population models. Sinauer Associates, Sunderland.
- Clark, W.G. 1999. Effects of an erroneous natural mortality rate on a simple age-structured stock assessment. *Canadian Journal of Fisheries and Aquatic Sciences* 56:1721–1731.

- Collier, C., W. Smith, C.B. Stewart, and S. Taylor. 2013. Mark recapture study to determine population size of Cape Fear River striped bass. Final Report, Coastal Recreational Fishing License Grant 2F25. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 34 p.
- Cowan, J.H., K.A. Rose, E.S. Rutherford, and E.D. Houde. 1993. Individual-based model of young-of-the-year striped bass population dynamics. II. Factors affecting recruitment in the Potomac River, Maryland. *Transactions of the American Fisheries Society* 122(3):439–458.
- Dahlberg, M.D., 1979. A review of survival rates of fish eggs and larvae in relation to impact assessments. *Marine Fisheries Review* 41:1–12.
- Darsee, S.P., T. Mathes and J. Facendola. 2019. North Carolina Striped Bass monitoring. Federal Aid in Sport Fish Restoration, Project F-56 Segment 26. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 61 p.
- Daugherty, D.J., and T.M. Sutton. 2005. Use of a chase boat for increasing electrofishing efficiency for flathead catfish in lotic systems. *North American Journal of Fisheries Management* 25(4):1528–1532.
- Denson, M.R., K. Brenkert, W.E. Jenkins, and T.L. Darden. 2012. Assessing red drum juvenile stocking in a South Carolina estuary using genetic identification. *North American Journal of Fisheries Management* 32:32–43.
- Dial Cordy and Associates, Inc. 2006. Cape Fear River anadromous fish egg and larval survey. Draft report to U.S. Army Corps of Engineers, Contract W912HN-05-D-0014, Wilmington, North Carolina.
- Dial Cordy and Associates, Inc. 2017. Restoring access to historic migratory fish habitat in the Cape Fear River Basin. Draft Report. Prepared for National Fish and Wildlife Foundation. Wilmington, North Carolina.
- Dolan, C.R., and L.E. Miranda. 2003. Immobilization thresholds of electrofishing relative to fish size. *Transactions of the American Fisheries Society* 132(5):969–976.
- Doroshev, S.I. 1970. Biological features of the eggs, larvae, and young of the striped bass [*Roccus saxatilis* (Walbaum)] in connection with the problem of its acclimatization in the USSR. *Journal of Ichthyology* 10(2):235–248.
- Ellis, T.A., J.E. Hightower, and J.A. Buckel. 2018. Relative importance of fishing and natural mortality for spotted seatrout (*Cynoscion nebulosus*) estimated from a tag-return model and corroborated with survey data. *Fisheries Research* 199:81–93.
- Farrae, D. 2019. 2018 Striped bass genotyping and parentage analysis. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Fisk, J.M., and C.W. Morgeson. 2016. Cape Fear River striped bass survey—2015. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.

- Gabriel, W.L., M.P. Sissenwine, and W.J. Overholtz. 1989. Analysis of spawning stock biomass per recruit: an example for Georges Bank haddock. *North American Journal of Fisheries Management* 9(4):383–391.
- Gelman, A., and J. Hill. 2007. *Data analysis using regression and multilevel/hierarchical models*. Cambridge, New York.
- Gibbons, J.W., and K.M. Andrews. 2004. PIT tagging: simple technology at its best. *Bio-Science* 54:447–454.
- Goodyear, C.P. 1993. Spawning stock biomass per recruit in fisheries management: foundation and current use. Pages 67–81 *In*: S.J. Smith, J.J. Hunt, and D. Rivard (editors), *Risk evaluation and biological reference points for fisheries management*. Canadian Special Publication of Fisheries and Aquatic Sciences 120.
- Greene, K.E., J.L. Zimmerman, R.W. Laney, and J.C. Thomas-Blate. 2009. Atlantic coast diadromous fish habitat: a review of utilization, threats, recommendations for conservation, and research needs. Atlantic States Marine Fisheries Commission, Habitat Management Series No. 9, Washington D.C. 464 p.
- Hadley, J. 2015. Survey of commercial fishing license holders for personal consumption of seafood caught with commercial gear. Final Report. North Carolina Department of Environmental Quality, Division of Marine Fisheries, Morehead City, North Carolina. 13 p.
- Hansson, S., and L.G. Rudstam. 1995. Gillnet catches as an estimate of fish abundance: a comparison between vertical gillnet catches and hydroacoustic abundance of Baltic Sea herring (*Clupea harengus*) and sprat (*Sprattus sprattus*). *Canadian Journal of Fisheries and Aquatic Sciences* 52:75–83.
- Hardy, J.D., Jr. 1978. Development of fishes of the mid-Atlantic Bight: An atlas of egg, larval and juvenile stages, volume III, aphredoderidae through rachycentridae. U.S. Department of the Interior, Fish and Wildlife Service, Biological Services Program Report No. FWS/OBS-78/12:1–394, Washington, D.C.
- Harris, J.E., and J.E. Hightower. 2015. Estimating mortality rates for Albemarle Sound-Roanoke River striped bass using an integrated modeling approach. Final Report, NCDMF Reference Number 3209.
- Hassler, W.W., and S.D. Taylor. 1984. The status, abundance, and exploitation of striped bass in the Roanoke River and Albemarle Sound, North Carolina, 1982, 1983. Completion Report, Project AFC-19. NCDMF Publication No. 136. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Hawkins, J.H. 1979. Anadromous fisheries research program—Neuse River. Progress report, Project AFCS 13-2. North Carolina Department of Natural Resources and Community Development. Division of Marine Fisheries, Morehead City, North Carolina. 101 p.
- Hawkins, J.H. 1980. Investigations of anadromous fishes of the Neuse River, North Carolina. North Carolina Department of Natural Resources and Community Development, Division of Marine Fisheries, Special Scientific Report 34, Morehead City, North Carolina.

- Hilborn, R., E.K. Pikitch, and M.K. McAllister. 1994. A Bayesian estimation and decision analysis for an age-structured model using biomass survey data. *Fisheries Research* 19(1-2):17–30.
- Hill, J., J.W. Evans, and M.J. Van Den Avyle. 1989. Species profiles: life histories and environmental requirements of coastal fishes and invertebrates (South Atlantic)—striped bass. Biological Report 82(11.118), U.S. Department of the Interior, Fish and Wildlife Service, Washington, D.C. 35 p.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. *Canadian Journal of Fisheries and Aquatic Sciences*, 52(2):364–368.
- Hoff, P.D. 2009. A first course in Bayesian statistical methods. Springer, New York.
- Homan, J.M., K.R. Rundle, R.D. Barwick, and K.W. Ashley. 2010. Review of striped bass monitoring programs in the Central/Southern Management Area, North Carolina—2009. Final Report, Project F-22, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Humphries, E.T. 1965. Spawning grounds of the striped bass, *Morone saxatilis* (Walbaum), in the Tar River, North Carolina. Master's thesis. East Carolina University, Greenville, North Carolina. 50 p.
- Humphries, M., and J.W. Kornegay. 1985. An evaluation of the use of bony structures for aging Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*). Project F-22, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Jiang, H., K.H. Pollock, C. Brownie, J.M. Hoenig, R.J. Latour, B.K. Wells, and J.E. Hightower. 2007. Tag return models allowing for harvest and catch and release: evidence of environmental and management impacts on striped bass fishing and natural mortality rates. *North American Journal of Fisheries Management* 27:387–396.
- Jiao, Y., N.W.R. Lapointe, P.L. Angermeier, and B.R. Murphy. 2009. Hierarchical demographic approaches for assessing invasion dynamics of non-indigenous species: An example using northern snakehead (*Channa argus*). *Ecological Modelling* 220:1681–1689.
- Jones, T.W., and W.J. Collart. 1997. Assessment of Tar River Striped Bass egg production, age, growth and sex composition, 1996. Final Report, Federal Aid Project F-23. North Carolina Wildlife Resources Commission, Raleigh, North Carolina. 10 p.
- Kimura, D.K. 2008. Extending the von Bertalanffy growth model using explanatory variables. *Canadian Journal of Fisheries and Aquatic Sciences* 65:1879–1891.
- Knight, E.H. 2015. Maturation and fecundity of the Neuse River and Tar-Pamlico rivers striped bass populations (*Morone saxatilis*) stocks in 2013. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Kornegay, J., and E.T. Humphries. 1975. Spawning of the striped bass in the Tar River, North Carolina. *Proceedings of the Annual Conference of the Southeastern Association of Game and Fish Commissioners* 29:317–325.

- Lee, L.M., and J.E. Rock. 2018. The forgotten need for spatial persistence in catch data from fixed station surveys. *Fishery Bulletin* 116(1):69–74.
- Lewis, R.M., and R.R. Bonner, Jr. 1966. Fecundity of the striped bass, *Roccus saxatilis* (Walbaum). *Transactions of the American Fisheries Society* 95(3):328–331.
- Li, Y., and Y. Jiao. 2015. Evaluation of stocking strategies for endangered white abalone using a hierarchical demographic model. *Ecological Modelling* 299:14–22.
- Li, Y., Y. Jiao, and J.A. Browder. 2016. Assessment of seabird bycatch in the US Atlantic pelagic longline fishery, with an extra exploration on modeling spatial variation. *ICES Journal of Marine Science* 73:2687–2694.
- Liao, H., C. Jones, C. Morgan, and J. Ballenger. 2009. Finfish ageing for Virginia catches and application of Virtual Population Analysis to provide management advice. Final Report 2008. VMRC/ODU, Center for Quantitative Fisheries Ecology. 149 p.
- Liao, H., A.F. Sharov, C.M. Jones, and G.A. Nelson. 2013. Quantifying the effects of aging bias in Atlantic striped bass stock assessment. *Transactions of the American Fisheries Society* 142(1):193–207.
- Limburg, K.E., and J.R. Waldman. 2009. Dramatic declines in north Atlantic diadromous fishes. *BioScience* 59(11):955–965.
- Lorenzen, K. 1996. The relationship between body weight and natural mortality in juvenile and adult fish: a comparison of natural ecosystems and aquaculture. *Journal of Fish Biology* 49(4):627–647.
- Lorenzen, K. 2000. Allometry of natural mortality as a basis for assessing optimal release size in fish-stocking programmes. *Canadian Journal of Fisheries and Aquatic Sciences* 57:2374–2381.
- Lorenzen, K. 2005. Population dynamics and potential of fisheries stock enhancement: practical theory for assessment and policy analysis. *Philosophical Transactions of the Royal Society of London, Series B* 360(1453):171–189.
- Lupton, B.Y., and P.S. Phalen. 1996. Designing and implementing a trip ticket program. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 32 p + appendices.
- Manooch, C.S. 1973. Food habits of yearling and adult striped bass, *Morone saxatilis* (Walbaum) from Albemarle Sound, North Carolina. *Chesapeake Science* 14(2):73–86.
- Manooch, C.S., and R.A. Rulifson. 1989. Roanoke River Water Flow Committee Report: a recommended water flow regime for the Roanoke River, North Carolina, to benefit anadromous striped bass and other below-dam resources and users. NOAA Technical Memorandum NMFS-SEFC-216.
- Mansueti, R.J. 1958. Eggs, larvae and young of the striped bass, *Roccus saxatilis*. Chesapeake Biological Laboratory Contribution No. 112. Maryland Department of Resources and Education, Solomons, Maryland.
- Markle, D.F., and G.C. Grant. 1970. The summer food habits of young-of-the-year striped bass in three Virginia rivers. *Chesapeake Science* 11(1):50–54.



- Martino, E.J., and E.D. Houde. 2010. Recruitment of striped bass in Chesapeake Bay: spatial and temporal environmental variability and availability of zooplankton prey. *Marine Ecology Progress Series* 409:213–228.
- Marvin, D.P. 2012. The success of Columbia Basin passive integrated transponder (PIT) tag information system. *American Fisheries Society Symposium* 76:95–134.
- McInerny, M.C., and T.K. Cross. 2000. Effects of sampling time, intraspecific density, and environmental variables on electrofishing catch per effort of largemouth bass in Minnesota lakes. *North American Journal of Fisheries Management* 20(2):328–336.
- Midway, S.R., T. Wagner, S.A. Arnott, P. Biondo, F. Martinez-Andrade, and T.F. Wadsworth. 2015. Spatial and temporal variability in growth of southern flounder (*Paralichthys lethostigma*). *Fisheries Research* 167:323–332.
- Minami, M., C.E. Lennert-Cody, W. Gao, and M. Román-Verdesoto. 2007. Modeling shark bycatch: the zero-inflated negative binomial regression model with smoothing. *Fisheries Research* 84(2):210–221.
- Monteleone, D.M., and E.D. Houde. 1990. Influence of maternal size on survival and growth of striped bass *Morone saxatilis* Walbaum eggs and larvae. *Journal of Experimental Marine Biology and Ecology* 140(1-2):1–11.
- Morgan, R.P., II, and V.J. Rasin, Jr. 1973. Effects of salinity and temperature on the development of eggs and larvae of striped bass and white perch: Appendix X *In*: Hydrographic and ecological effects of enlargement of the Chesapeake and Delaware Canal. Final Report No. DACW-61-71-C-0062. U.S. Army Corps of Engineers Philadelphia, Pennsylvania.
- Morgeson, C.W., and J.M. Fisk. 2018. Cape Fear River anadromous spawning activity survey—2016. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Morris, W.F., and D.F. Doak. 2004. Buffering of life histories against environmental stochasticity: accounting for a spurious correlation between the variabilities of vital rates and their contributions to fitness. *The American Naturalist* 163:579–590.
- Nichols, P.R., and D.E. Louder. 1970. Upstream passage of anadromous fish through navigation locks and use of the stream for nursery and spawning habitat, Cape Fear River, North Carolina, 1962–1966. U.S. Fish and Wildlife Service Circular 352.
- North Carolina Department of Environmental Quality (NCDEQ). 2016. North Carolina coastal habitat protection plan. North Carolina Department of Environmental Quality, Raleigh, North Carolina.
- North Carolina Division of Marine Fisheries (NCDMF). 2004. North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environment and Natural Resources, Division of Marine Fisheries, Morehead City, North Carolina. 374 p.
- NCDMF. 2014. November 2014 Revision to Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environmental and Natural Resources, Division of Marine Fisheries, Elizabeth City, North Carolina. 15 p.

- NCDMF. 2017. North Carolina cooperative striped bass creel survey in the Central and Southern Management Area (CSMA). Final Report, Grant F-79. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- NCDMF. 2019a. Supplement A to Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environment and Natural Resources, North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 826 p.
- NCDMF. 2019b. North Carolina Division of Marine Fisheries License and Statistics Section Annual Report. North Carolina Department of Environmental Quality, Division of Marine Fisheries, Morehead City, North Carolina.
- NCDMF. 2020. North Carolina Division of Marine Fisheries 2019 Annual Fisheries Management Plan Review. North Carolina Department of Environmental Quality, Division of Marine Fisheries, Morehead City, North Carolina.
- NCDMF and North Carolina Wildlife Resources Commission (NCWRC). 2013. Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environmental and Natural Resources, Division of Marine Fisheries, Elizabeth City, North Carolina. 429 p.
- NCWRC and NCDMF. 2011. Estuarine striped bass in North Carolina: scale ageing methods. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Nelson, K.L., and A.E. Little. 1991. Early life history characteristics of Striped Bass in the Tar and Neuse rivers, North Carolina. Final Report, Project F-22-11, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Northeast Fisheries Science Center (NEFSC). 2013. 57th Northeast Regional Stock Assessment Workshop (57th SAW) Assessment Summary Report. U.S. Department of Commerce, Northeast Fisheries Science Center Reference Document 13-14. 39 p.
- O'Donnell T., and D. Farrae. 2017. South Carolina Department of Natural Resources. Final Report, North Carolina Wildlife Resources Commission 2016 Striped Bass Genotyping and Parentage Analyses 2016.
- Olsen, J.E., and R. Rulifson. 1992. Maturation and fecundity of Roanoke River-Albemarle Sound striped bass. *Transactions of the American Fisheries Society* 121:524–537.
- Plummer, M. 2003. JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. *Proceedings of the 3rd International Workshop on Distributed Statistical Computing* 124(125.10):1–10.
- Pollock, K.H., C.M. Jones, and T.L. Brown. 1994. Angler survey methods and their applications in fisheries management. *American Fisheries Society Special Publication* 25.
- Prescott, J.C. 2019. Migration ecology of striped bass (*Morone saxatilis*) in the Cape Fear River, North Carolina. Master's Thesis. University of North Carolina Wilmington, Wilmington, North Carolina.

- Quinn, T.J., and R.B. Deriso. 1999. Quantitative fish dynamics. Oxford University Press, New York.
- R Core Team. 2013. R: A language an environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. [Available at <https://www.R-project.org/>].
- Raabe, J.K., J.E. Hightower, T.A. Ellis, and J.J. Facendola. 2019. Evaluation of fish passage at a nature-like rock ramp fishway on a large coastal river. *Transactions of the American Fisheries Society* 148:798–816.
- Rachels, K.T., and B.R. Ricks. 2015. Neuse River striped bass monitoring programs, population dynamics, and recovery strategies. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Rachels, K.T., and B.R. Ricks. 2018. Exploring causal factors of spawning stock mortality in a riverine striped bass population. *Marine and Coastal Fisheries* 10:424–434.
- Reynolds, J.B. 1996. Electrofishing. Pages 221–253 *In*: B.R. Murphy and D.W. Willis (editors), *Fisheries techniques*, 2<sup>nd</sup> edition. American Fisheries Society, Bethesda, Maryland.
- Richards, R.A., and P.J. Rago. 1999. A case history of effective fishery management: Chesapeake Bay striped bass. *North American Journal of Fisheries Management* 19:356–375.
- Ricker, W.E. 1975. Computation and interpretation of biological statistics of fish populations. *Bulletin of the Fisheries Research Board of Canada*. 382 p.
- Ricks, B.R., and C. Buckley. 2018. Neuse River striped bass monitoring programs, 2016–2017. Final Report, Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Robins, C.R., R.M. Bailey, C.E. Bond, J.R. Brooker, E.A. Lachner, R.N. Lea, and W.B. Scott. 1991. Common and scientific names of fishes from the United States and Canada, Fifth Edition. American Fisheries Society Special Publication 20. Bethesda, Maryland.
- Rock, J., D. Zapf, C. Wilson, and D. Mumford. 2016. Improving estimates of striped bass discards in the Central Southern Management Area (CSMA) through a recreation access site survey and an expanded observer program. Final Report, CRFL Grant 2011-F-001. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 76 p.
- Rock, J., D. Zapf, J. Facendola, and C. Stewart. 2018. Assessing critical habitat, movement patterns, and spawning grounds of anadromous fishes in the Tar-Pamlico, Neuse, and Cape Fear rivers using telemetry tagging techniques. Final Report, CRFL Grant 2013-F-103. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 109 p.
- Rogers, B.A., D.T. Westin, and S.B. Saila. 1977. Life stage duration studies on Hudson River striped bass. NOAA Sea Grant Marine Technical Report 31. University of Rhode Island, Applied Marine Research Group, Kingston, Rhode Island.

- Ruetz III, C.R., D.G. Uzarski, D.M. Krueger, and E.S. Rutherford. 2007. Sampling a littoral fish assemblage: comparison of small-mesh fyke netting and boat electrofishing. *North American Journal of Fisheries Management* 27(3):825–831.
- Rulifson, R.A. 1990. Abundance and viability of striped bass eggs spawned in the Roanoke River, North Carolina, in 1989. North Carolina Project No. 90-11, Albemarle-Pamlico Estuarine Study. North Carolina Department of Environmental Management, Health and Natural Resources and U.S. Environmental Protection Agency, Raleigh, North Carolina. 96 p.
- Rulifson, R.A. 1991. Comparing the abundance and viability of striped bass eggs spawned in the Roanoke River, North Carolina, at two locations in 1991. Interim report to the North Carolina Striped Bass Study Management Board. Institute for Coastal and Marine Resources, and Department of Biology, East Carolina University, Greenville, NC.
- Rulifson, R.A. 2014. Origin of Central Southern Management Area striped bass using otolith chemistry, and recommendations for fishery management. Final Report, Coastal Recreational Fishing License Grant 2011-F-005. 67 p.
- Rulifson, R.A., and D. Bass. 1991. Food analyses of young-of-year. Pages 217–219 *In*: NOAA Technical Memorandum NMFS-SEFC-291.
- Rulifson, R.A., M.T. Huish, and R.W.W. Thoesen. 1982a Status of anadromous fisheries in southeast U.S. estuaries. Pages 413–425 *In*: V. Kennedy (editor), *Estuarine comparisons*. Academic Press Inc., New York, NY.
- Rulifson, R.A., M.T. Huish, and R.W. Thoesen. 1982b. Anadromous fish in the Southeastern United States and recommendations for development of a management plan. U.S. Fish and Wildlife Service, Fisheries Resource, Region 4, Atlanta, Georgia. 525 p.
- Rulifson, R.A., and J.J. Isely. 1995. Striped bass egg abundance and viability in the Roanoke River, North Carolina, and young-of-year survivorship, for 1993. Completion Report to Virginia Power, Innsbrook Technical Center, Glenn Allen, Virginia. Institute for Coastal and Marine Resources, East Carolina University, Greenville, North Carolina.
- Rulifson, R.A., and W. Laney. 1999. Striped bass stocking programs in the United States: ecological and resource management issues. Research Document 99/007. Fisheries and Oceans Canada. 40 p.
- Schoenebeck, C.W., and M.J. Hansen. 2005. Electrofishing catchability of walleyes, largemouth bass, smallmouth bass, northern pike, and muskellunge in Wisconsin lakes. *North American Journal of Fisheries Management* 25(4):1341–1352.
- Seber, G.A.F. 1982. The estimation of animal abundance and related parameters. Blackburn Press, Caldwell, New Jersey.
- Secor, D.H. 2000. Longevity and resilience of Chesapeake Bay striped bass. *ICES Journal of Marine Science* 57(4):808–815.
- Secor, D.H., T.M. Trice, and H.T. Hornick. 1995 Validation of otolith-based ageing and a comparison of otolith and scale-based ageing in mark-recaptured Chesapeake Bay striped bass, *Morone saxatilis*. *Fishery Bulletin* 93(1):186–190.

- Setzler, E.M., W.R. Boynton, K.V. Wood, H.H. Zion, L. Lubbers, N.K. Mountford, P. Frere, L. Tucker, and J.A. Mihursky. 1980. Synopsis of biological data on striped bass. NOAA Technical Report, NMFS Circular 443: FAO Synopsis No. 121. 69 p.
- Shapovalov, L. 1936. Food of the striped bass. *California Fish and Game* 22:267–271.
- Sholar, T.M. 1977. Anadromous Fisheries Research Program—Cape Fear River System. Completion Report, Project AFCS-12. North Carolina Department of Natural Resources and Community Development, Division of Marine Fisheries, Morehead City, North Carolina. 81 p.
- Smith, J.A. 2009. Spawning activity and migratory characteristics of American shad and striped bass in the Cape Fear River, North Carolina. Master's thesis. North Carolina State University, Raleigh, North Carolina.
- Smith, J.A., and J.E. Hightower. 2012. Effect of low-head lock-and-dam structures on migration and spawning of American shad and striped bass in the Cape Fear River, North Carolina. *Transactions of the American Fisheries Society* 141(2):402–413.
- Smith, C.A., and K.M. Patoka. 2018. Characteristics of the 2016 and 2017 Roanoke River striped bass spawning population. Project F-108, Federal Aid in Sport Fish Restoration. North Carolina Wildlife Resources Commission, Raleigh, North Carolina.
- Smith, C.M., and R.A. Rulifson. 2015. Overlapping habitat use of multiple anadromous fish species in a restricted coastal watershed. *Transactions of the American Fisheries Society*, 144(6):1173-1183.
- Speas, D.W., C.J. Walters, D.L. Ward, and R.S. Rogers. 2004. Effects of intraspecific density and environmental variables on electrofishing catchability of brown and rainbow trout in the Colorado River. *North American Journal of Fisheries Management* 24(2):586–596.
- Stevens, D.E. 1966. Food habits of striped bass, *Morone saxatilis*, in the Sacramento-San Joaquin Delta. Pages 68–96 *In*: J.L. Turner and D.W. Kelley (compilers), *Ecological studies of the Sacramento-San Joaquin Delta. Part H. Fishes of the delta.* California Department Fish Game Fishery Bulletin 136.
- Stewart, C., and Y. Li. 2019. Mark recapture study to determine population size of Cape Fear River striped bass. Final Report, CRFL Grant 2013-F-010. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 28 p.
- Street, M.W., A.S. Deaton, W.S. Chappell, and P.D. Mooreside. 2005. North Carolina coastal habitat protection plan. North Carolina Department of Environment and Natural Resources, Division of Marine Fisheries, Morehead City, North Carolina. 656 p.
- Sullivan, C. 1956. The importance of size grouping in population estimates employing electric shockers. *Progress Fish-Culturist* 18(4):188–190.
- Thompson, W.F., and F.H. Bell. 1934. Biological statistics of the Pacific halibut fishery. 2. Effect of changes in intensity upon total yield and yield per unit of gear. Report Internal Fisheries (Pacific Halibut) Commission 8. 49 p.

- Thorson, J.T., and M.H. Prager. 2011. Better catch curves: incorporating age-specific natural mortality and logistic selectivity. *Transactions of the American Fisheries Society* 140(2):356–366.
- Trent, L., and W.W. Hassler. 1968. Gill net selection, migration, size and age composition, sex ratio, harvest efficiency, and management of striped bass in the Roanoke River, North Carolina. *Chesapeake Science* 9(4):217–232.
- Trumbo, B.A., M.D. Kaller, A.R. Harlan, T. Pasco, W.E. Kelso, and D.A. Rutherford. 2015. Effectiveness of continuous versus point electrofishing for fish assemblage assessment in shallow, turbid aquatic habitats. *North American Journal of Fisheries Management* 36:398–406.
- Vetter, E. 1988. Estimation of natural mortality in fish stocks: a review. *Fishery Bulletin* 86:25–43.
- von Bertalanffy, L. 1938. A quantitative theory of organic growth (inquiries on growth laws. II). *Human Biology* 10:181–213.
- Ware, F.J. 1971. Some early life history of Florida's inland striped bass, *Morone saxatilis*. *Procedures of the Annual Conference of the Southeast Game Fish Commission* 24:439–447.
- Warren, W.G. 1994. The potential of sampling with partial replacement for fisheries surveys. *ICES Journal of Marine Science* 51(3):315–324.
- Warren, W.G. 1995. Juvenile abundance index workshop—consultant's report. Appendix 1 *In*: P.J. Rago, C.D. Stephen, and H.M. Austin (editors), *Report of the juvenile abundances indices workshop*. Atlantic States Marine Fisheries Commission, Special Report No. 48, Washington, D.C. 83 p.
- Welch, T.J., M.J. Van Den Avyle, R.K. Betsill, and E.M. Driebe. 1993. Precision and relative accuracy of striped bass age estimates from otoliths, scales and anal fin rays and spines. *North American Journal of Fisheries Management* 13:616–620.
- Wickham, H. 2009. *ggplot2: elegant graphics for data analysis*. Springer-Verlag, New York.
- Wilke, C.O. 2019. *Ridgeline plots in 'ggplot2'*.
- Winslow, S.E. 2007. North Carolina striped bass tagging and return summary. January 1980–September 30, 2007. North Carolina Division of Marine Fisheries. Morehead City, North Carolina.
- Winslow, E., N.S. Sanderlin, G.W. Judy, J.H. Hawkins, B.F. Holland, Jr., C.A. Fischer, and R.A. Rulifson. 1983. North Carolina anadromous fisheries management program. Annual Report, Project AFCS-22. North Carolina Department of Natural Resources and Community Development, Division of Marine Fisheries, Morehead City, North Carolina. 207 p.
- Woodroffe, J.R. 2011. Historical ecology of striped bass stocking in the south-eastern United States. Master's thesis. East Carolina University, Greenville, North Carolina.
- Zar, J.H. 1999. *Biostatistical analysis*, fourth edition. Prentice Hall, New Jersey.

- Zhu, J., M.N. Maunder, A.M. Aires-da-Silva, and Y. Chen. 2016. Estimation of growth within Stock Synthesis models: management implications when using length-composition data. *Fisheries Research* 180:87–91.
- Zuur, A.F., E.N. Ieno, N.J. Walker, A.A. Saveliev, and G.M. Smith. 2009. *Mixed effects models and extensions in ecology with R*. Springer-Verlag, New York. 574 p.
- Zuur, A.F., A.A. Saveliev, and E.N. Ieno. 2012. *Zero inflated models and generalized linear mixed models with R*. Highland Statistics Ltd, United Kingdom. 324 p.

**10 TABLES**

**Table 1.1.** Stocking numbers of Phase II (5–7 inches total length) striped bass by system and year for the Tar-Pamlico, Neuse, and Cape Fear rivers, 1980–2018.

<b>Year Class</b>	<b>Tar-Pamlico River</b>	<b>Neuse River</b>	<b>Cape Fear River</b>
1980			14,874
1981		47,648	
1982	76,674		
1983			
1984	26,000		56,437
1985		39,769	
1986			
1987	17,993		
1988		71,092	
1989			77,242
1990		61,877	
1991	30,801		
1992		116,820	
1993	118,600		
1994	183,254	79,933	
1995	140,972		
1996		100,760	
1997	24,031		
1998		83,195	
1999	17,954		
2000		108,000	
2001	37,000		
2002		147,654	
2003	159,996		
2004		168,011	172,055
2005	267,376		
2006		99,595	102,283
2007	69,871	69,953	
2008	91,962		92,580
2009	61,054	104,061	112,674



**Table 1.1. (continued)** Stocking numbers of Phase II (5–7 inches total length) striped bass by system and year for the Tar-Pamlico, Neuse, and Cape Fear rivers, 1980–2018.

<b>Year Class</b>	<b>Tar-Pamlico River</b>	<b>Neuse River</b>	<b>Cape Fear River</b>
<b>2010<sup>1</sup></b>	114,012	107,142	210,105
<b>2011</b>	107,767	102,089	130,665
<b>2012<sup>2</sup></b>	45,667	90,178	127,078
<b>2013</b>	123,416	113,834	195,882
<b>2014</b>	92,727	78,899	141,752
<b>2015</b>	52,922	109,146	116,011
<b>2016</b>	121,190	134,559	63,914
<b>2017</b>	101,987	14,203 <sup>3</sup>	154,024
<b>2018</b>	186,609	149,076	152,593

---

<sup>1</sup> First year of *in situ* broodstock collection from the Cape Fear River

<sup>2</sup> First year of *in situ* broodstock collection from the Tar-Pamlico and Neuse rivers

<sup>3</sup> Poor spawning of broodstock led to low stocking numbers

**Table 1.2.** Percent hatchery contribution from striped bass genetic samples collected in the Tar-Pamlico, Neuse, and Cape Fear rivers by NCDMF and NCWRC staff, 2013–2018. (Source: South Carolina Department of Natural Resources)

<b>Year</b>	<b>Agency</b>	<b>System</b>	<b>n Total</b>	<b>n Hatchery</b>	<b>n Unknown</b>	<b>% Hatchery</b>	<b>% Unknown</b>
<b>2013</b>	NCWRC	Upper Tar-Pamlico	196	162	34	83	17
		Upper Neuse	195	130	65	67	33
		Cape Fear	219	138	81	63	37
<b>2014</b>	NCWRC	Upper Tar-Pamlico	205	174	31	85	15
		Upper Neuse	299	247	52	83	17
		Cape Fear	292	223	71	76	24
<b>2015</b>	NCWRC	Upper Tar-Pamlico	208	194	14	93	7
		Upper Neuse	241	176	65	73	27
		Cape Fear	233	166	67	71	29
<b>2016</b>	NCDMF	Tar-Pamlico	190	164	26	86	14
		Neuse	150	142	8	95	5
		Cape Fear					
	NCWRC	Upper Tar-Pamlico	195	171	24	88	12
		Upper Neuse	61	44	17	72	28
		Cape Fear	213	196	17	92	8
<b>2017</b>	NCDMF	Tar-Pamlico	147	102	45	70	31
		Neuse	118	66	52	56	44
		Cape Fear	110	93	17	85	15
	NCWRC	Upper Tar-Pamlico	137	96	41	70	30
		Upper Neuse	233	198	35	85	15
		Cape Fear	119	106	13	89	11
<b>2018</b>	NCDMF	Tar-Pamlico	206	74	132	36	64
		Neuse	86	46	40	54	47
		Cape Fear	96	81	15	84	16
	NCWRC	Upper Tar-Pamlico	166	67	99	41	59
		Upper Neuse	322	250	72	78	12
		Cape Fear	119	110	9	93	7

**Table 2.1.** Summary (mean, minimum, maximum and number of samples) striped bass length data (TL in inches) from CSMA commercial harvest, 2000–2018.

Year	Tar-Pamlico R. / Pungo R.				Neuse / Bay R.			
	Mean	Min	Max	n	Mean	Min	Max	n
2000	23	20	35	126	25	22	31	5
2001	23	21	26	116	25	23	31	12
2002	24	19	39	96	25	19	29	31
2003	23	18	37	173	24	19	37	19
2004	24	20	42	131	25	19	37	74
2005	23	20	37	127	24	20	36	70
2006	22	18	37	119	24	19	36	144
2007	22	19	33	112	22	19	27	63
2008	22	18	43	84	23	19	44	39
2009	22	19	31	99	22	18	31	85
2010	22	19	26	194	23	19	32	263
2011	23	18	27	284	23	19	42	195
2012	24	15	30	254	24	19	29	96
2013	25	18	40	225	25	18	39	301
2014	22	18	39	52	24	20	38	56
2015	24	19	40	97	24	19	44	97
2016	24	17	29	257	23	19	28	78
2017	24	19	31	151	24	19	50	97
2018	23	19	32	76	24	18	38	163

**Table 2.2.** Commercial estimates of striped bass discards (standard error in parentheses) in the Tar-Pamlico/Pungo rivers by mesh size, 2013–2018.

Year	Live Releases			Dead			Release Mortalities		Total Dead	
	Small Mesh	Large Mesh	Combined	Small Mesh	Large Mesh	Combined	Small Mesh	Large Mesh	Small Mesh	Large Mesh
2013	484 (123)	490 (150)	975 (244)	59 (13)	230 (73)	289 (85)	208	211	267	442
2014	258 (83)	490 (133)	749 (143)	33 (11)	233 (80)	266 (91)	112	212	145	445
2015	149 (46)	145 (51)	296 (87)	41 (15)	184 (75)	224 (90)	65	63	106	246
2016	421 (97)	470 (171)	891 (242)	30 (11)	131 (36)	161 (46)	181	203	210	333
2017	269 (104)	143 (64)	411 (159)	37 (13)	93 (38)	130 (51)	115	60	152	154
2018	416 (214)	346 (145)	762 (344)	25 (7)	86 (30)	111 (36)	179	148	204	234

**Table 2.3.** Commercial estimates of striped bass discards (standard error in parentheses) in the Neuse/Bay rivers by mesh size, 2013–2018.

Year	Live Releases			Dead			Release Mortalities		Total Dead	
	Small Mesh	Large Mesh	Combined	Small Mesh	Large Mesh	Combined	Small Mesh	Large Mesh	Small Mesh	Large Mesh
2013	110 (32)	132 (45)	243 (69)	34 (8)	204 (53)	237 (61)	47	58	81	261
2014	182 (61)	74 (22)	256 (76)	54 (20)	108 (35)	162 (54)	78	32	133	139
2015	56 (20)	14 (6)	71 (25)	45 (17)	68 (27)	112 (43)	23	7	68	74
2016	57 (14)	91 (36)	149 (47)	10 (3)	88 (25)	98 (28)	25	39	36	127
2017	51 (22)	35 (17)	86 (37)	20 (7)	81 (31)	101 (38)	21	15	44	96
2018	180 (96)	117 (48)	297 (138)	29 (8)	96 (29)	124 (37)	78	51	107	145

**Table 2.4.** Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries.

Zone	Year	Recreational Fishing Effort				Recreational Harvest		Striped Bass Discards				
		Total Angler Fishing Trips	Total Fishing Effort (Angler Hours)	Striped Bass Angler Trips	Striped Bass Effort (Angler Hours)	numbers	pounds	n over creel	n undersized	n legal sized	n slot sized	Total
Neuse River	2004	26,663	162,424	7,445	39,942	3,985	14,845	29	5,721	1,221	0	6,971
	2005	64,301	249,396	9,678	42,107	1,641	6,540	13	6,473	630	77	7,193
	2006	39,181	162,559	6,260	24,053	1,244	4,079		7,797	1,979	0	9,776
	2007	31,052	142,093	4,965	20,966	2,616	7,115	140	4,858	1,484	0	6,482
	2008	28,134	136,575	3,174	12,954	405	1,510	2,838	4,801	2,450	51	10,140
	2009	17,519	77,634	2,474	12,995	249	868		443	704	138	1,285
	2010	19,540	83,108	2,340	9,177	109	361		699	1,440	13	2,152
	2011	24,407	97,302	5,657	21,393	1,080	3,809		7,426	2,434	913	10,773
	2012	70,649	210,197	8,703	34,652	1,508	5,742	334	13,660	9,741	664	24,400
	2013	62,013	201,924	10,433	45,068	2,563	9,604	312	6,709	3,286	1,191	11,498
	2014	56,805	213,867	7,840	35,829	1,230	5,603	0	5,810	3,050	1,044	9,903
	2015	56,636	250,634	6,515	27,747	1,373	4,804	0	4,904	3,184	387	8,476
	2016	49,869	210,111	7,107	30,422	1,506	5,619	0	10,788	3,599	2,189	16,575
	2017	60,899	270,485	10,450	50,648	3,188	12,337	519	27,870	16,343	1,479	46,210
2018	45,237	160,827	6,076	26,228	965	3,090	17	3,459	7,296	986	11,758	
<b>Total</b>		<b>652,905</b>	<b>2,629,136</b>	<b>99,117</b>	<b>434,181</b>	<b>23,661</b>	<b>85,926</b>	<b>4,202</b>	<b>111,419</b>	<b>58,841</b>	<b>9,132</b>	<b>183,593</b>

**Table 2.4. (continued)** Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries.

Zone	Year	Recreational Fishing Effort				Recreational Harvest		Striped Bass Discards				
		Total Angler Fishing Trips	Total Fishing Effort (Angler Hours)	Striped Bass Angler Trips	Striped Bass Effort (Angler Hours)	numbers	pounds	n over creel	n undersized	n legal sized	n slot sized	Total
Tar-Pamlico River	2004	13,880	74,984	3,427	13,666	663	2,886	0	3,465	263	0	3,728
	2005	18,334	68,588	4,662	17,668	572	2,511	0	8,423	310	0	8,733
	2006	15,012	72,475	2,964	12,297	675	1,442	0	2,588	278	0	2,866
	2007	21,623	102,968	4,144	17,001	346	1,655	0	12,393	114	0	12,507
	2008	11,521	59,030	2,899	13,283	175	647	0	5,138	295	37	5,470
	2009	15,298	68,715	2,412	10,474	233	794	0	2,347	512	288	3,147
	2010	12,008	52,227	3,913	15,102	1,510	4,696	22	3,925	843	338	5,128
	2011	15,260	60,509	6,209	26,258	1,234	4,253	9	8,062	2,687	1,124	11,882
	2012	30,626	109,560	8,936	34,027	2,049	8,221	17	10,298	3,480	2,246	16,040
	2013	39,446	137,943	8,811	35,645	2,108	7,289	134	10,311	6,401	1,090	17,937
	2014	22,514	89,749	6,945	30,131	1,898	7,163	728	12,793	2,052	531	16,105
	2015	38,513	147,296	10,724	47,305	2,147	8,082	40	12,329	4,566	426	17,361
	2016	46,700	199,478	14,909	72,897	4,861	18,502	203	29,089	5,844	4,544	39,680
	2017	48,876	182,534	14,636	63,843	3,495	12,566	0	51,334	9,522	803	61,659
2018	34,648	130,200	9,274	38,548	2,046	6,403	854	22,366	4,028	904	28,151	
	<b>Total</b>	<b>384,259</b>	<b>1,556,255</b>	<b>104,865</b>	<b>448,144</b>	<b>24,011</b>	<b>87,110</b>	<b>2,008</b>	<b>194,861</b>	<b>41,195</b>	<b>12,331</b>	<b>250,395</b>
Pungo River	2004	5,532	40,573	1,910	10,183	1,493	5,227	56	2,543	259	0	2,858
	2005	7,029	34,386	2,074	9,595	1,619	5,914	139	713	76	0	928
	2006	8,470	44,599	1,387	5,716	562	1,831	33	2,163	57	0	2,253
	2007	13,089	64,273	1,862	8,688	635	2,024	7	4,422	109	0	4,538
	2008	13,232	71,210	548	2,176	263	833	0	1,782	571	3	2,356

**Table 2.4. (continued)** Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries.

Zone	Year	Recreational Fishing Effort				Recreational Harvest		Striped Bass Discards				
		Total Angler Fishing Trips	Total Fishing Effort (Angler Hours)	Striped Bass Angler Trips	Striped Bass Effort (Angler Hours)	numbers	pounds	n over creel	n undersized	n legal sized	n slot sized	Total
Pungo River	2009	13,090	67,410	756	3,142	413	1,399	7	1,681	553	292	2,533
	2010	5,970	29,308	306	1,075	138	480	7	576	118	9	710
	2011	5,579	27,996	740	3,889	414	1,412	0	1,171	276	86	1,533
	2012	9,415	50,264	700	3,285	365	1,277	88	2,385	400	0	2,873
	2013	12,665	69,902	892	5,336	796	2,644	0	2,282	669	75	3,026
	2014	7,440	44,458	459	2,192	173	602	0	582	2,002	66	2,650
	2015	5,767	32,743	711	3,644	414	1,383	0	5,038	279	0	5,317
	2016	8,806	46,520	1,268	5,670	330	1,139	0	17,997	534	46	18,578
	2017	14,534	81,889	1,013	5,031	652	2,070	31	22,582	622	11	23,246
	2018	10,785	66,683	1,019	5,080	360	1,391	0	8,304	768	0	9,072
	<b>Total</b>	<b>141,401</b>	<b>772,215</b>	<b>15,645</b>	<b>74,703</b>	<b>8,627</b>	<b>29,626</b>	<b>368</b>	<b>74,221</b>	<b>7,293</b>	<b>589</b>	<b>82,471</b>
Cape Fear River	2013	22,251	103,412	257	870	0	0	92	0	263	0	355
	2014	6,931	28,622	438	2,164	0	0	721	0	830	0	1,551
	2015	9,056	55,463	209	702	0	0	176	0	22	0	199
	2016	9,936	43,226	391	1,464	0	0	12	0	616	0	628
	2017	2,159	11,057	26	159	0	0	0	0	14	0	14
	2018	6,062	24,568	24	61	0	0	0	0	140	0	140
		<b>Total</b>	<b>50,332</b>	<b>241,780</b>	<b>1,345</b>	<b>5,419</b>	<b>0</b>	<b>0</b>	<b>1,001</b>	<b>0</b>	<b>1,885</b>	<b>0</b>

**Table 2.4. (continued)** Recreational effort, harvest, and discards estimates for striped bass in the Tar-Pamlico, Pungo, Neuse, and Cape Fear rivers and tributaries.

Zone	Year	Recreational Fishing Effort				Recreational Harvest		Striped Bass Discards				
		Total Angler Fishing Trips	Total Fishing Effort (Angler Hours)	Striped Bass Angler Trips	Striped Bass Effort (Angler Hours)	numbers	pounds	n over creel	n undersized	n legal sized	n slot sized	Total
All CSMA	2004	46,075	277,981	12,782	63,791	6,141	22,958	85	11,729	1,743	0	13,557
	2005	89,664	352,370	16,414	69,370	3,832	14,965	152	15,609	1,016	77	16,854
	2006	62,663	279,633	10,611	42,066	2,481	7,352	33	12,548	2,314	0	14,895
	2007	65,764	309,334	10,971	46,655	3,597	10,794	147	21,673	1,707	0	23,527
	2008	52,887	266,815	6,621	28,413	843	2,990	2,838	11,721	3,316	91	17,966
	2009	45,907	213,759	5,642	26,611	895	3,061	7	4,471	1,769	718	6,965
	2010	37,518	164,643	6,559	25,354	1,757	5,537	29	5,200	2,401	360	7,990
	2011	45,246	185,807	12,606	51,540	2,728	9,474	9	16,659	5,397	2,123	24,188
	2012	110,689	370,021	18,338	71,964	3,922	15,240	439	26,343	13,621	2,910	43,313
	2013	136,374	513,181	20,394	86,918	5,467	19,537	539	19,302	10,619	2,357	32,816
	2014	93,690	376,696	15,682	70,316	3,301	13,368	1,449	19,185	7,934	1,641	30,209
	2015	109,972	486,136	18,159	79,398	3,934	14,269	217	22,272	8,052	813	31,353
	2016	115,311	499,335	23,675	110,453	6,697	25,260	215	57,874	10,593	6,779	75,461
	2017	126,467	545,965	26,125	119,680	7,334	26,973	549	101,787	26,501	2,293	131,129
2018	96,732	382,278	16,393	69,917	3,371	10,884	871	34,128	12,232	1,890	49,122	
<b>Total</b>		<b>1,228,898</b>	<b>5,199,385</b>	<b>220,972</b>	<b>962,447</b>	<b>56,299</b>	<b>202,662</b>	<b>7,579</b>	<b>380,500</b>	<b>109,215</b>	<b>22,052</b>	<b>519,345</b>



**Table 2.5.** Annual weighted relative abundance index of striped bass (number of individuals per sample), total number of striped bass collected, and the number of gill net samples (n) in the Tar-Pamlico, Pungo, and Neuse rivers (2004–2018) and the Cape Fear and New rivers (2008–2018). The Percent Standard Error (PSE) represents a measure of precision of the index.

Year	Tar-Pamlico and Pungo rivers				Neuse River				Cape Fear and New rivers <sup>4</sup>			
	Index	n Striped Bass	n samples	PSE [Index]	Index	n Striped Bass	n samples	PSE [Index]	Index	n Striped Bass	n samples	PSE [Index]
2004	1.2	184	160	16	1.04	158	160	26				
2005	2.66	396	152*	14	1.37	200	152 <sup>5</sup>	23				
2006	2.38	371	160	17	1.74	268	160	17				
2007	1.57	241	160	22	1.16	177	160	19				
2008	1.61	249	160	21	1.25	193	161	23	0.04	3	84	100
2009	1.18	182	160	16	0.9	142	160	26	0.03	3	119	67
2010	2.11	329	160	17	2.02	311	160	23	0.01	1	120	100
2011	2.15	328	160	20	2.14	325	160	18	0.04	4	120	50
2012	0.94	143	160	20	0.84	127	160	20	0.03	3	120	67
2013	1.41	215	160	18	0.98	149	160	24	0.02	2	120	50
2014	1.43	217	160	16	1.82	273	160	20	0	0	120	
2015	1.14	173	160	18	1.65	251	160	18	0.14	15	120	36
2016	1.16	178	160	14	1.17	178	160	14	0.11	12	120	45
2017	1.21	186	160	17	1.41	218	160	16	0.08	9	120	50
2018	2.26	346	160	21	1.34	204	160	19	0.03	3	113	67

<sup>4</sup> Sampling in the Cape Fear and New rivers began in 2008

<sup>5</sup> In 2005, fewer stations were sampled due to high gasoline prices

**Table 2.6.** NCWRC annual catch summary for the Tar River striped bass electrofishing survey, 1996–2018.

<b>Year</b>	<b>n Sample Events</b>	<b>Total Catch</b>	<b>Males</b>	<b>Females</b>	<b>Effort</b>	<b>Mean Index</b>	<b>SD [Index]</b>	<b>SE [Index]</b>	<b>Peak Index</b>
<b>1996</b>	3	535	373	162	98,640	19.5	0.6	0.4	20.3
<b>1997</b>	3	1,275	1,045	230	103,572	44.3	13.7	7.9	53.1
<b>1998</b>	14	1,061	897	164	91,263	41.6	30.8	8.2	97.3
<b>1999</b>	8	561	334	227	50,793	36.7	28.7	10.1	93.7
<b>2000</b>	9	547	348	199	41,443	51.8	37.7	12.6	132.1
<b>2001</b>	6	326	240	86	24,814	46.8	15	6.1	68
<b>2002</b>	7	369	260	109	40,798	35.2	23.4	8.8	68.4
<b>2003</b>	6	211	169	42	23,862	31.1	13.8	5.6	51.7
<b>2004</b>	7	318	225	93	32,401	36.3	11.6	4.4	51
<b>2005</b>	13	1,429	1,390	39	63,456	86.9	53.1	14.7	184.9
<b>2006</b>	7	530	437	93	35,300	55.1	18.6	7	82.2
<b>2007</b>	8	317	264	53	41,019	29	12.2	4.3	43.5
<b>2008</b>	10	505	469	36	42,564	43.4	35.3	11.2	116.8
<b>2009</b>	6	347	265	82	23,532	48.1	33.6	13.7	98.3
<b>2010</b>	4	392	313	79	12,600	99.8	81.1	40.6	200
<b>2011</b>	4	202	100	102	18,800	37.1	19.8	9.9	54.7
<b>2012</b>	7	249	195	54	33,630	29.7	24.6	9.3	71.6
<b>2013</b>	7	315	241	74	32,400	34.2	13.5	5.1	44.7
<b>2014</b>	9	339	243	96	43,200	27	15.9	5.3	58.7
<b>2015</b>	11	418	354	64	46,800	35.1	21.9	6.6	90
<b>2016</b>	12	286	247	39	36,000	27.3	14.7	4.2	52
<b>2017</b>	5	180	128	49	23,400	25.5	9.9	4.4	34.7
<b>2018</b>	16	221	166	48	45,000	18.2	20.1	5	80

**Table 2.7.** NCWRC annual catch summary for the Neuse River striped bass electrofishing survey, 1994–2018.

<b>Year</b>	<b>n Sample Events</b>	<b>Total Catch</b>	<b>Males</b>	<b>Females</b>	<b>Effort</b>	<b>Mean Index</b>	<b>SD [Index]</b>	<b>SE [Index]</b>	<b>Peak Index</b>
1994	5	121	92	28	26,452	18.7	10.5	4.7	29.7
1995	5	125	107	18	26,381	15.3	13.2	6.6	33.2
1996	24	226	168	58	69,489	10	16.1	3.3	48
1997	26	143	114	29	76,537	6	7.1	1.4	20.8
1998	21	219	176	43	61,125	11.9	15.6	3.4	44
1999	15	292	242	50	49,562	20.4	15.6	4	62
2000	24	352	241	111	67,449	18.4	19.2	3.9	66.7
2001	22	155	132	23	57,680	8.6	11.2	2.4	46.6
2002	22	100	82	18	68,340	5.1	4.9	1	21
2003	40	401	303	98	112,305	11.8	17.6	2.8	90.5
2004	14	73	54	19	40,858	6.5	3.6	1	12.6
2005	14	65	56	9	51,094	4.5	6.6	1.8	24
2006	15	58	53	5	36,528	6.8	12.2	3.1	43.7
2007	23	170	138	32	62,372	9.8	11	2.3	51.2
2008	23	138	107	31	81,116	4.4	4.4	0.9	16.6
2009	18	360	328	31	59,094	14.1	18.1	4.3	57.3
2010	17	141	122	19	52,116	10	12.6	3.1	44.4
2011	19	176	115	60	54,129	13.9	12.4	2.8	38.4
2012	28	144	116	27	63,468	8.9	15.5	2.9	66.6
2013	29	322	265	56	71,490	15	12.4	2.3	53.7
2014	39	284	201	83	91,120	10.1	14.7	2.3	71.6
2015	42	226	198	28	47,560	15.5	27.6	4.3	137.4
2016	42	93	71	22	45,579	7.4	6.6	1	29.5
2017	61	200	155	45	81,692	6.7	11.2	1.4	55.8
2018	56	282	236	46	77,132	12.5	11.5	1.5	44.1

**Table 2.8.** NCWRC annual catch summary for the Cape Fear River striped bass electrofishing survey, 2003–2018.

<b>Year</b>	<b>n Sample Events</b>	<b>Effort</b>	<b>n Females</b>	<b>n Males</b>	<b>Total Catch</b>	<b>Mean Index</b>	<b>SD [Index]</b>	<b>SE [Index]</b>	<b>Peak Index</b>
<b>2003</b>	11	18,562	12	4	16	4.5	5.5	1.7	14
<b>2004</b>	8	8,843	20	17	42	25.4	20.1	7.1	57.8
<b>2005</b>	20	61,200	35	42	103	6.5	7.3	1.6	26
<b>2006</b>	12	25,429	2	2	5	1	1.9	0.5	5.8
<b>2007</b>	22	46,557	28	30	120	10.8	15.6	3.3	60
<b>2008</b>	21	45,900	35	64	100	8.8	11.2	2.5	38
<b>2009</b>	21	44,677	27	57	103	9.2	8.3	1.8	24
<b>2010</b>	24	43,200	110	62	182	15.2	13.7	2.8	56
<b>2011</b>	24	42,300	59	37	105	9	12.3	2.5	54
<b>2012</b>	26	45,521	64	55	119	9.2	10	2	30
<b>2013</b>	23	41,400	28	65	99	8.6	14.7	3.1	52
<b>2014</b>	24	43,123	30	71	154	12.9	15.7	3.2	55.6
<b>2015</b>	20	36,259	78	102	193	19.1	27.9	6.2	104
<b>2016</b>	25	45,408	45	145	202	15.9	28.7	5.7	102
<b>2017</b>	19	34,036	47	59	107	11.3	24.7	5.7	86
<b>2018</b>	15	27,315	20	28	58	7.7	6.8	1.7	23.8

**Table 1.9.** Total number of striped bass PIT tagged by all gears and tagger affiliation in the Cape Fear River, 2011–2018.

<b>Tagger</b>	<b>Gear</b>	<b>2011</b>	<b>2012</b>	<b>2013</b>	<b>2014</b>	<b>2015</b>	<b>2016</b>	<b>2017</b>	<b>2018</b>	<b>All</b>
NCDMF	Electrofisher	133	235	336	410	484	388	262	342	2,590
	Gill Net (P915)	11			2	4	3	4		24
	Gill Net (run-around)	9				3	2	6		20
	Hook and line	23	8		8	9	14	11		73
	Trotline			1						1
NCWRC	Electrofisher	72	88	50	99	154	128	33	33	657
Tournament	Hook and line	16	21	38	31	20	33	34	17	210
Volunteer	Hook and line	21	42	34	45	10	24	9		185
All	All	285	394	459	595	684	592	359	392	3,760

**Table 2.10.** Total number of striped bass PIT tagged by gear and tagger affiliation included in the tagging model in the Cape Fear River, 2012–2018.

<b>Tagger</b>	<b>Gear</b>	<b>2012</b>	<b>2013</b>	<b>2014</b>	<b>2015</b>	<b>2016</b>	<b>2017</b>	<b>2018</b>	<b>All</b>
NCDMF	Electrofisher	235	336	410	484	388	262	342	2,457
	Hook and line	8		8	9	14	11		50
NCWRC	Electrofisher	88	50	99	154	128	33	33	585
Tournament	Hook and line	21	38	31	20	33	34	17	194
Volunteer	Hook and line	42	34	45	10	24	9		164
All	All	394	458	593	677	587	349	392	3,450

**Table 2.11.** Mean, standard deviation (SD), minimum, and maximum total length (TL) of striped bass tagged by year, gear, and tagger affiliation included in the tagging model for the Cape Fear River, 2012–2018.

<b>Group</b>	<b>Level</b>	<b>n</b>	<b>Mean TL (mm)</b>	<b>SD [Mean TL]</b>	<b>Minimum TL (mm)</b>	<b>Maximum TL (mm)</b>
<b>Year</b>	2012	394	544.6	118.2	219	846
	2013	458	534.3	109	192	835
	2014	593	540.8	119.2	212	800
	2015	677	508.5	128.9	284	891
	2016	586	525.7	101.8	329	889
	2017	349	540.4	103.4	298	867
	2018	392	569	101.4	337	809
<b>Gear</b>	Hook and line	408	557.8	115.4	330	838
	Electrofishing	3,041	531.5	114.4	192	891
<b>Tagger</b>	NCDMF	2,507	525.2	114.5	219	867
	NCWRC	584	558.7	109.1	192	891
	Tournament	194	537	110.7	330	823
	Volunteer	164	590.7	116.3	355	838

**Table 2.12.** Total number of striped bass PIT tag recaptures by all gears in the Cape Fear River, 2011–2018.

<b>Year</b>	<b>Tagged</b>	<b>Recapture Year</b>								<b>Total Recaptured</b>	<b>Percent Recaptured</b>
		<b>2011</b>	<b>2012</b>	<b>2013</b>	<b>2014</b>	<b>2015</b>	<b>2016</b>	<b>2017</b>	<b>2018</b>		
<b>2011</b>	285	6	8	10	3	2	4	.	.	33	11.6
<b>2012</b>	394	.	4	14	12	6	.	.	1	37	9.4
<b>2013</b>	459	.	.	18	14	9	8	3	3	55	12
<b>2014</b>	595	.	.	.	14	23	5	3	4	49	8.2
<b>2015</b>	684	.	.	.	.	9	8	11	2	30	4.4
<b>2016</b>	592	.	.	.	.	.	10	15	7	32	5.4
<b>2017</b>	359	.	.	.	.	.	.	7	11	18	5
<b>2018</b>	392	.	.	.	.	.	.	.	5	5	1.3
<b>All</b>	3,760	6	12	42	43	49	35	39	33	259	6.9

**Table 2.13.** Total number of striped bass PIT tag recaptures, from electrofishing gear, included in the tagging model for the Cape Fear River, 2012–2018.

Year	Tagged	Recapture Year							Total Recaptured	Percent Recaptured
		2012	2013	2014	2015	2016	2017	2018		
2012	394	4	14	12	5	.	.	1	36	9.1
2013	458	.	18	14	8	8	3	3	54	11.8
2014	593	.	.	14	21	5	3	4	47	7.9
2015	677	.	.	.	9	7	11	2	29	4.3
2016	587	.	.	.	.	10	15	7	32	5.5
2017	349	.	.	.	.	.	7	11	18	5.2
2018	392	.	.	.	.	.	.	5	5	1.3
All	3,450	4	32	40	43	30	39	33	221	6.4

**Table 2.14.** Distance (miles) between release and recapture sites of striped bass included in the tagging model by days at large in the Cape Fear River, 2012–2018.

Days at Large	n	Median	Mean	SD[Mean]	Minimum	Maximum
8 to 20	15	1	2.7	5	0	20
21 to 100	37	1	6.2	11.4	0	54
101 to 200	5	20	14.4	11.5	1	25
201 to 300	25	3	11.2	15	0	65
301 to 400	49	1	4.4	9.9	0	43
401 to 500	23	1	4.8	8.2	0	25
501 to 1,000	40	1	7.2	14.3	0	62
1,001 to 1,500	20	1	7	15.1	0	65
1,501 to 2,000	6	1	1.5	0.8	1	3
2,001 to 2,500+	1	56	56		56	56
All	221	1	6.5	12.3	0	65

**Table 2.15.** Mean, standard deviation (SD), minimum, and maximum number of days at large of striped bass recaptured by year, 2012–2018.

<b>Year</b>	<b>n</b>	<b>Mean</b>	<b>SD[Mean]</b>	<b>Minimum</b>	<b>Maximum</b>
<b>2012</b>	36	594.4	418.5	14	2,232
<b>2013</b>	54	559.8	485.3	12	1,870
<b>2014</b>	47	434.9	443.9	8	1,695
<b>2015</b>	29	412.6	348.7	21	1,371
<b>2016</b>	32	398.9	217.1	8	826
<b>2017</b>	18	231.6	189	12	467
<b>2018</b>	5	28.6	22.5	12	57
<b>All</b>	221	457.5	406.9	8	2,232

**Table 2.16.** Mean, standard deviation (SD), minimum, and maximum total length (TL) of striped bass recaptured by year in the Cape Fear River, 2012–2018.

<b>Year</b>	<b>n</b>	<b>Mean TL (mm)</b>	<b>SE[Mean TL]</b>	<b>Minimum TL (mm)</b>	<b>Maximum TL (mm)</b>
<b>2012</b>	36	611.6	94	456	845
<b>2013</b>	54	592.5	69.9	469	747
<b>2014</b>	47	600.2	89.1	380	814
<b>2015</b>	29	560.5	104.1	359	760
<b>2016</b>	32	570.7	83.5	382	766
<b>2017</b>	18	577.2	96.4	397	766
<b>2018</b>	5	481.2	51.3	402	534
<b>All</b>	221	586.7	89.1	359	845



**Table 2.17.** Mean, standard deviation (SD), minimum, and maximum growth (mm) of recaptured striped bass by days at large in the Cape Fear River, 2012–2018.

<b>Days at Large</b>	<b>n</b>	<b>Mean (mm)</b>	<b>SD[Mean]</b>	<b>Minimum (mm)<sup>6</sup></b>	<b>Maximum (mm)</b>
8 to 20	11	2.7	2.8	0	9
21 to 100	29	11.8	13.6	0	69
101 to 200	4	29.5	24.4	3	62
201 to 300	22	64.5	45.8	4	192
301 to 400	49	67.6	45.3	7	255
401 to 500	21	80.5	43.10	19	154
501 to 1,000	41	126.9	63.5	15	221
1,001 to 1,500	21	133.3	73.4	29	306
1,501 to 2,000	6	242.8	55.9	181	332
2,001 to 2,500+	1	367.0		367	367

**Table 2.18.** Mean, standard deviation (SD), minimum, and maximum growth (mm) of striped bass recaptured by year in the Cape Fear River, 2012–2018.

<b>Year</b>	<b>n</b>	<b>Mean (mm/day)</b>	<b>SD[Mean]</b>	<b>Minimum (mm/day)<sup>6</sup></b>	<b>Maximum (mm/day)</b>
<b>2012</b>	33	0.18	0.10	0.02	0.39
<b>2013</b>	52	0.16	0.14	0.01	0.78
<b>2014</b>	47	0.19	0.15	0.00	0.77
<b>2015</b>	24	0.20	0.12	0.04	0.57
<b>2016</b>	30	0.21	0.13	0.04	0.54
<b>2017</b>	16	0.24	0.18	0.08	0.75
<b>2018</b>	3	0.23	0.20	0.11	0.46
<b>All</b>	205	0.19	0.14	0.00	0.78

---

<sup>6</sup> Negative values were removed

**Table 3.1.** Summary of parameter values used to develop the demographic matrix model.

Parameter	Notation	Cape Fear	Neuse	Tar-Pamlico	Reference
Maximum age (yr)	$T$	7	11	11	Age-length data from three rivers 2004–2017
Asymtotic length (mm)	$L_{\infty}$	759.7	874.4	838.7	Posterior median estimated using age-length data from three rivers 2004–2017
Growth coefficient ( $\text{yr}^{-1}$ )	$K$	0.35	0.185	0.197	
Age at which fish has a length of zero (yr)	$t_0$	-0.003	-1.914	-1.696	
Maximum age for calculating $M$ (yr)	$t_{max}$	7	11	11	Maximum age assumed for matrix model
Minimum age for calculating $M$ (yr)	$t_{min}$		1		Minimum age assumed for matrix model
Mean constant natural mortality lower bound ( $\text{yr}^{-1}$ )	$m_1$	0.6 for age 1-3; 0.1 for age 4+			Harris and Hightower 2017;
Mean constant natural mortality upper bound ( $\text{yr}^{-1}$ )	$m_2$	1 for age 1-3; 0.5 for age 4+			Bradley et al. 2018
Coefficient of variation	$CV$	U(20%, 40%)			Jiao et al. 2009; Li and Jiao 2015
Fishery selectivity	$g$	0.014 for age 1; 0.13 for age 2; 0.6 for age 3; 0.94 for age 4; 0.99 for age 5; 1 for age > 5	0 for age < 3; 0.28 for age 3; 1 for age > 3	0 for age < 3; 0.12 for age 3; 1 for age > 3	Estimated from a catch-curve analysis using 2017 fishery-dependent data for Neuse and Tar-Pamlico Rivers, and 2017 fishery-independent data for Cape Fear River
Fishing mortality ( $\text{yr}^{-1}$ )	$F$	0, 0.2, 0.4, 0.6, 0.8, 1			Hypothetical scenarios based on Rachels and Ricks 2015, Bradley et al. 2018
Sex ratio		1:1			
Proportion of viable eggs	$x$	0.64			Bradley et al. 2018b
Survival of offspring from birth to next census	$S_0$	0.000017			Estimated using Table 1 in Dahlberg 1979
Fecundity (number of eggs per mature female)	$E$	$\ln(E_t)=12.484+0.205t$	$\ln(E_t)=12.52+0.214t$	$\ln(E_t)=12.429+0.203t$	Estimated using survey data from Neuse River and Tar-Pamlico River 2013–2014

**Table 3.1.** (*continued*) Summary of parameter values used to develop the demographic matrix model.

<b>Parameter</b>	<b>Notation</b>	<b>Cape Fear</b>	<b>Neuse</b>	<b>Tar-Pamlico</b>	<b>Reference</b>
Maturity	$w$		0 for age $\leq 2$ , 1 for age $\geq 5$		Survey data from Neuse River and Tar-Pamlico River 2013–2014; Olsen and Rulifson 1992; Boyd 2011
Maturity lower bound	$w_1$		0.29 for age 3, 0.94 for age 4		
Maturity upper bound	$w_2$		1 for ages 3 and 4		

**Table 3.2.** Initial year age structure for fishery management strategy evaluation.

Age	Proportion
<b>1</b>	0.35
<b>2</b>	0.16
<b>3</b>	0.12
<b>4</b>	0.1
<b>5</b>	0.08
<b>6</b>	0.08
<b>7</b>	0.06
<b>8</b>	0.05
<b>9</b>	0
<b>10</b>	0
<b>11</b>	0

**Table 3.3.** Population growth rate estimates from the matrix model. Pr is the probability of population growth rate greater than one.

<i>F</i>	Cape Fear				Neuse				Tar-Pamlico			
	Median	Lower	Upper	Pr	Median	Lower	Upper	Pr	Median	Lower	Upper	Pr
<b>0</b>	1.01	0.70	1.39	0.52	1.13	0.83	1.48	0.80	1.10	0.81	1.44	0.74
<b>0.2</b>	0.94	0.64	1.30	0.36	1.05	0.76	1.39	0.63	1.02	0.74	1.36	0.56
<b>0.4</b>	0.88	0.60	1.24	0.24	0.99	0.71	1.34	0.47	0.97	0.69	1.31	0.41
<b>0.6</b>	0.83	0.56	1.20	0.15	0.94	0.66	1.30	0.35	0.92	0.65	1.28	0.32
<b>0.8</b>	0.79	0.52	1.14	0.10	0.90	0.62	1.26	0.27	0.89	0.61	1.24	0.24
<b>1</b>	0.75	0.49	1.10	0.07	0.87	0.59	1.24	0.23	0.86	0.59	1.22	0.20

**Table 4.1.** Cape Fear River tagging model parameters and priors.  $U$  denotes the uniform distribution.

Parameters	Values	Reference
<b><u>Constant parameters</u></b>		
Survival from tagging procedure	$\phi=1$	
Immediate tag retention probability	$\rho=1$	
Tag reporting rate	$\lambda=1$	
Tag loss	$\Omega=0$	
<b><u>Priors</u></b>		
Instantaneous total mortality ( $\text{yr}^{-1}$ )	$\bar{Z} \sim U(0.1, 1.5)$	Bradley et al. 2018; Harris and Hightower 2017
Instantaneous survey mortality ( $\text{month}^{-1}$ )	$U \sim U(0, 0.1)$	
Standard deviation of log-total mortality	$\sigma_Z \sim U(0.001, 1)$	

**Table 4.2.** Estimated instantaneous total mortality ( $Z$ ,  $\text{yr}^{-1}$ ) due to natural causes and fishing, estimated abundance (number) and estimated capture probability ( $\alpha$ ) from the tagging model in the Cape Fear River. Median—posterior median; Lower and Upper—lower and upper 95% credible intervals.

Year	$Z$ ( $\text{yr}^{-1}$ )			$N$ (number)			$\alpha$		
	Median	Lower	Upper	Median	Lower	Upper	Median	Lower	Upper
<b>2012</b>	0.96	0.53	1.43	10,983	5,418	23,479	0.036	0.017	0.073
<b>2013</b>	0.58	0.21	1.00	4,532	3,024	6,921	0.101	0.066	0.151
<b>2014</b>	1.13	0.71	1.47	7,372	4,623	11,708	0.080	0.051	0.128
<b>2015</b>	0.81	0.37	1.29	3,778	2,655	5,825	0.179	0.116	0.255
<b>2016</b>	0.63	0.24	1.09	3,335	2,191	5,573	0.176	0.105	0.268
<b>2017</b>	0.53	0.18	0.97	1,578	1,168	2,293	0.221	0.152	0.299
<b>2018</b>	0.73	0.21	1.41	1,914	1,415	2,765	0.205	0.142	0.277

**Table 4.3.** Estimated striped bass effort and catch in the Cape Fear River. (Source: Costal Angling Program (CAP) Central Southern Management Area (CSMA) recreational striped bass creel survey)

Year	Month	n Striped Bass Trips	n Striped Bass Hours	Harvest		Discard (numbers)				Total Catch (n fish)
				numbers	pounds	legal sized	over creel	undersized	slot	
2013	Jan									
	Feb									
	Mar									
	Apr	92	399					81		81
	May	165	470			263		11		274
	<b>Total</b>	257	870			263		92		355
	<b>PSE</b>	48.6	63.1			90.8		55.6		
2014	Jan									
	Feb									
	Mar	134	558							
	Apr	138	833			708		703		1,412
	May	161	748			122		17		139
	<b>Total</b>	433	2,140			830		721		1,551
	<b>PSE</b>	42.9	45.9			72.7		77.5		
2015	Jan									
	Feb									
	Mar	110	422			22				
	Apr	19	181					162		162
	May	79	100					15		15
	<b>Total</b>	209	702			22		176		199
	<b>PSE</b>	50.1	53			100		57.4		
2016	Jan									
	Feb									
	Mar	179	750			10		12		22
	Apr	87	315			17				17
	May	126	399			588				588
	<b>Total</b>	391	1,464			616		12		628
	<b>PSE</b>	46.4	44.4			95.8		100		

**Table 4.3. (continued)** Estimated striped bass effort and catch in the Cape Fear River. (Source: Costal Angling Program (CAP) Central Southern Management Area (CSMA) recreational striped bass creel survey)

Year	Month	n Striped Bass Trips	n Striped Bass Hours	Harvest		Discard (numbers)				Total Catch (n fish)
				numbers	pounds	legal sized	over creel	undersized	slot	
2017	Jan									
	Feb	26	159			14				14
	Mar									
	Apr									
	May									
	Total	26	159			14				14
	PSE	100.0	100			100				
2018	Jan									
	Feb									
	Mar	18	35							
	Apr									
	May	7	26			140				140
	Total	24	61			140				140
	PSE	77.1	71.5			70.8				

**Table 5.1.** Fit of the candidate models. Com = commercial; Rec = recreational; DO = dissolved oxygen (mg/L); K = the number of parameters; AIC<sub>c</sub> = Akaike’s information criterion corrected for small sample size; Δ<sub>i</sub> = Akaike difference; w<sub>i</sub> = Akaike weight. The candidate models from Rachels and Ricks (2018) are formatted in **bold**.

Model	<b>K</b>	<b>AIC<sub>c</sub></b>	<b>Δ<sub>i</sub></b>	<b>w<sub>i</sub></b>	<b>R<sup>2</sup></b>
<b>Com effort</b>	<b>2</b>	<b>-20.41</b>	<b>0</b>	<b>0.152</b>	<b>0.21</b>
<b>Com effort, Com harvest</b>	<b>3</b>	<b>-19.27</b>	<b>1.14</b>	<b>0.086</b>	<b>0.43</b>
Rec effort	2	-19.15	1.26	0.081	0.11
Rec discard	2	-18.89	1.52	0.071	0.09
<b>Com harvest</b>	<b>2</b>	<b>-18.88</b>	<b>1.53</b>	<b>0.071</b>	<b>0.08</b>
Rec total catch	2	-18.77	1.64	0.067	0.075
Rec effort, Com effort	3	-18.68	1.73	0.064	0.39
Rec harvest, Rec effort	3	-18.5	1.91	0.058	0.38
<b>DO</b>	<b>2</b>	<b>-18</b>	<b>2.41</b>	<b>0.045</b>	<b>0.001</b>
Rec harvest	2	-17.99	2.42	0.045	0
Rec total removal	2	-17.99	2.42	0.045	0
<b>Com effort, Temperature</b>	<b>3</b>	<b>-17.61</b>	<b>2.8</b>	<b>0.037</b>	<b>0.32</b>
<b>Com effort, DO</b>	<b>3</b>	<b>-16.62</b>	<b>3.79</b>	<b>0.023</b>	<b>0.25</b>
Rec harvest, Com effort	3	-16.57	3.84	0.022	0.25
Rec effort, Temperature	3	-16.1	4.31	0.018	0.21
Rec effort, DO	3	-15.69	4.72	0.014	0.18
Rec discard, Rec effort	3	-15.36	5.05	0.012	0.15
Rec effort, Com harvest	3	-15.26	5.15	0.012	0.14
<b>Com harvest, Temperature</b>	<b>3</b>	<b>-14.94</b>	<b>5.47</b>	<b>0.010</b>	<b>0.12</b>
Rec harvest, Com harvest	3	-14.86	5.55	0.009	0.11
Rec discard, Temperature	3	-14.76	5.65	0.009	0.1
Rec discard, DO	3	-14.65	5.76	0.009	0.09
Rec discard, Rec harvest	3	-14.61	5.8	0.008	0.09
<b>Com harvest, DO</b>	<b>3</b>	<b>-14.59</b>	<b>5.82</b>	<b>0.008</b>	<b>0.08</b>
<b>DO, Temperature</b>	<b>3</b>	<b>-13.86</b>	<b>6.55</b>	<b>0.006</b>	<b>0.015</b>
Rec harvest, Temperature	3	-13.82	6.59	0.006	0.01
Rec harvest, DO	3	-13.72	6.69	0.005	0.002
Rec discard, Rec effort, Rec harvest	4	-12.82	7.59	0.003	0.4
<b>Com effort, DO, Temperature</b>	<b>4</b>	<b>-11.88</b>	<b>8.53</b>	<b>0.002</b>	<b>0.34</b>
<b>Com harvest, DO, Temperature</b>	<b>4</b>	<b>-10.51</b>	<b>9.9</b>	<b>0.001</b>	<b>0.24</b>
Rec discard, Rec harvest, Com harvest	4	-10.37	10.04	0.001	0.23
<b>Com effort, Com harvest, DO, Temperature</b>	<b>5</b>	<b>-6.55</b>	<b>13.86</b>	<b>0.000</b>	<b>0.54</b>



**Table 6.1.** Estimated parameter values of the von Bertalanffy age-length relationship and their associated standard errors (SE) where total length was measured in millimeters (n=166).

Parameter	Value	SE
$L_{\infty}$	787	65
$K$	0.26	0.084
$t_0$	-0.94	0.72

**Table 6.2.** Estimated parameter values of the length-weight relationship and their associated standard errors (SE) where total length was measured in millimeters and weight was measured in grams (n=198).

Parameter	Value	SE
$a$	2.4E-06	9.5E-07
$b$	3.2	6.2E-02

**Table 6.3.** Estimated natural mortality ( $M$ ) at age based on Lorenzen's (1996) approach. The values given represent instantaneous rates.

Age	$M$
1	0.60
2	0.45
3	0.38
4	0.34
5	0.31
6	0.30
7	0.29

**Table 6.4.** Estimated parameter values of the logistic length-maturity relationship and their associated standard errors (SE) where total length was measured in millimeters (n=170).

Parameter	Value	SE
$a$	-49	18
$b$	0.10	0.037

**Table 6.5.** Definitions of symbols used in the per-recruit equations.

Symbol	Definition	Units
$t$	age	years
$F_t$	fishing mortality at age $t$	year <sup>-1</sup>
$F_{full}$	fully-recruited fishing mortality	year <sup>-1</sup>
$S_t$	selectivity at age $t$	proportion
$Z_t$	total mortality at age $t$	year <sup>-1</sup>
$M_t$	natural mortality at age $t$	year <sup>-1</sup>
$N_1$	number of fish at age 1	numbers of fish
$N_t$	number of fish at age $t$	numbers of fish
$B_t$	population biomass at age $t$	kilograms
$w_t$	individual weight at age $t$	grams
$U_t$	population size at age $t$ for non-hatchery origin fish	numbers of fish
$h$	assumed proportion of hatchery fish in the population	proportion
$H_t$	population size at age $t$ for hatchery-origin fish	numbers of fish
$C_t$	catch at age $t$	numbers of fish
YPR	yield per recruit	numbers of fish
$W_t$	weight of catch at age $t$	kilograms
WPR	weight per recruit	kilograms
SSB	spawning stock biomass	kilograms
$SSU_t$	SSB at age $t$ for non-hatchery female fish	kilograms
$p$	proportion of individuals in the population that are female	proportion
$mat_t$	maturity at age $t$	proportion
$f$	proportion of fishing mortality that occurs before spawning	proportion
$m$	proportion of natural mortality that occurs before spawning	proportion
SSU/R	SSB per recruit for the non-hatchery female fish	kilograms
SSH <sub><math>t</math></sub>	SSB at age $t$ for hatchery-origin female fish	kilograms
SSH/R	SSB per recruit for hatchery-origin female fish	kilograms
SSB/R	SSB per recruit for the entire population	kilograms
EU <sub><math>t</math></sub>	total number of eggs at age $t$ for the non-hatchery female fish	numbers of eggs
EU/R	eggs per recruit for the non-hatchery female fish	numbers of eggs
EH <sub><math>t</math></sub>	total number of eggs at age $t$ for hatchery-origin female fish	numbers of eggs
EH/R	eggs per recruit for hatchery-origin female fish	numbers of eggs
E/R	eggs per recruit for the entire population	numbers of eggs
%SPR	spawning potential ratio	percentage

**Table 6.6.** Sample frequency at (genetic) age of striped bass collected in the Neuse River by the NCWRC's Spawning Stock Survey in 2017. Catches have been standardized to a collection time of 19 minutes.

Age	Frequency
1	0
2	8
3	123
4	88
5	37
6	36
7	7

**Table 6.7.** Estimates of fishing mortality ( $F$ ) at age derived from the catch curve analysis.

Age	$F$
1	0
2	0.0092
3	0.25
4	0.33
5	0.33
6	0.33
7+	0.33

**Table 7.1.** Number of scales, otoliths, and genetic (PBT) structures collected by NCDMF available for CSMA striped bass age determination, 1975–2018. Genetic (PBT) structures are only available from 2016–2018.

<b>Year</b>	<b>Scale</b>	<b>Otolith</b>	<b>PBT</b>		<b>Year</b>	<b>Scale</b>	<b>Otolith</b>	<b>PBT</b>
<b>1975</b>	77	0			<b>1997</b>	0	0	
<b>1976</b>	4	0			<b>1998</b>	1	8	
<b>1977</b>	2	0			<b>1999</b>	18	0	
<b>1978</b>	32	0			<b>2000</b>	57	0	
<b>1979</b>	29	0			<b>2001</b>	50	0	
<b>1980</b>	105	0			<b>2002</b>	204	0	
<b>1981</b>	0	0			<b>2003</b>	334	64	
<b>1982</b>	0	0			<b>2004</b>	254	66	
<b>1983</b>	16	0			<b>2005</b>	532	86	
<b>1984</b>	18	0			<b>2006</b>	484	115	
<b>1985</b>	9	0			<b>2007</b>	335	87	
<b>1986</b>	0	0			<b>2008</b>	242	114	
<b>1987</b>	2	0			<b>2009</b>	316	39	
<b>1988</b>	4	0			<b>2010</b>	671	156	
<b>1989</b>	7	0			<b>2011</b>	688	196	
<b>1990</b>	0	0			<b>2012</b>	766	248	
<b>1991</b>	0	0			<b>2013</b>	993	189	
<b>1992</b>	0	0			<b>2014</b>	376	181	
<b>1993</b>	0	0			<b>2015</b>	413	107	
<b>1994</b>	0	0			<b>2016</b>	592	123	322
<b>1995</b>	0	0			<b>2017</b>	599	132	261
<b>1996</b>	0	0			<b>2018</b>	719	219	201
					<b>Total</b>	8,949	2,130	784

**Table 7.2.** Mean percentage age bias (bias compared to genetic age) for each reader for overall age bias and age bias by method type (standard deviation in parentheses). Cells with no values indicate the reader performed no readings for that method type.

Reader ID	Overall % Age Bias	Otolith % Age Bias	Scale % Age Bias
1	-0.274 (14.0)	-0.899 (8.5)	-0.0935 (15.2)
2	15.3 (22.2)	-1.81 (7.3)	23.1 (22.3)
3	-0.603 (13.3)		-0.603 (13.3)
4	-1.12 (12.4)		-1.12 (12.4)
5	7.81 (17.3)		7.81 (17.3)

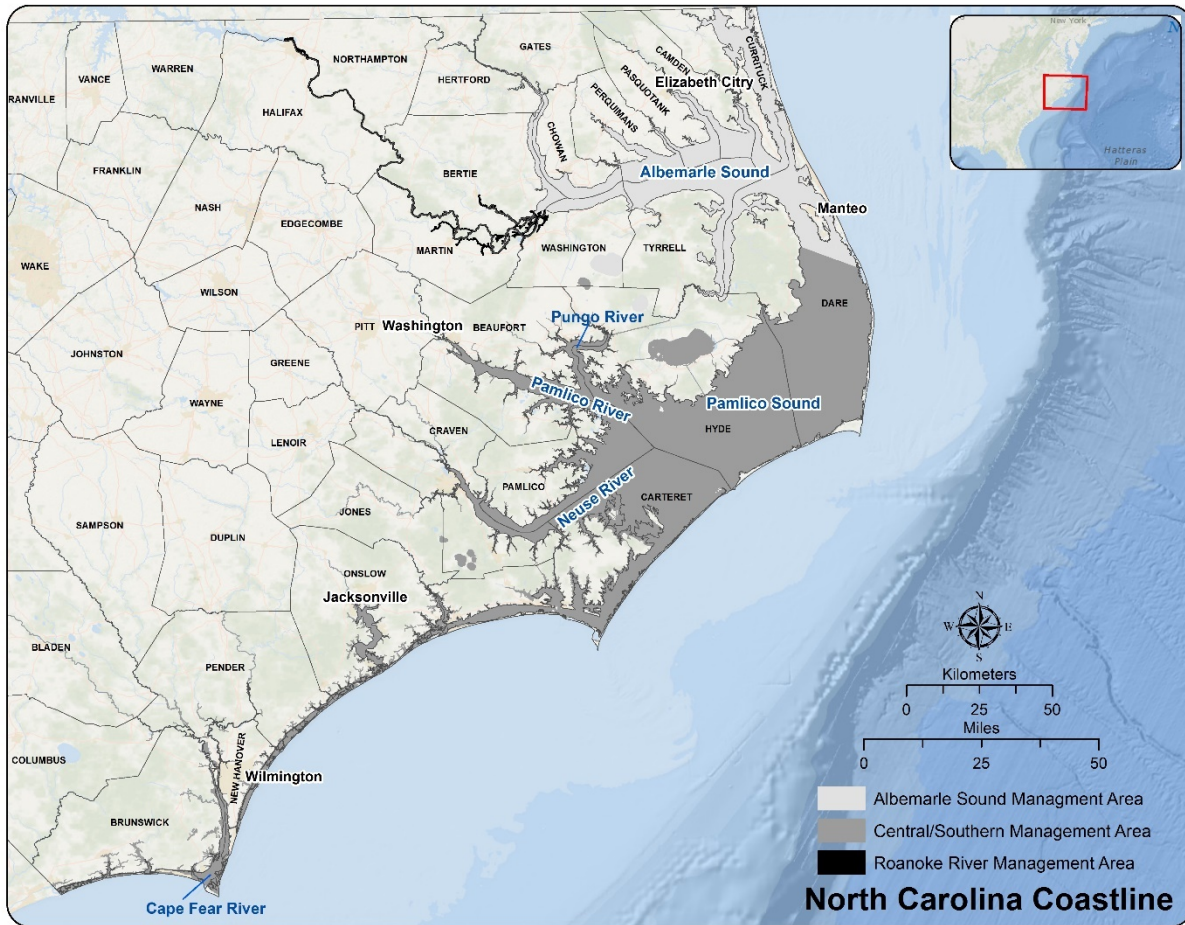
**Table 7.3.** Parameter estimates from Bayesian generalized linear mixed effects model for scale ages and otolith ages compared to genetic ages. Estimates are median of posterior distributions with confidence interval in parentheses.

Parameter	Estimates	
<b>Reader ID random effects</b>	$\alpha_1$	-0.858 (-1.31, -0.304)
	$\alpha_2$	2.47 (0.373, 8.96)
	$\alpha_3$	-0.931 (-1.47, -0.399)
	$\alpha_4$	-1.01 (-1.61, -0.505)
	$\alpha_5$	0.280 (-0.527, 2.63)
	$p_r$	1.70 (0.1, 10.7)
	$\sigma_r$	1.67 (0.589, 5.34)
<b>Ageing method fixed effects</b>		
<b>Otolith:</b>	$\gamma_1$	-1.19 (-2.88, 0.507)
<b>Scale:</b>	$\gamma_2$	7.90 (1.37, 16.6)
<b>Random error</b>		
	$p_e$	98.3 (89.3, 99.9)
	$\sigma_e$	15.5 (15.0, 16.0)

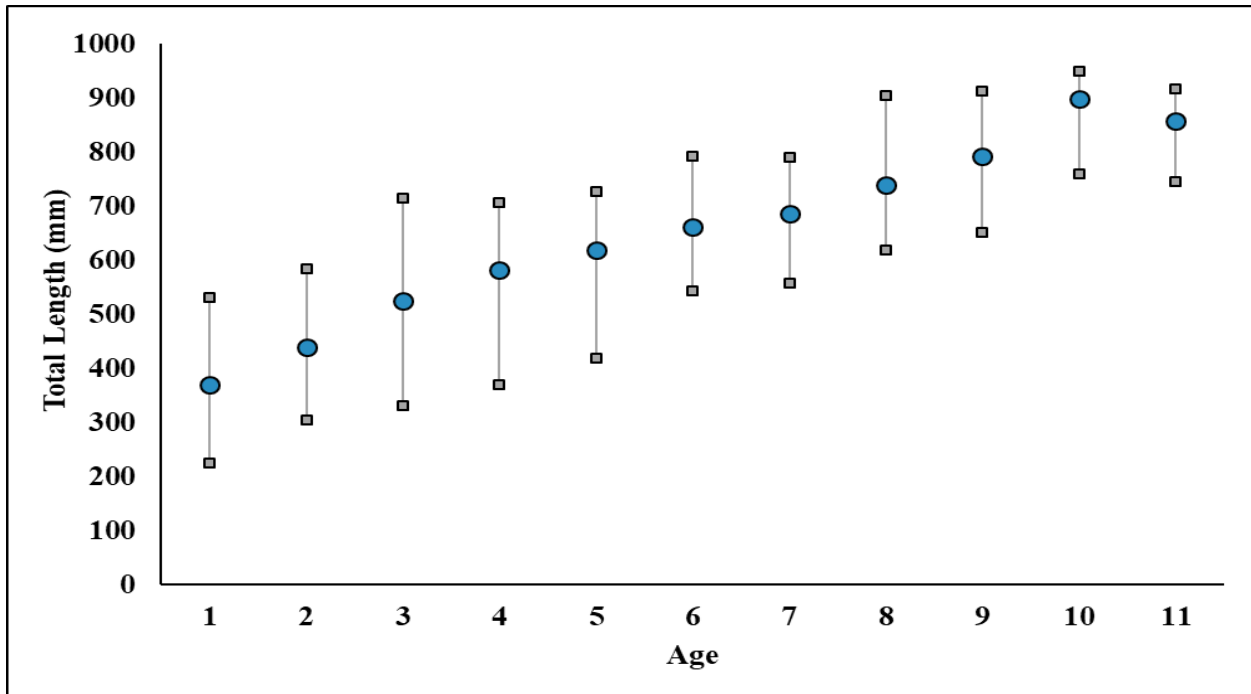
**Table 7.4.** Coefficient of variation (%) analyses results for between readers for scale ages. Values in parentheses are percent agreement. Values in **bold** are significant ( $P < 0.01$ ). Between reader coefficients of variation differ depending on which reader is the reference reader.

	Reader						
	1	2	3	4	5	6	7
1		<b>6.1 (31)</b>	<b>4.7 (65)</b>	<b>3.2 (59)</b>	3.8 (72)	<b>5.4 (24)</b>	<b>4.4 (19)</b>
2	<b>5.2 (31)</b>		<b>4.3 (38)</b>	<b>3.8 (46)</b>	<b>5.5 (30)</b>		
3	<b>5.1 (65)</b>	<b>6.3 (38)</b>		<b>3.2 (67)</b>	2.5 (88)		
Reader 4	<b>5.8 (59)</b>	<b>5.4 (46)</b>	<b>4.9 (67)</b>		<b>6.0 (66)</b>		
5	3.6 (72)	<b>7.0 (30)</b>	2.8 (88)	<b>3.1 (66)</b>			<b>4.3 (22)</b>
6	<b>5.2 (24)</b>						
7	<b>4.0 (19)</b>				<b>3.6 (22)</b>		

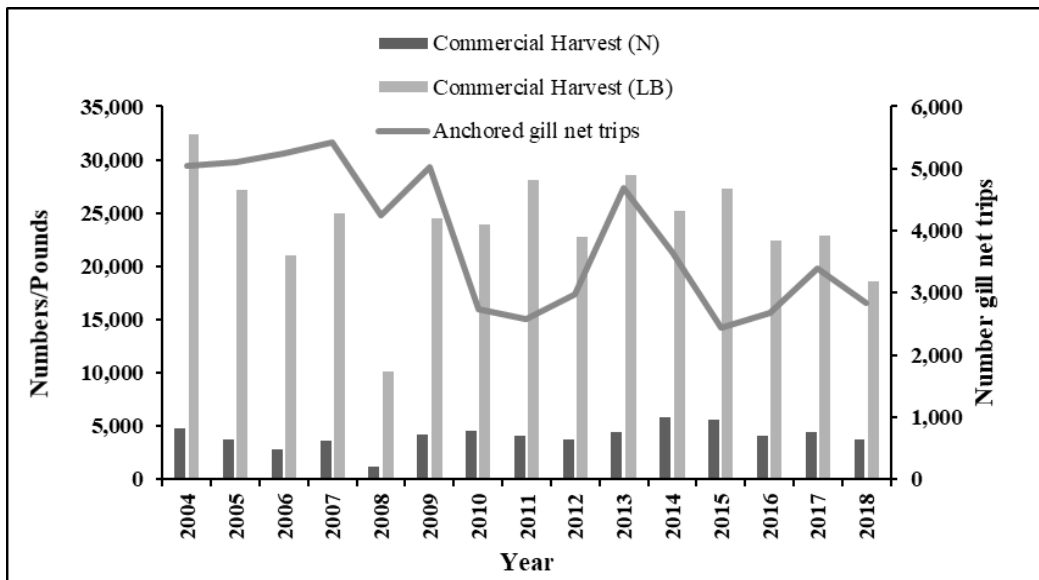
# 11 FIGURES



**Figure 1.1.** Boundary lines between the Albemarle Sound Management Area, Central Southern Management Area, and the Roanoke River Management Area.

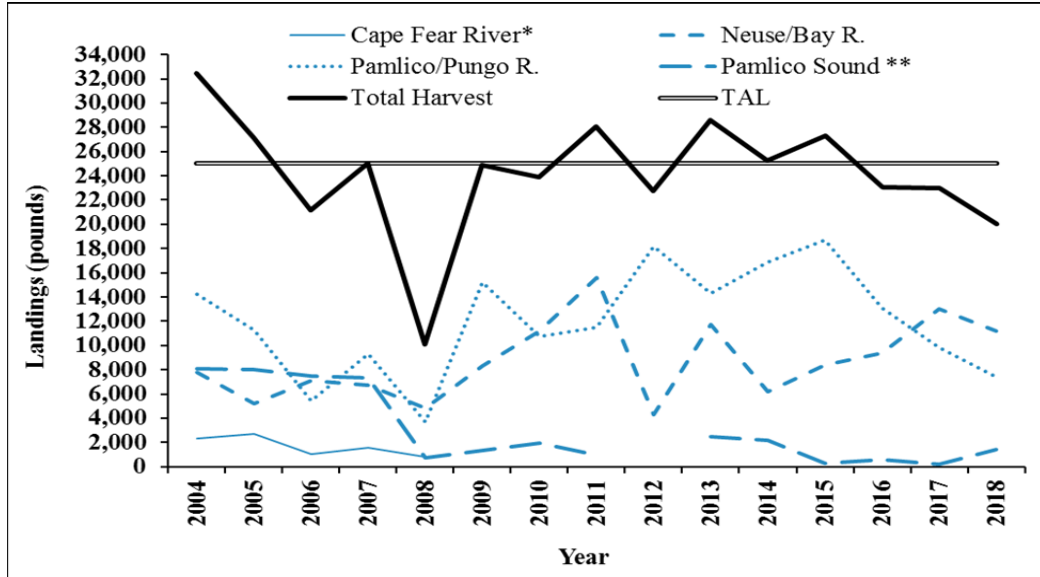


**Figure 1.2.** CSMA striped bass length at age based on otolith and genetic age samples collected by NCDMF, 2004–2018. Blue circles represent the mean size at a given age while the grey squares represent the minimum and maximum observed size for each age.

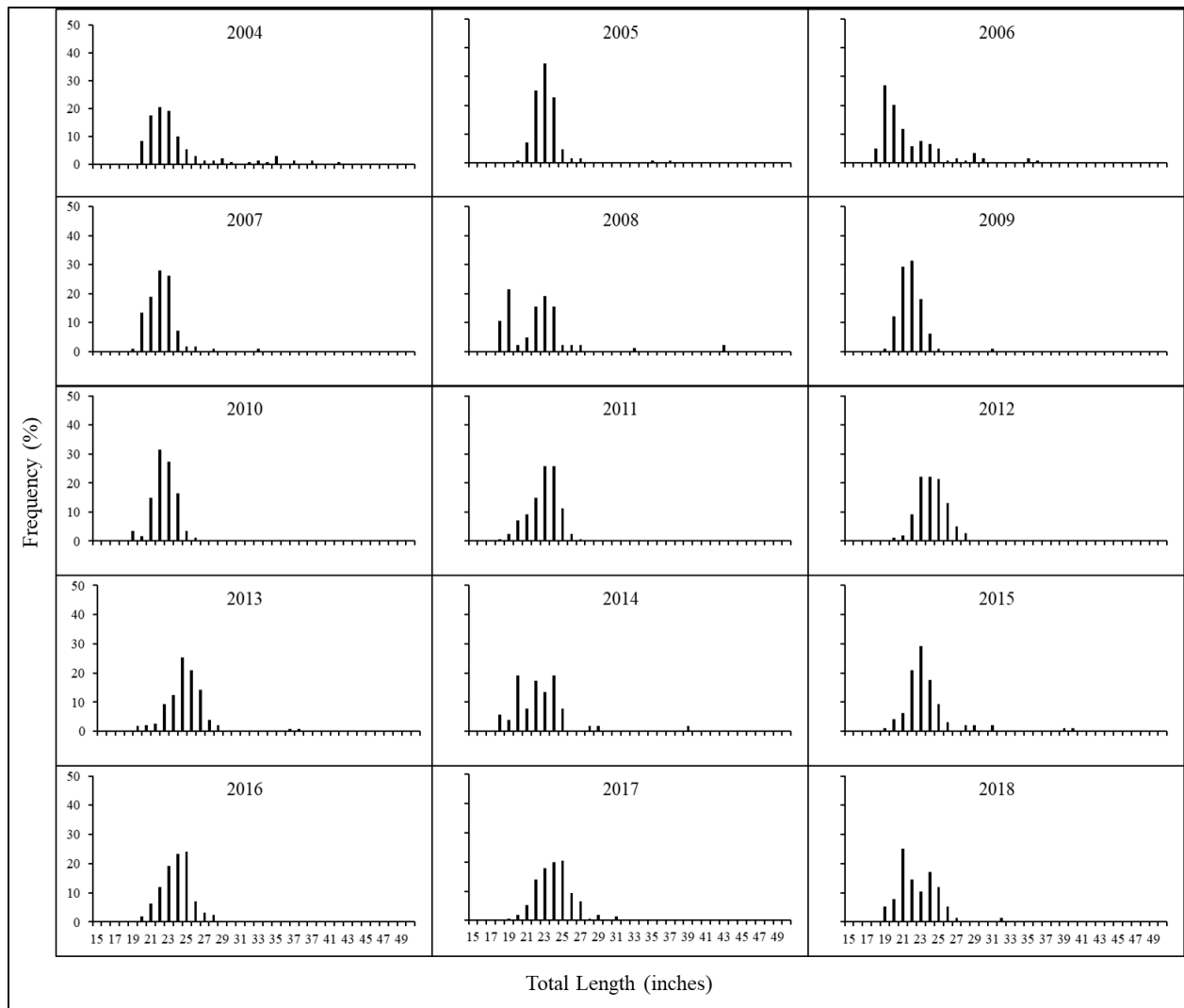




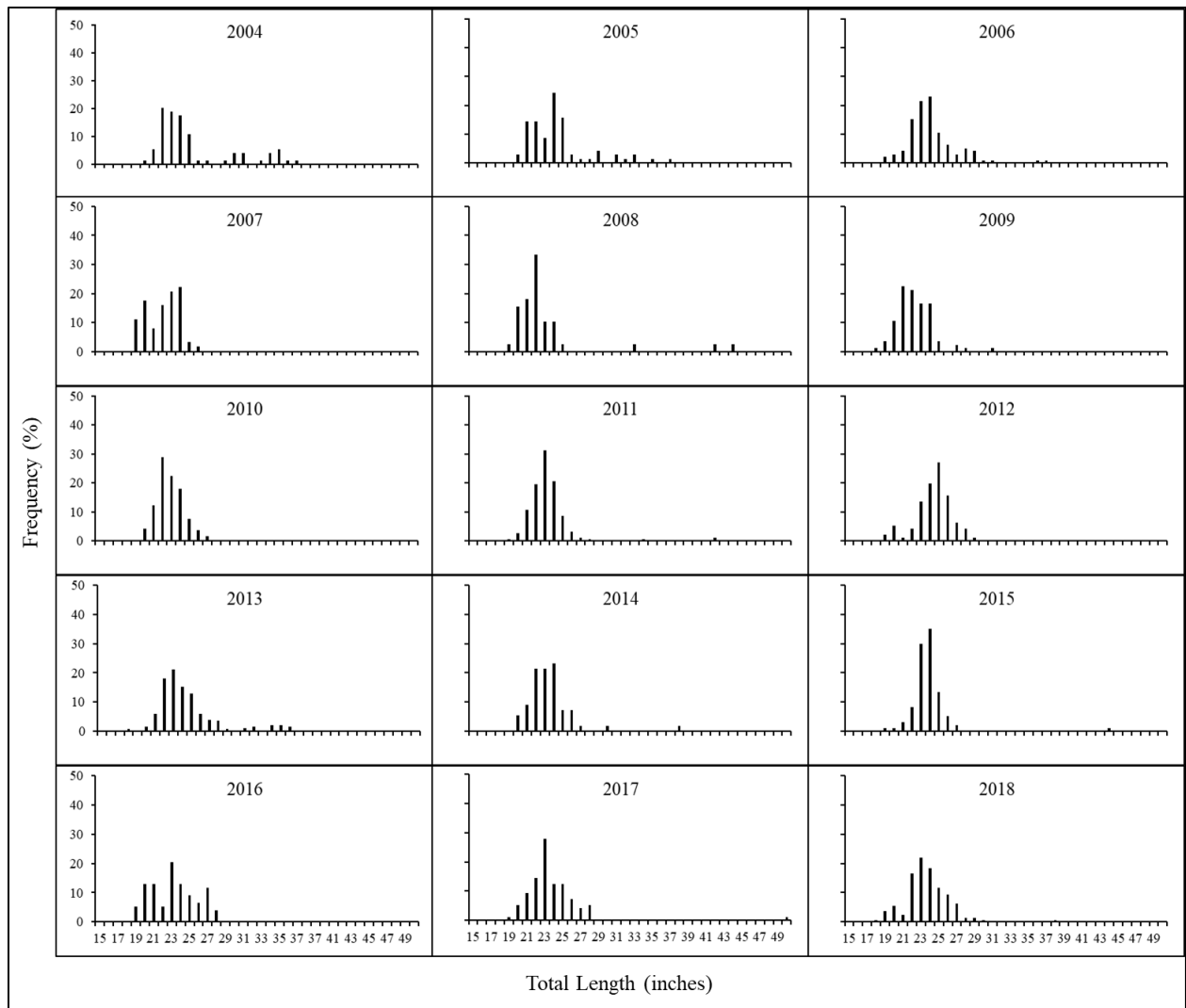
**Figure 2.1.** Commercial striped bass harvest in numbers and pounds and anchored gill-net trips in the Tar-Pamlico, Pungo, Neuse, and Bay rivers, 2004–2018.



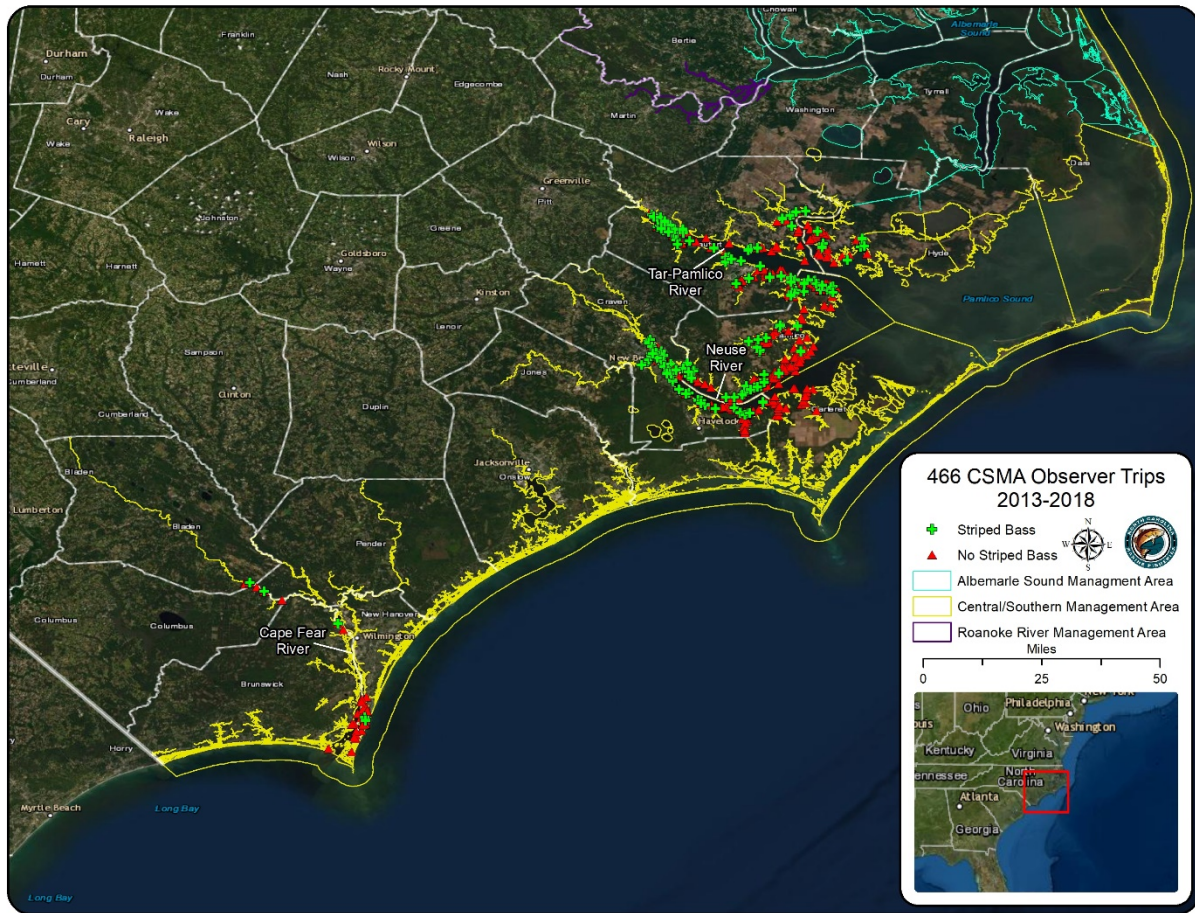
**Figure 2.2.** Commercial striped bass harvest by system, and the TAL in the CSMA, 2004–2018. \*There has been a harvest moratorium in the Cape Fear River since 2008. \*\*Landings data for the Pamlico Sound in 2012 are confidential.



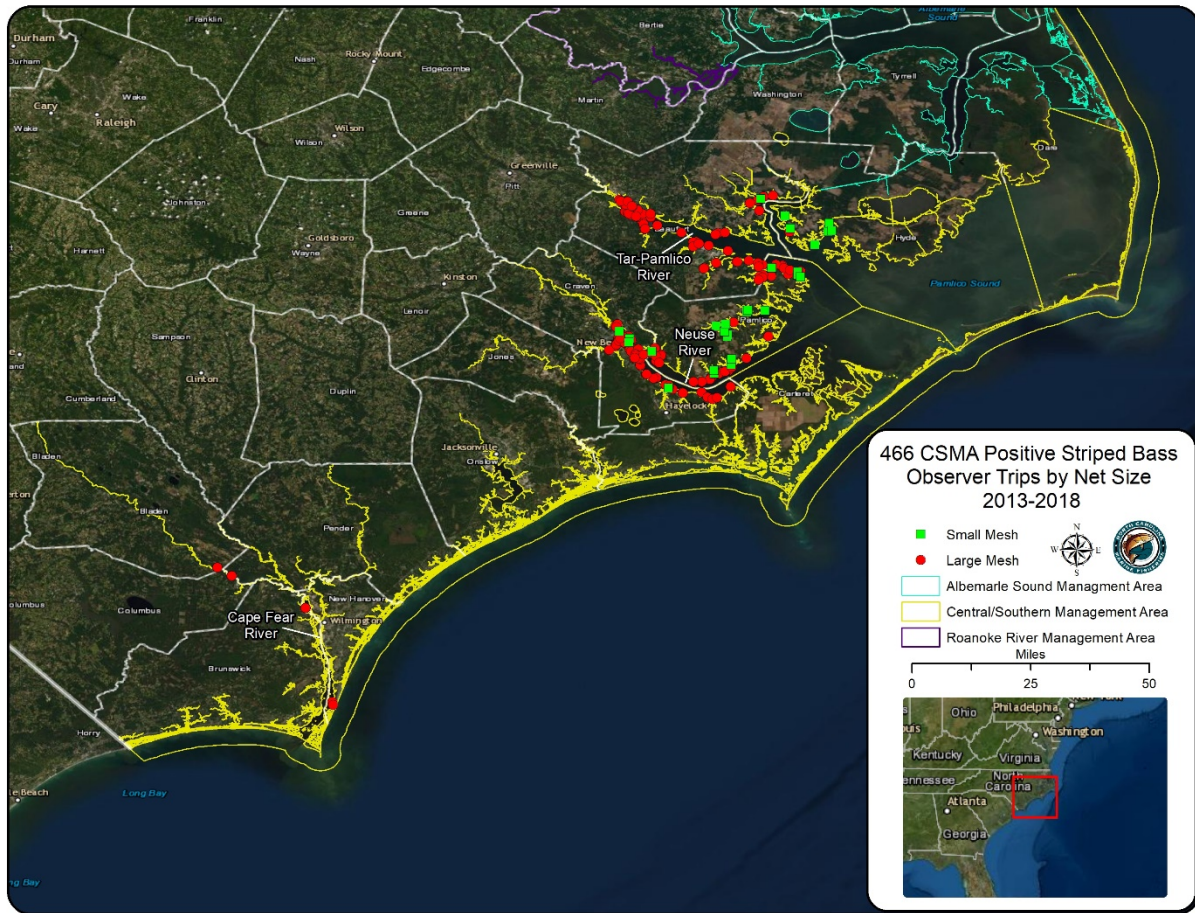
**Figure 2.3.** Length frequency of CSMA striped bass landed commercially in the Tar-Pamlico and Pungo rivers, 2004–2018.



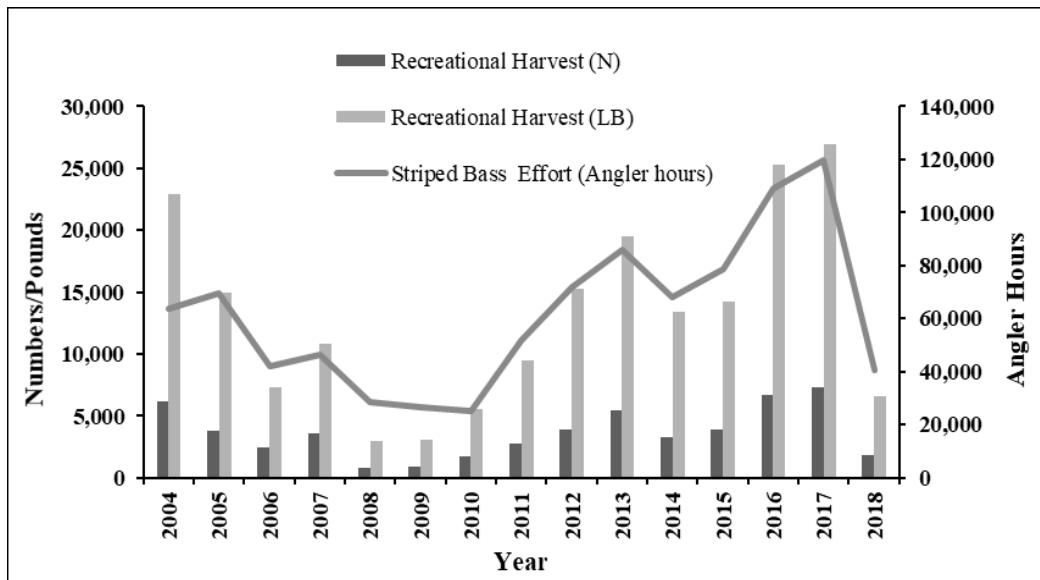
**Figure 2.4.** Length frequency of CSMA striped bass landed commercially in the Neuse and Bay rivers, 2004–2018.



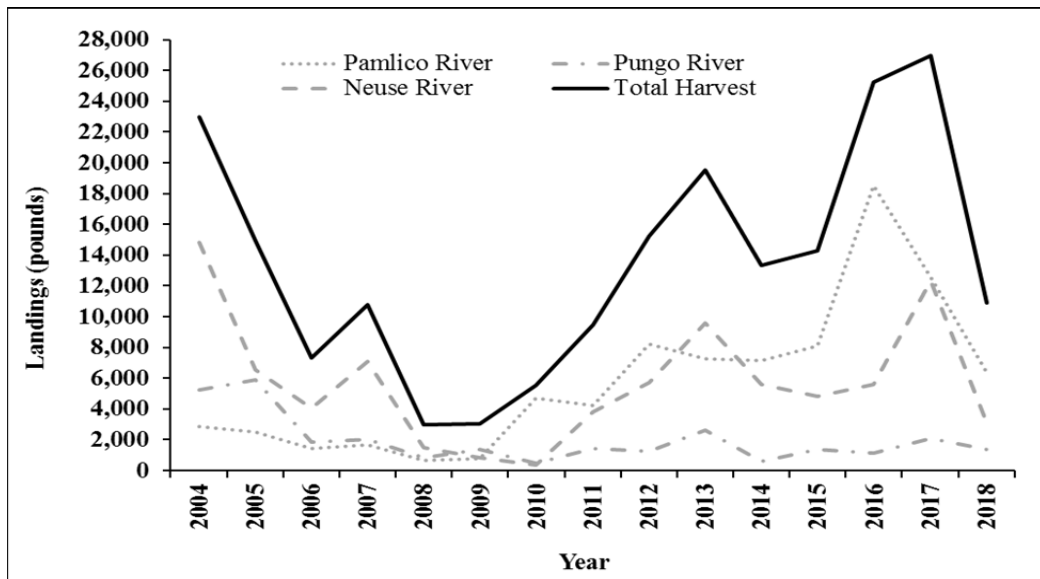
**Figure 2.5.** Program 466 CSMA observer trips by the presence or absence of striped bass, 2013–2018. The cross sign is an observer trip that encountered a striped bass (n=284), and the triangle is an observer trip that did not encounter striped bass (n=789).



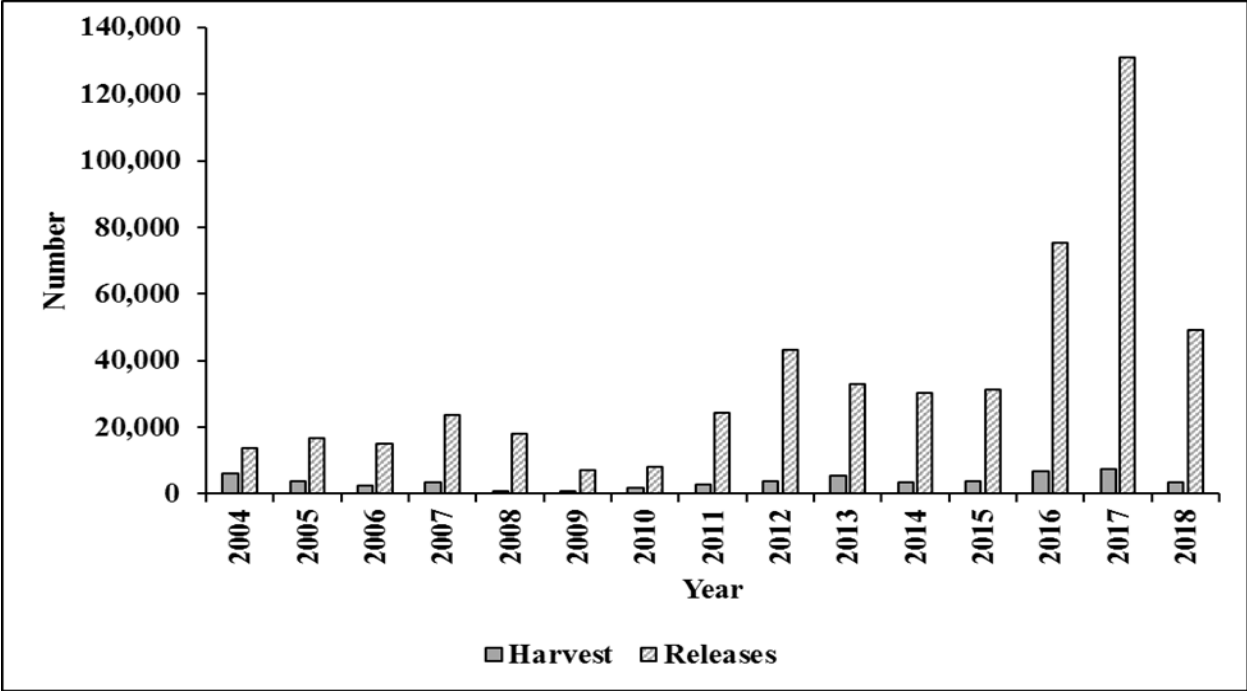
**Figure 2.6.** Program 466 CSMA observer trips by mesh size, 2013–2018. The square is a small mesh observer trip that encountered striped bass ( $n=38$ ), and the circle is a large mesh observer trip that encountered striped bass ( $n=246$ ). Eight large mesh observer trips accounted for 37 striped bass that are not presented on the map due to the absence of coordinates.



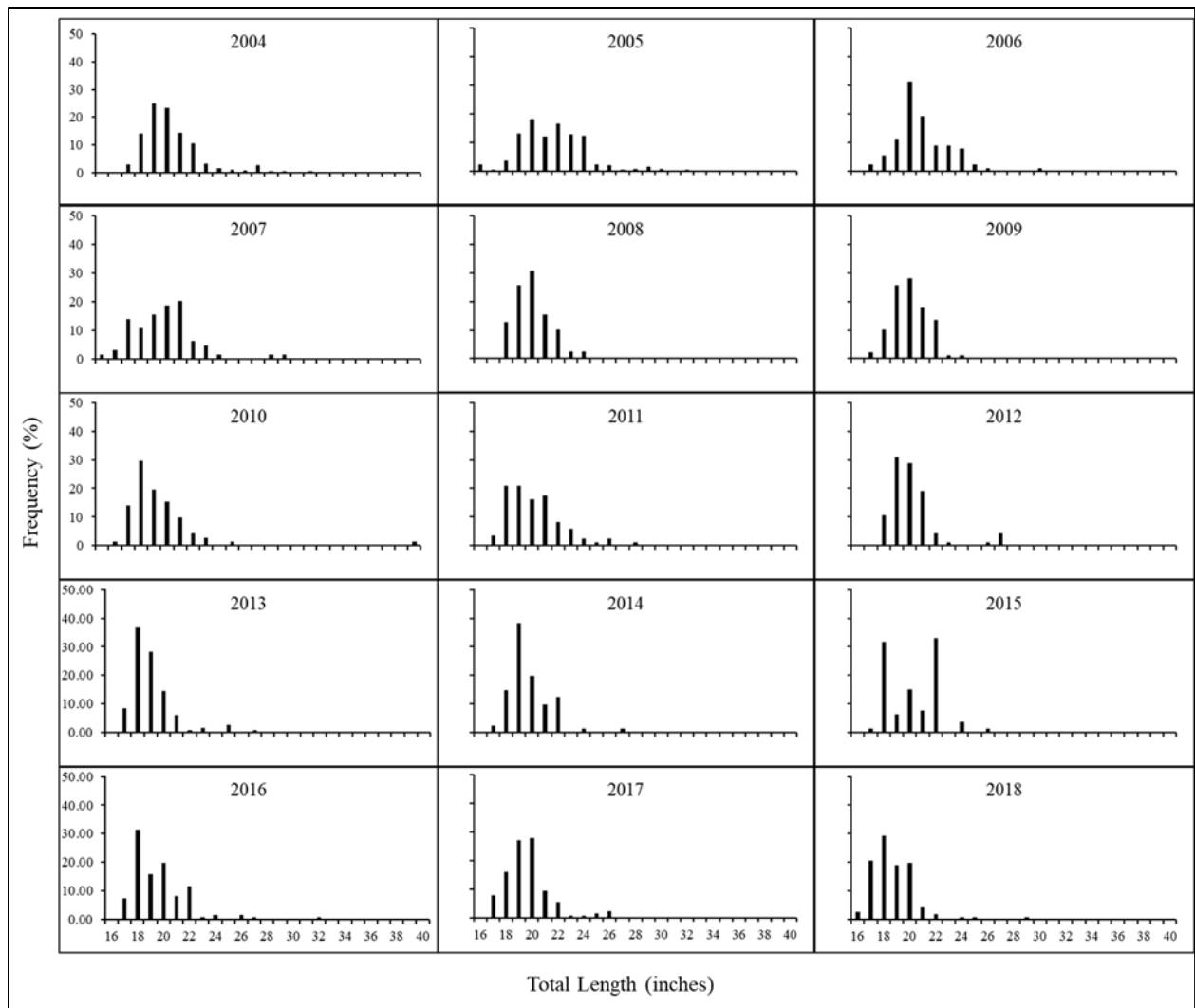
**Figure 2.7.** Recreational striped bass harvest in numbers and pounds and effort in angler hours for the Tar-Pamlico and Neuse rivers and tributaries, 2004–2018.



**Figure 2.8.** Recreational striped bass harvest in the Tar-Pamlico, Pungo, and Neuse rivers, 2004–2018.

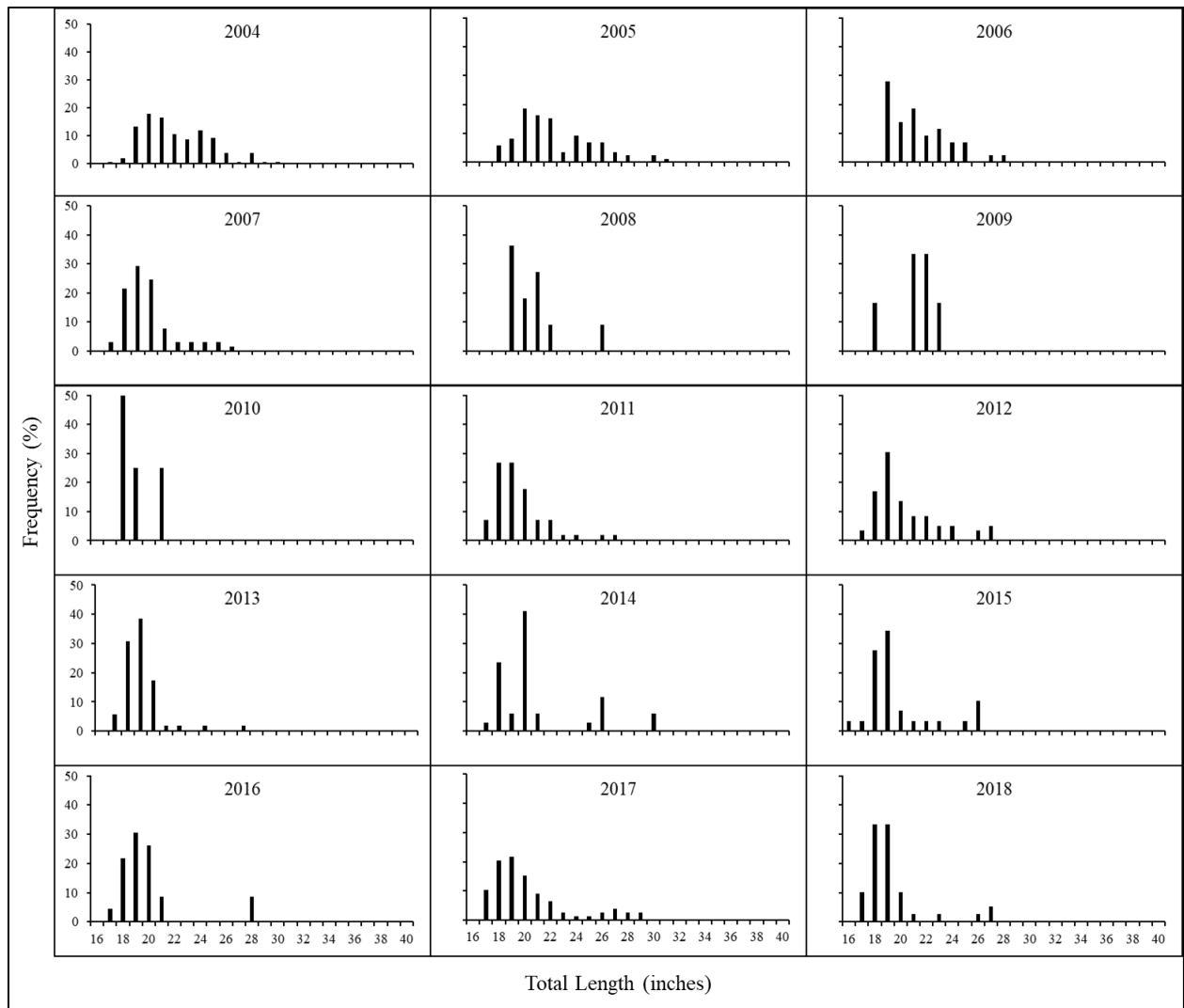


**Figure 2.9.** Annual recreational catch (released and/or harvested) of striped bass in the CSMA, 2004–2018.

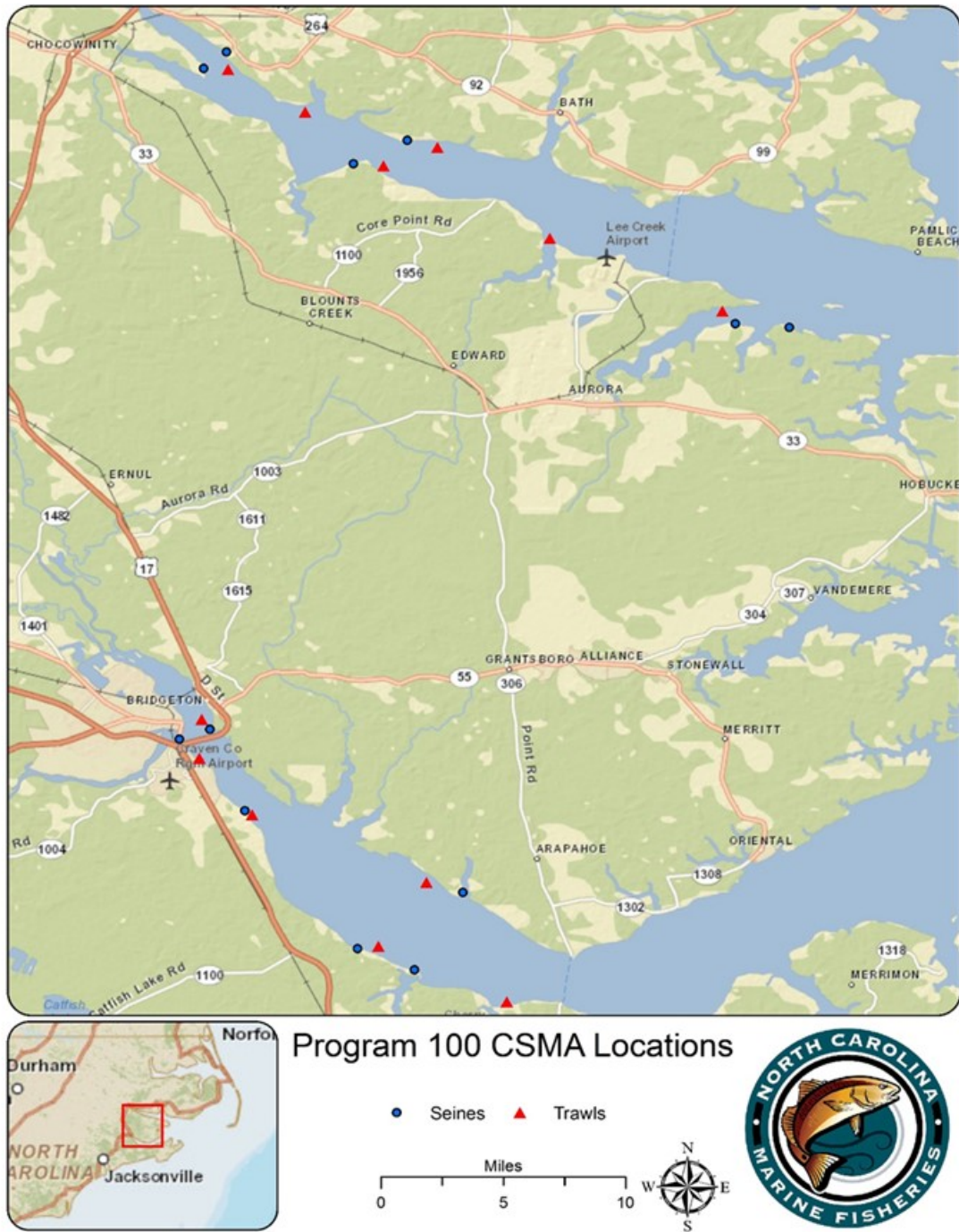


**Figure 2.10.** Length frequency of CSMA striped bass recreationally harvested in the Tar-Pamlico and Pungo rivers, 2004–2018.

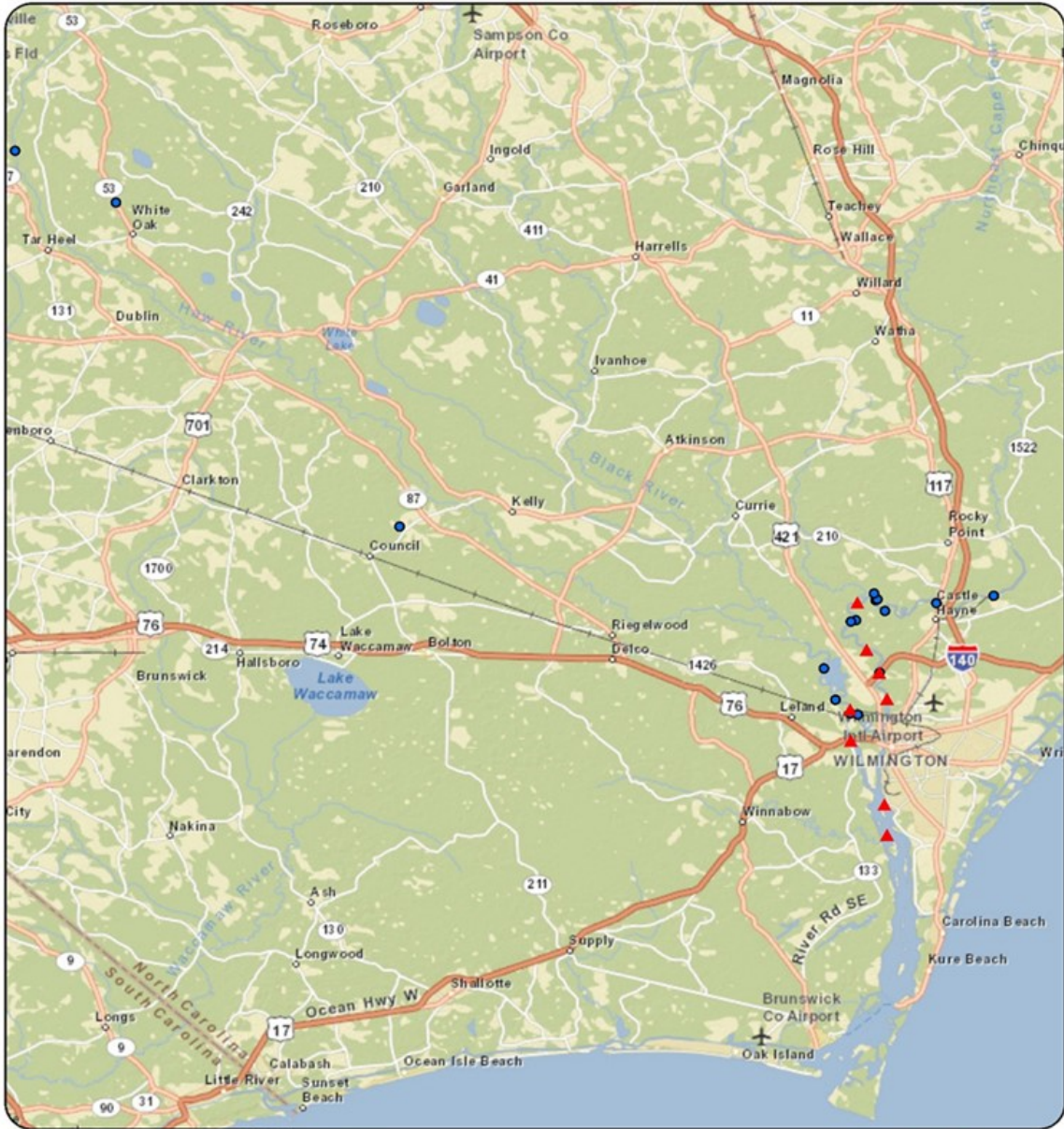




**Figure 2.11.** Length frequency of CSMA striped bass recreationally harvested in the Neuse River, 2004–2018.

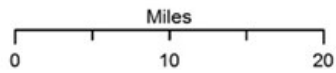


**Figure 2.12.** Location of Central Southern Management Area (CSMA) juvenile striped bass beach seine and trawl sites, Tar-Pamlico and Neuse rivers, NC.

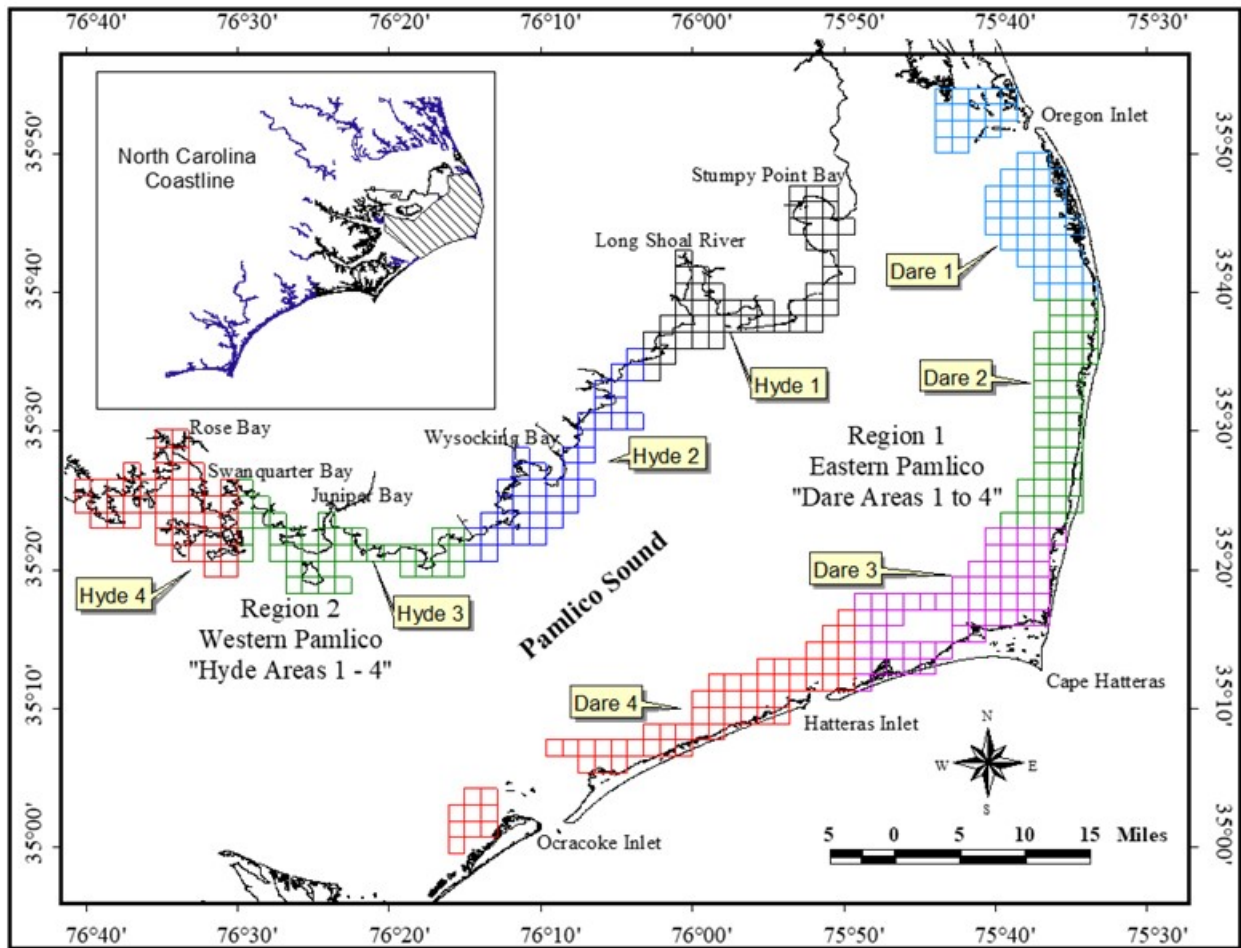


### Program 100 CFR Locations

● Seines ▲ Trawls

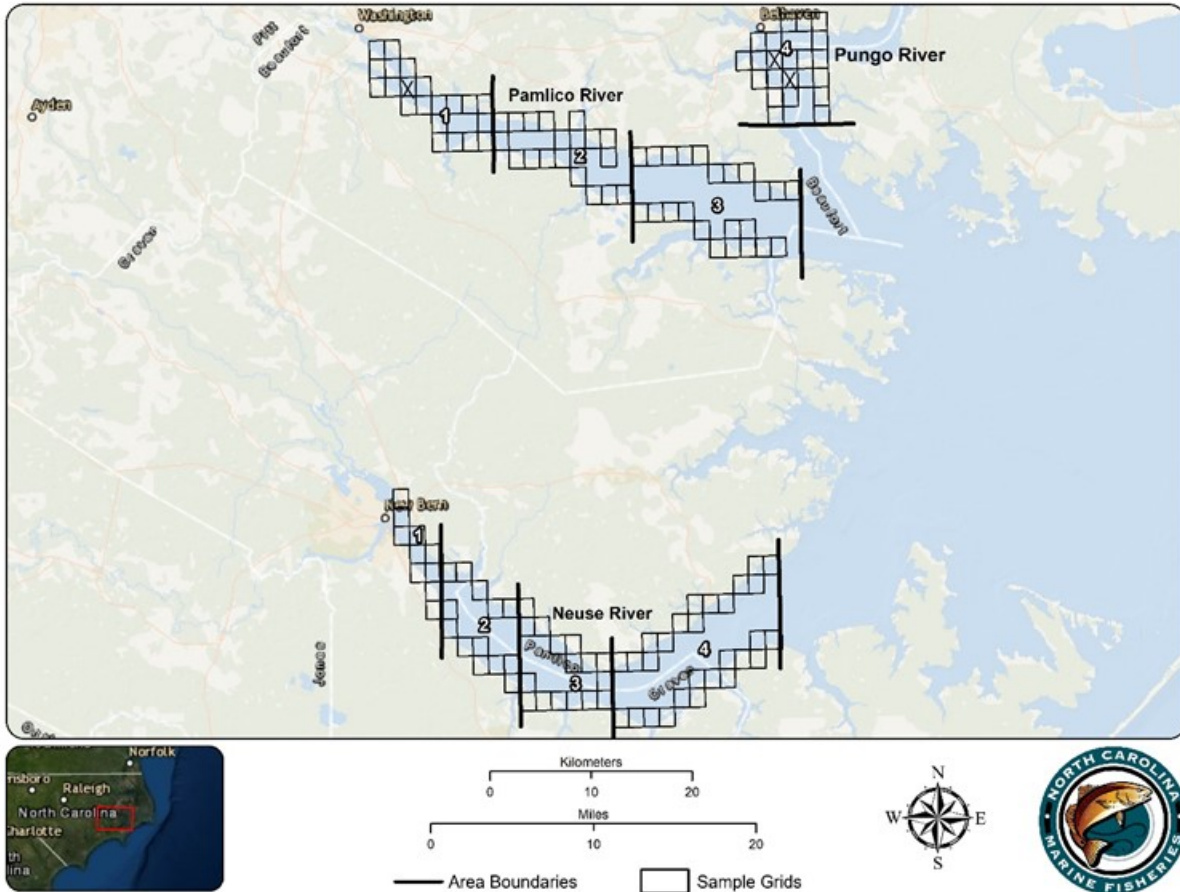


**Figure 2.13.** Location of Cape Fear River juvenile striped bass beach seine and trawl sites, Cape Fear River, NC.



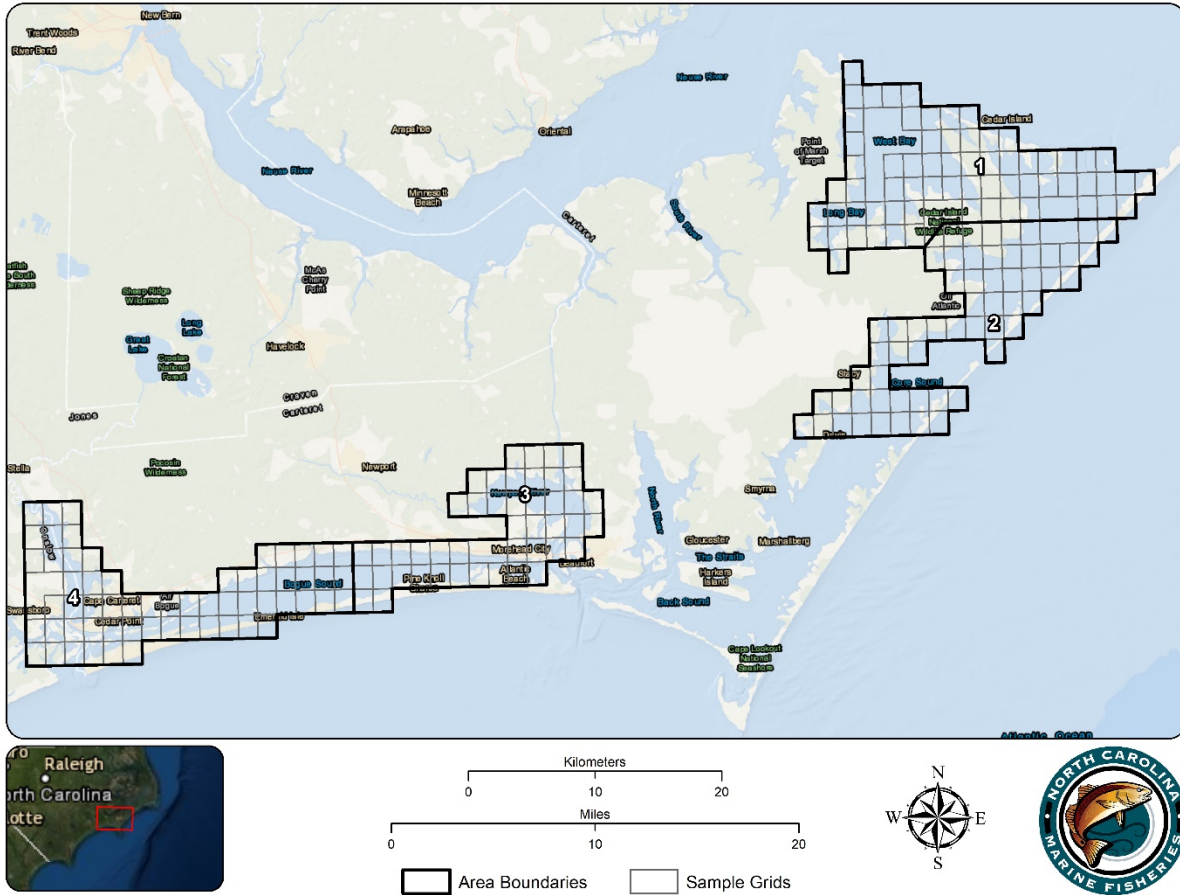
**Figure 2.14.** The sample regions and grid system for P915 in Dare (Region 1) and Hyde (Region 2) counties.

Program 915 Pamlico Region Area Map



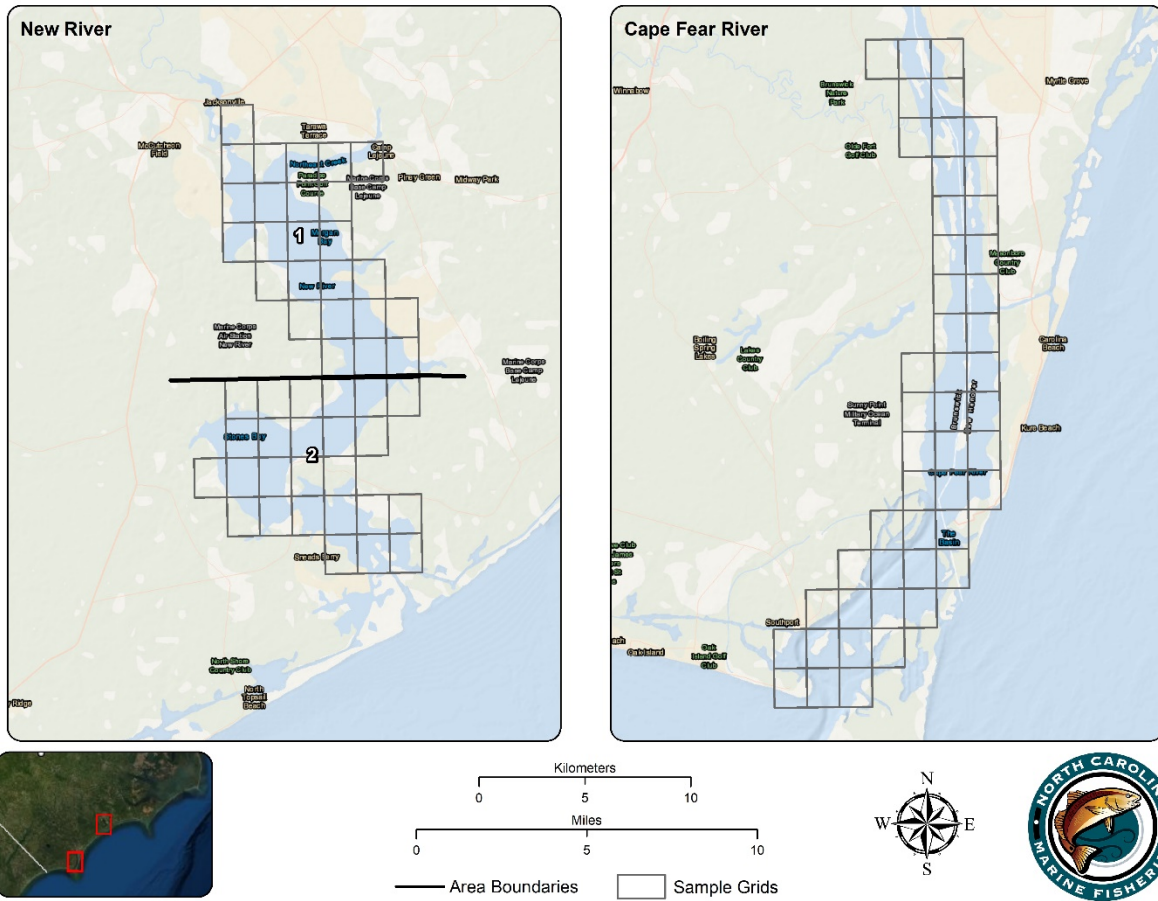
**Figure 2.15.** The sample areas and grid system for P915 in the Pamlico Region (Pamlico, Pungo and Neuse rivers) with areas numbered Pamlico/Pungo: 1—Upper, 2—Middle, 3—Lower, 4—Pungo; Neuse: 1—Upper, 2—Upper-middle, 3—Lower-middle, and 4—Lower).

### Program 915 Central Region Area Map

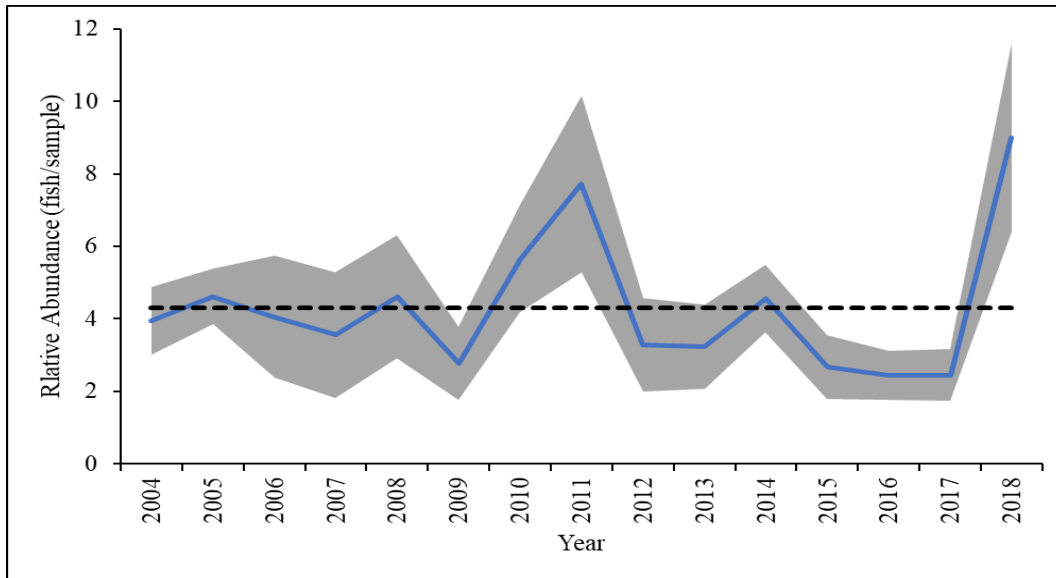


**Figure 2.16.** The sample areas and grid system for P915 in the Central Region with areas numbered (1—West Bay/Upper Core Sound, 2—Lower Core Sound, 3—Newport River/Bogue Sound, and 4—Bogue Sound/White Oak River). Sampling began May 2018.

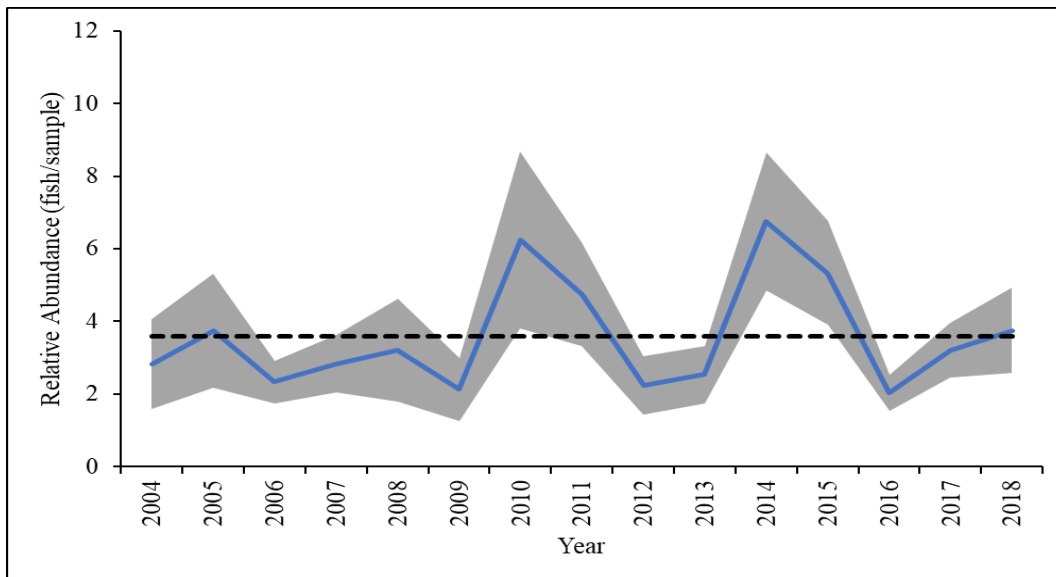
## Program 915 Southern Region Area Map



**Figure 2.17.** The sample areas and grid system for P915 in the Southern Region (New and Cape Fear rivers) with areas numbered (New: 1—Upper, 2—Lower, Cape Fear).

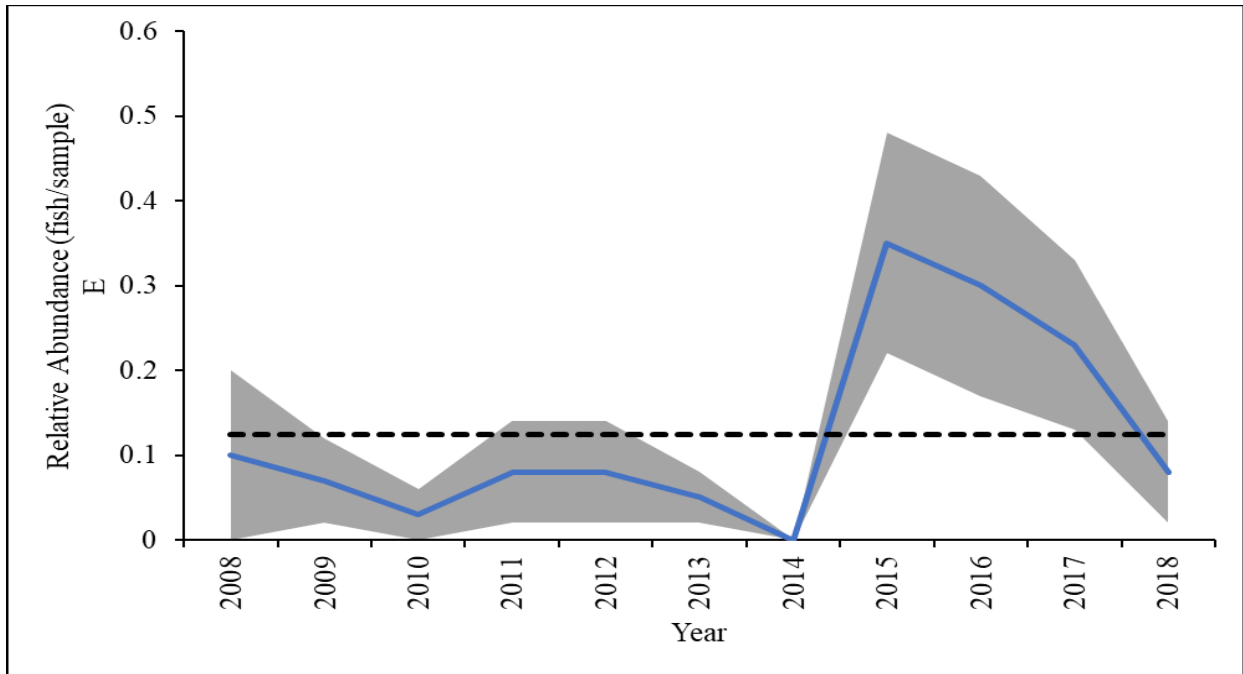


**Figure 2.18.** Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2004–2018 (Tar-Pamlico River, shallow sets, April and October–November). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index.

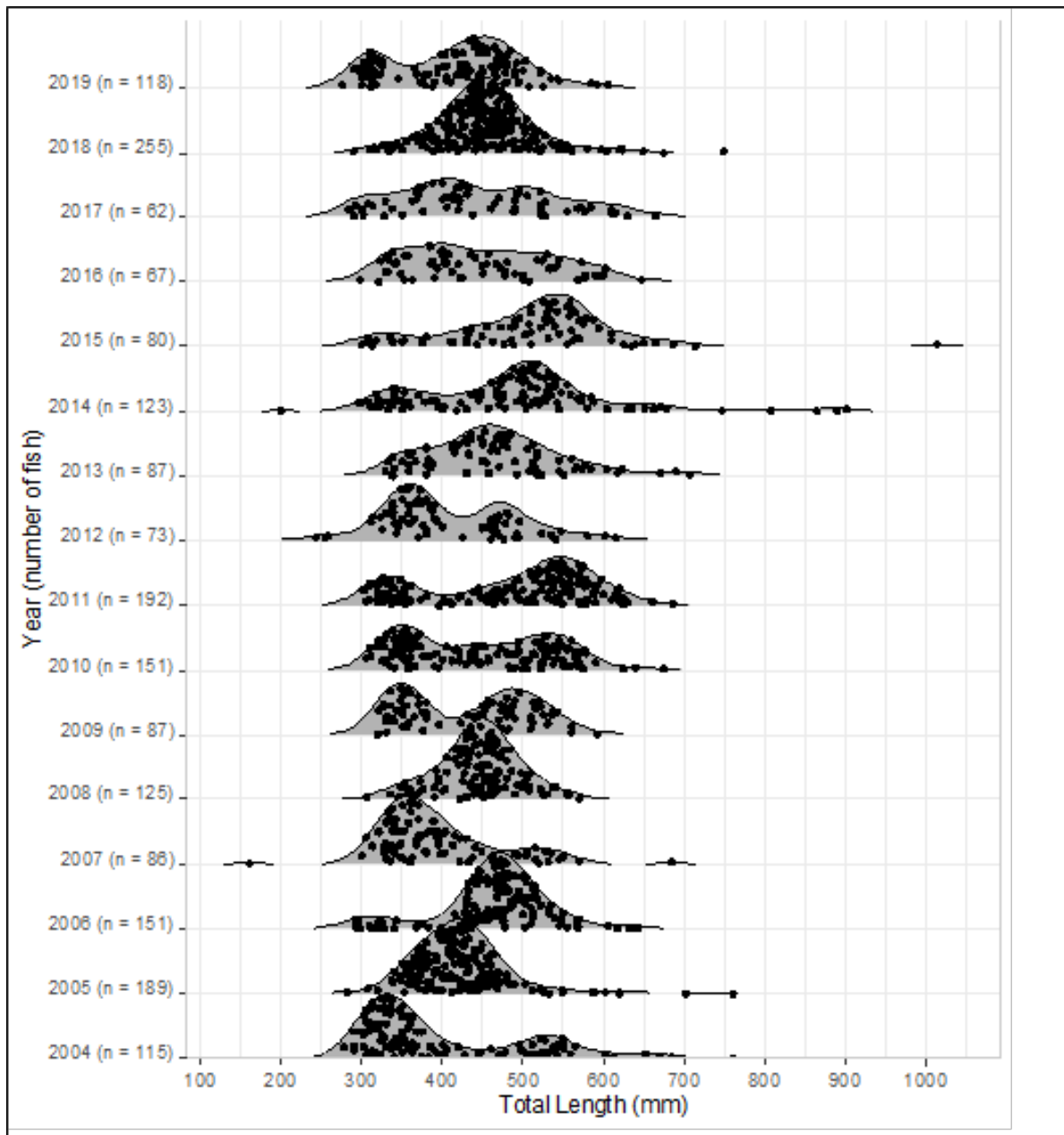


**Figure 2.19.** Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2004–2018 (Neuse River, shallow sets, April and October–November). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index.

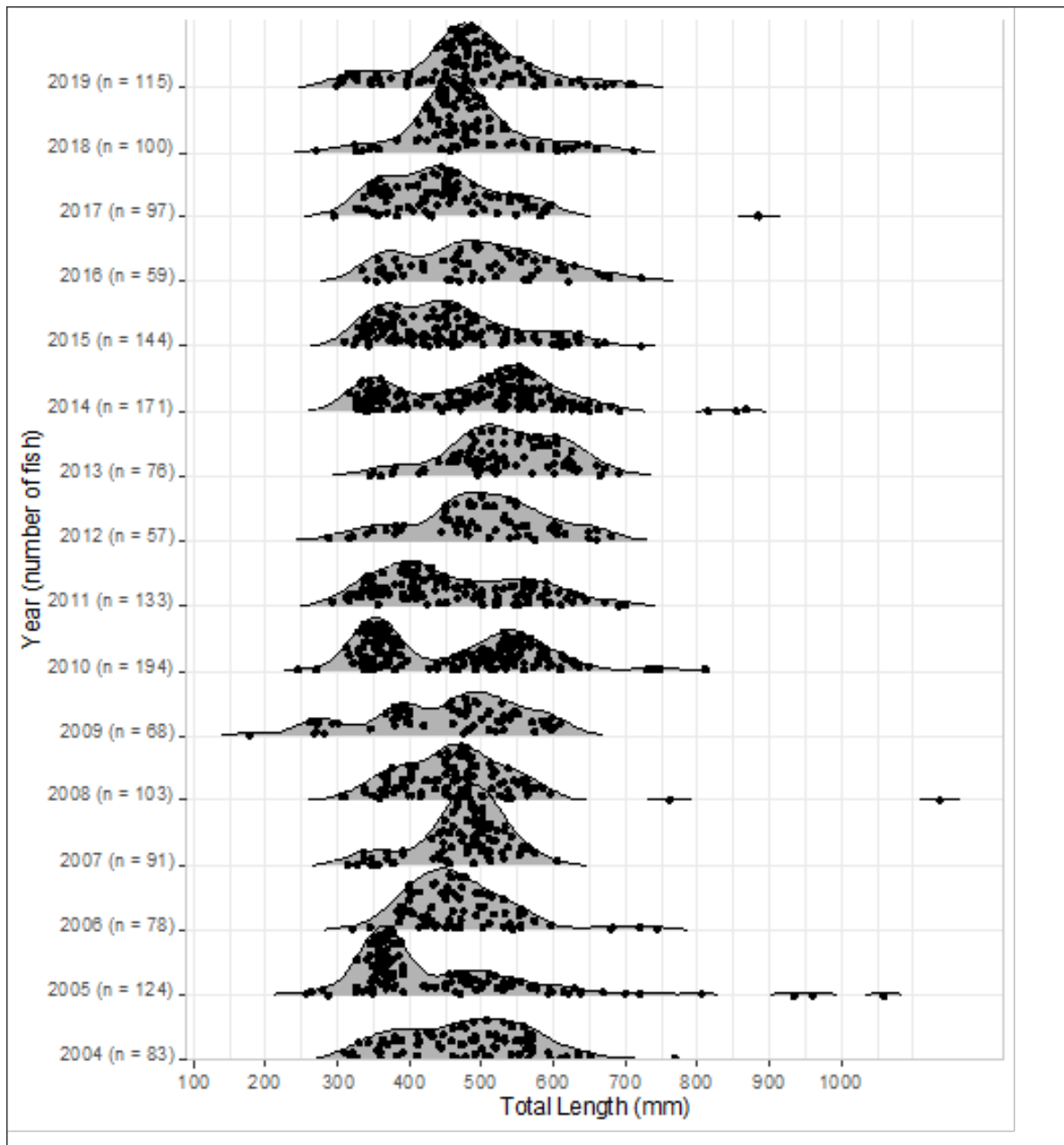




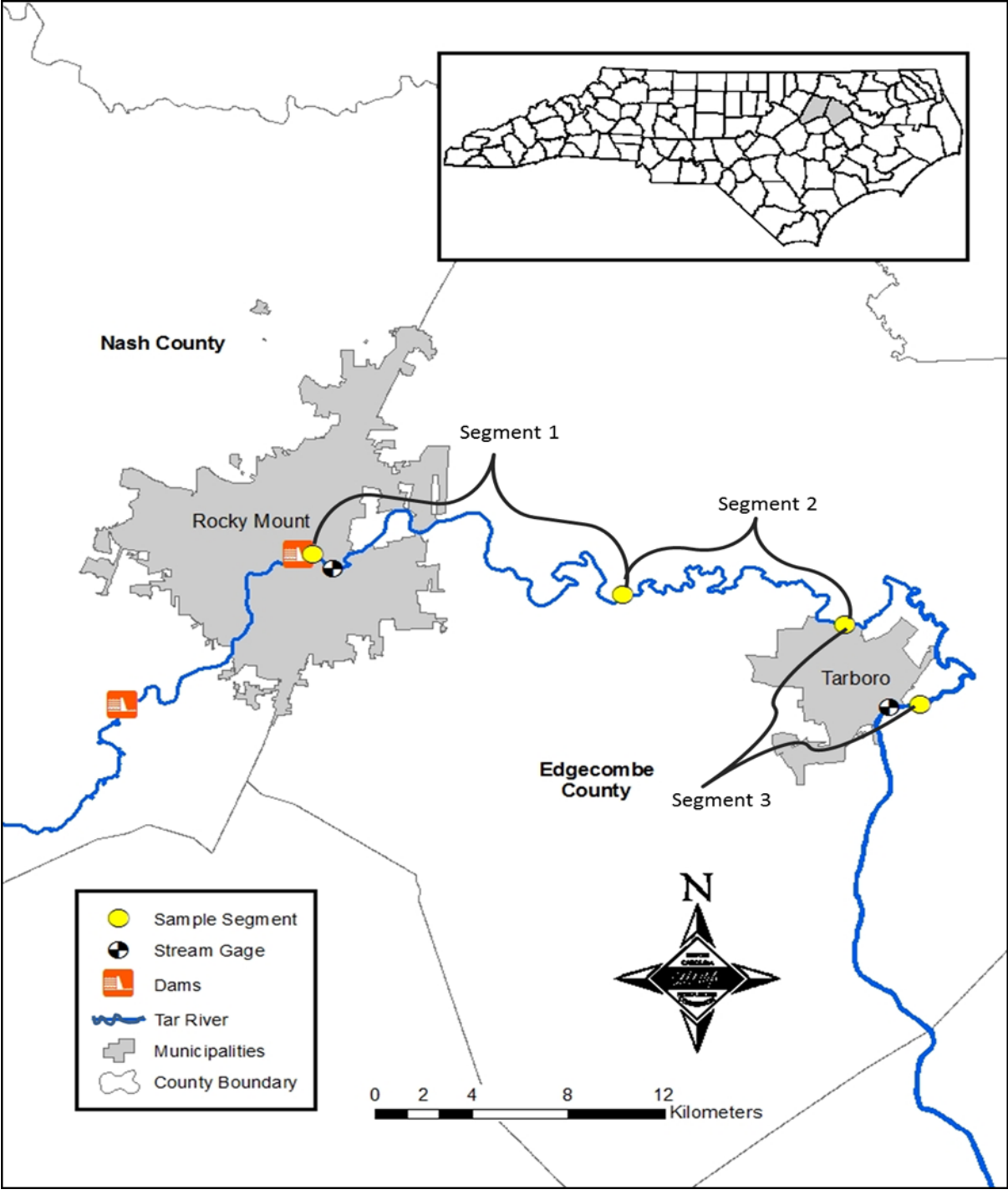
**Figure 2.20.** Striped bass annual weighted relative abundance index (# fish per sample; sample=240 yards of gill net) in P915, 2008–2018 (Cape Fear River, shallow sets). Dashed black line represents time-series average. Shaded area represents standard error. Soak times were not used in calculating the index.



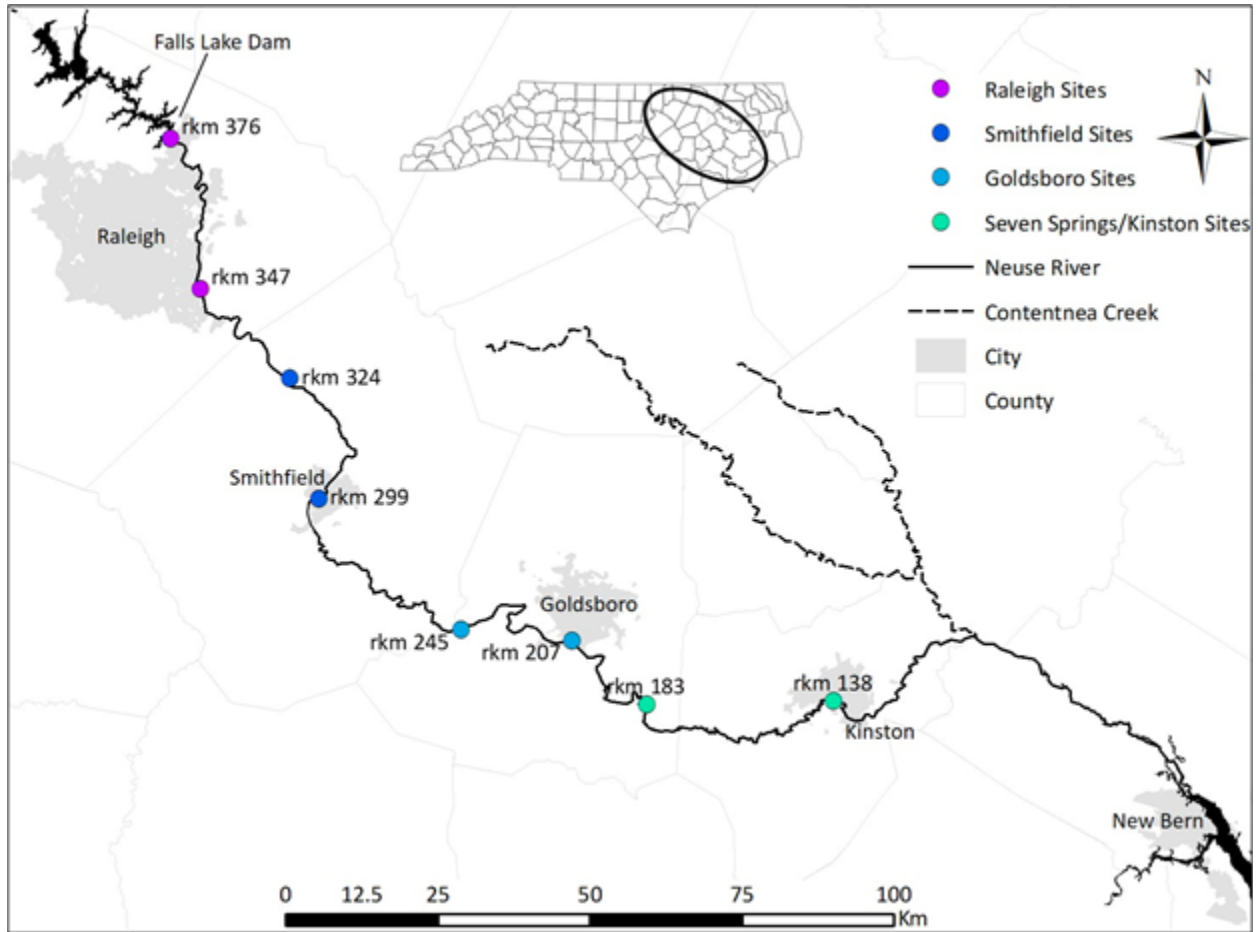
**Figure 2.21.** Length frequency distribution of CSMA striped bass captured in P915 in the Tar-Pamlico River, 2004–2019 (deep and shallow sets, April and October–November).



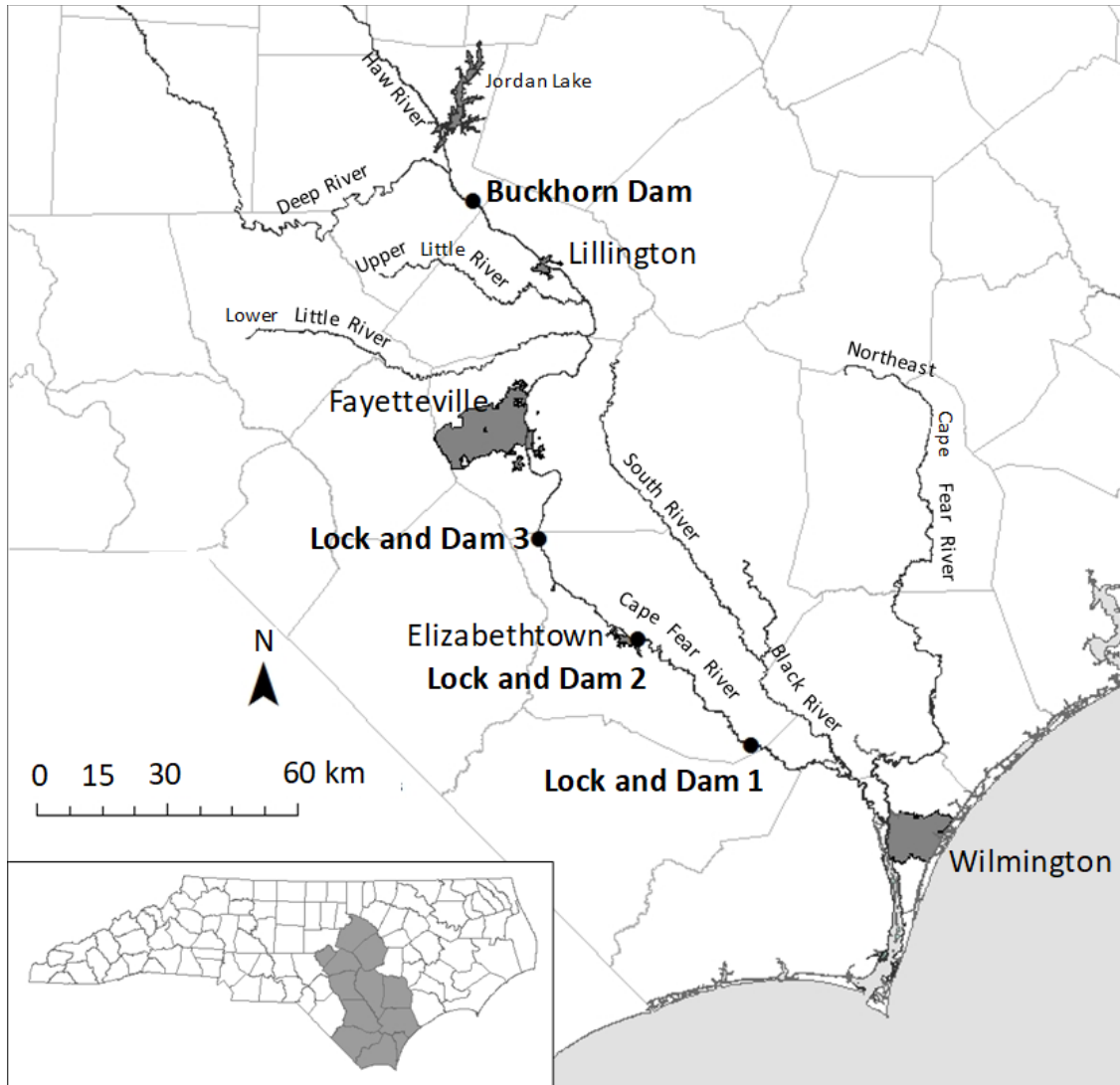
**Figure 2.22.** Length frequency distribution of CSMA striped bass captured in P915 the Neuse River, 2004–2019 (deep and shallow sets, April and October–November).



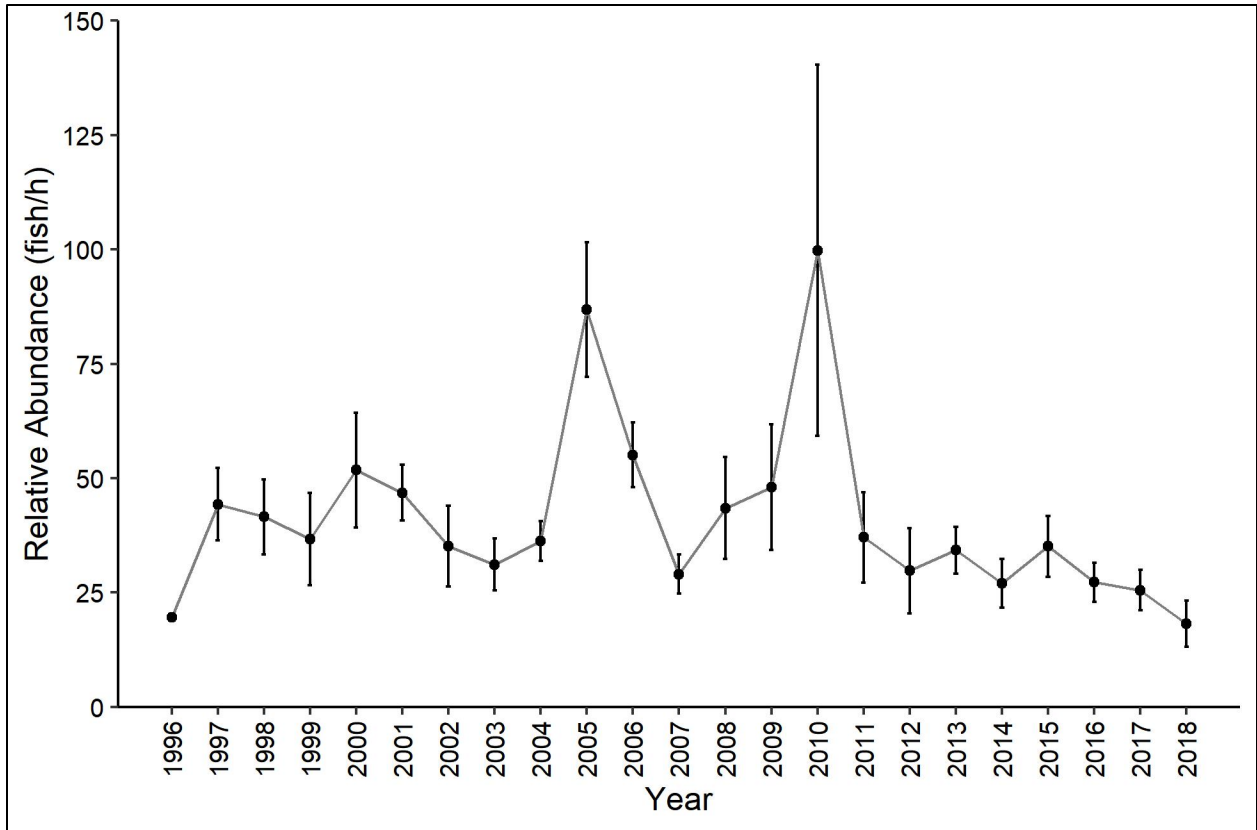
**Figure 2.23.** NCWRC electrofishing survey segments on the Tar-Pamlico River.



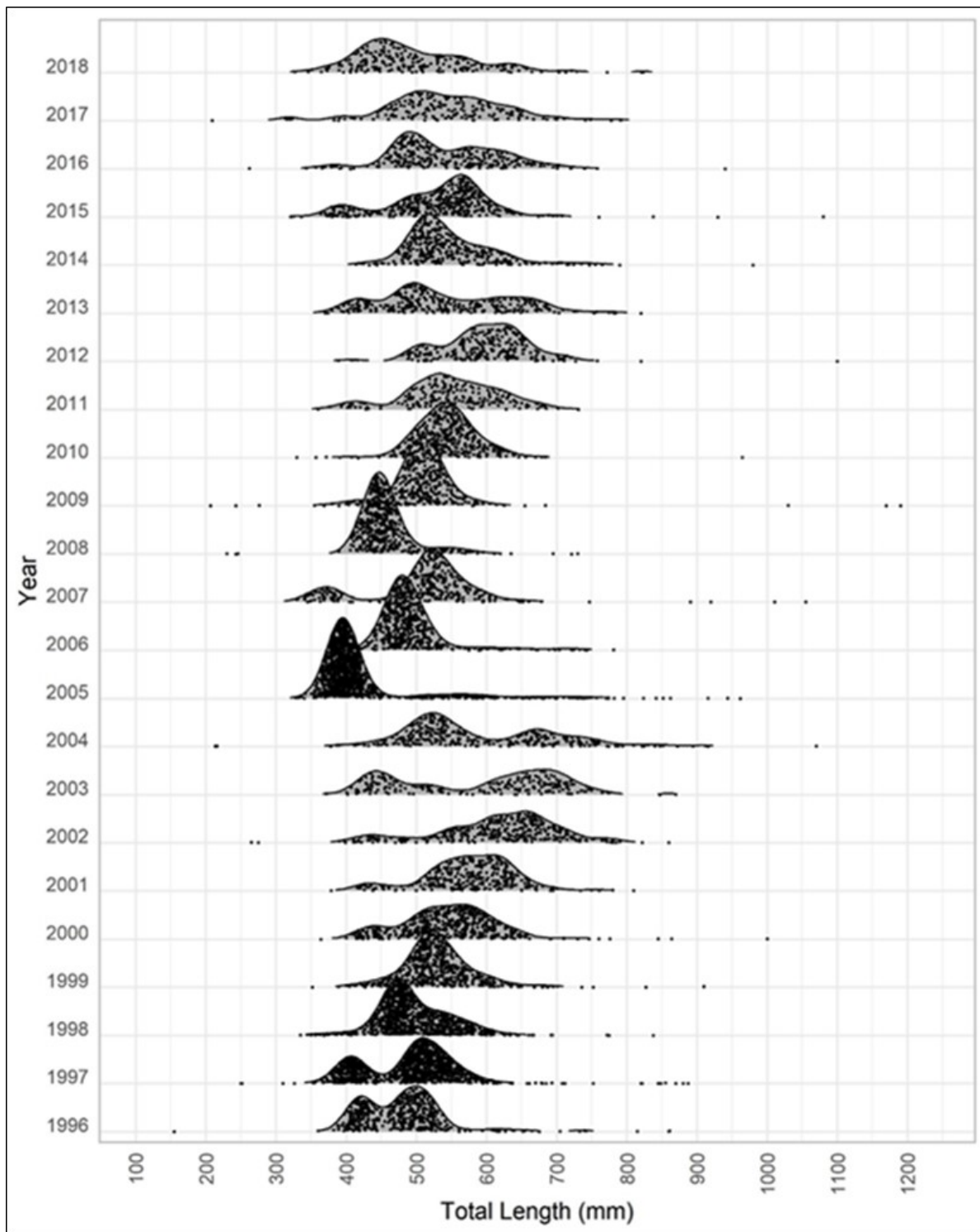
**Figure 2.24.** NCWRC electrofishing survey area on the Neuse River. The upstream and downstream extent of four sampling strata are by colored markers.



**Figure 2.25.** NCWRC electrofishing sampling sites (indicated by black circles in bold) at Lock and Dams 1, 2, 3, and Buckhorn Dam on the Cape Fear River.

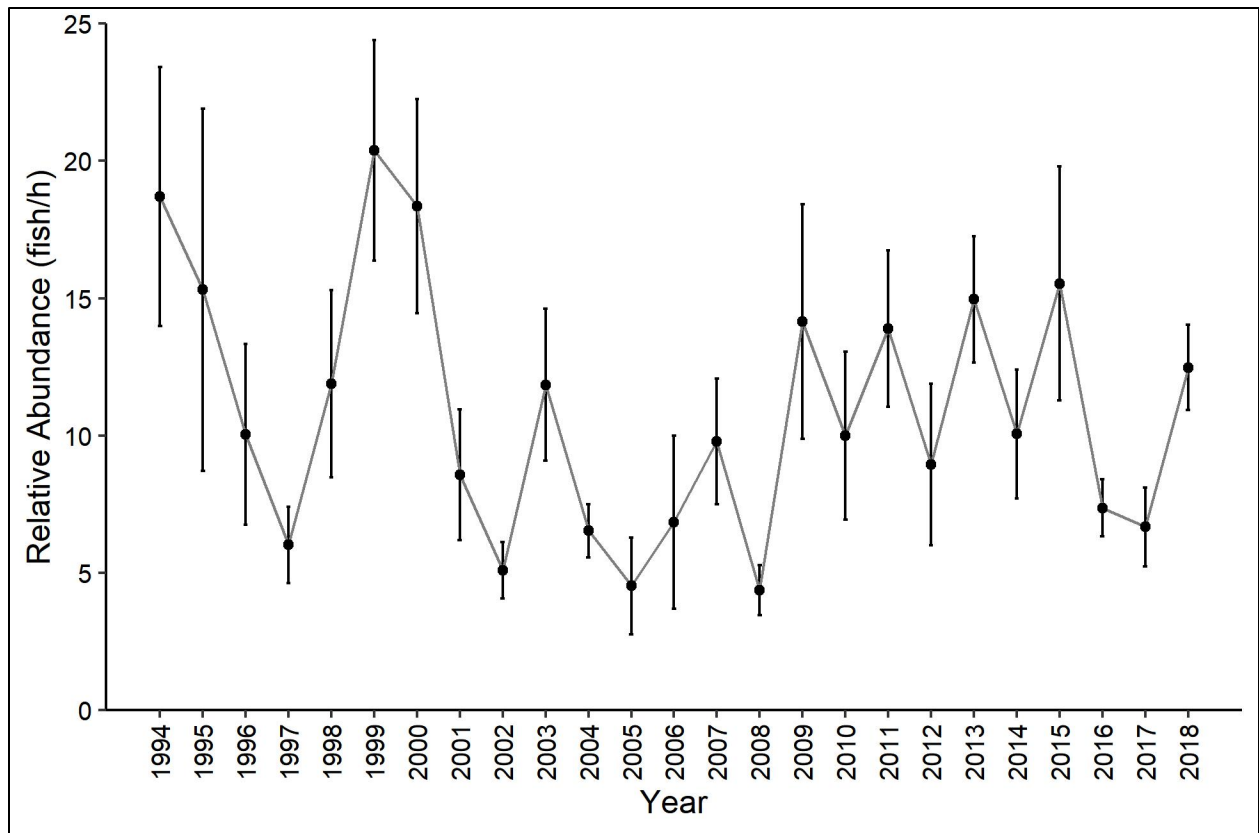


**Figure 2.26.** Relative abundance (with associated standard error) of striped bass collected during the NCWRC Tar River electrofishing surveys, 1996–2018.

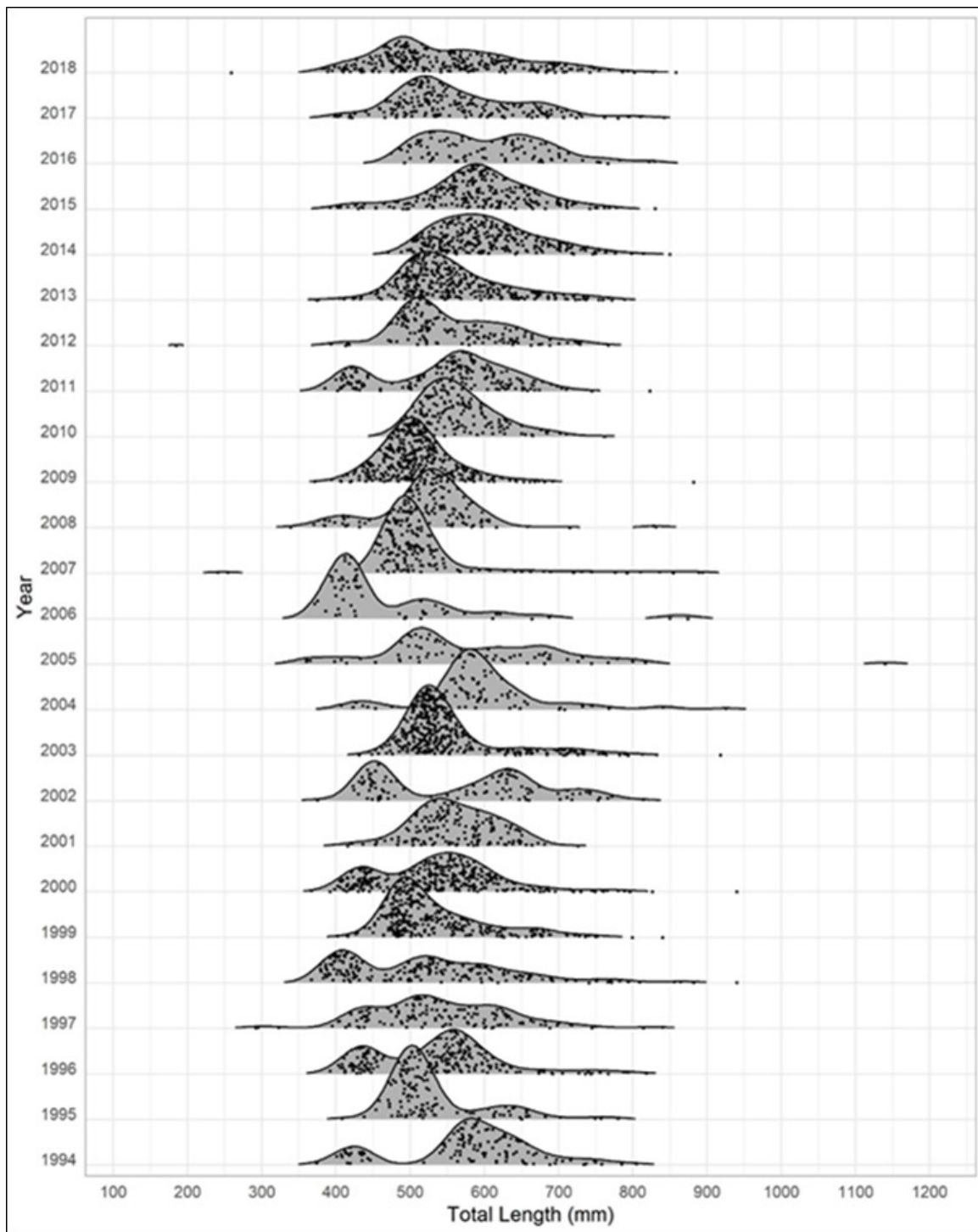


**Figure 2.27.** Length distributions for striped bass collected during the NCWRC Tar River electrofishing surveys, 1996–2018. Dots indicate individual length measurements.

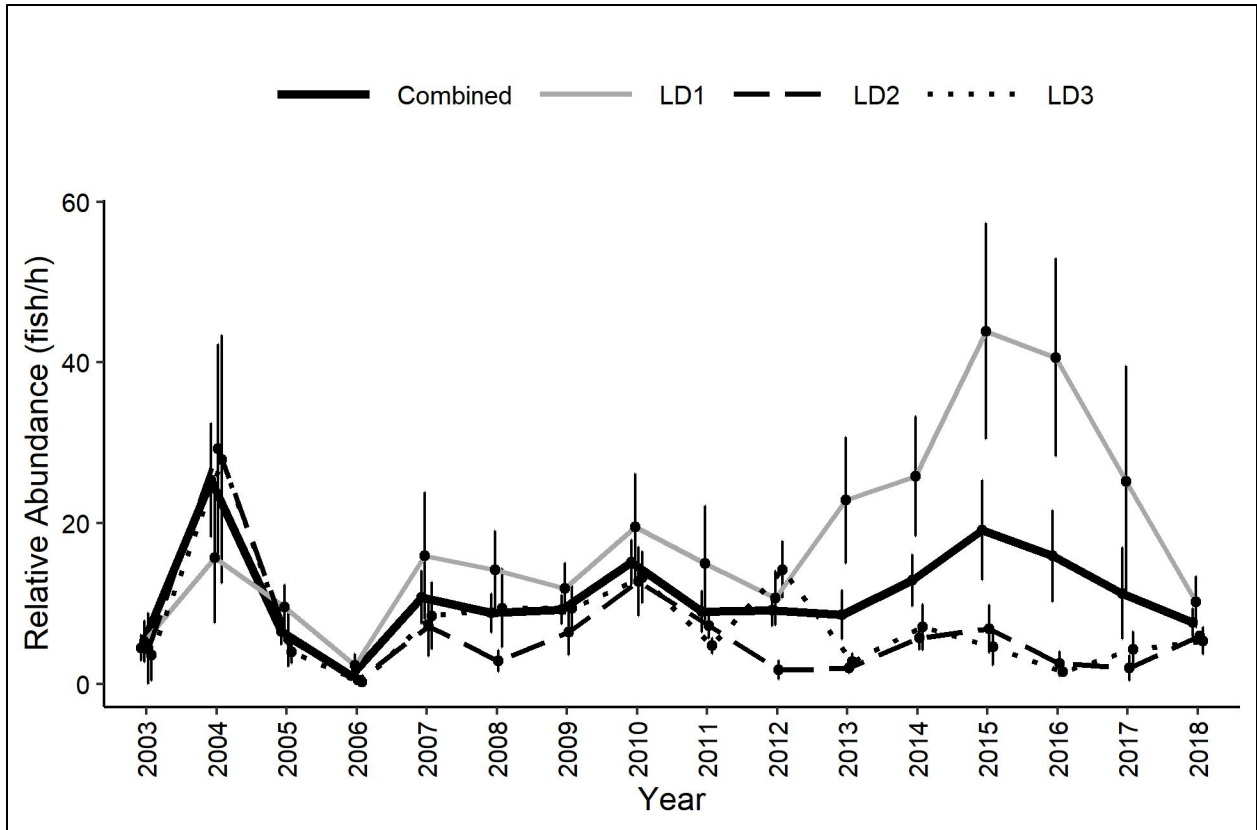




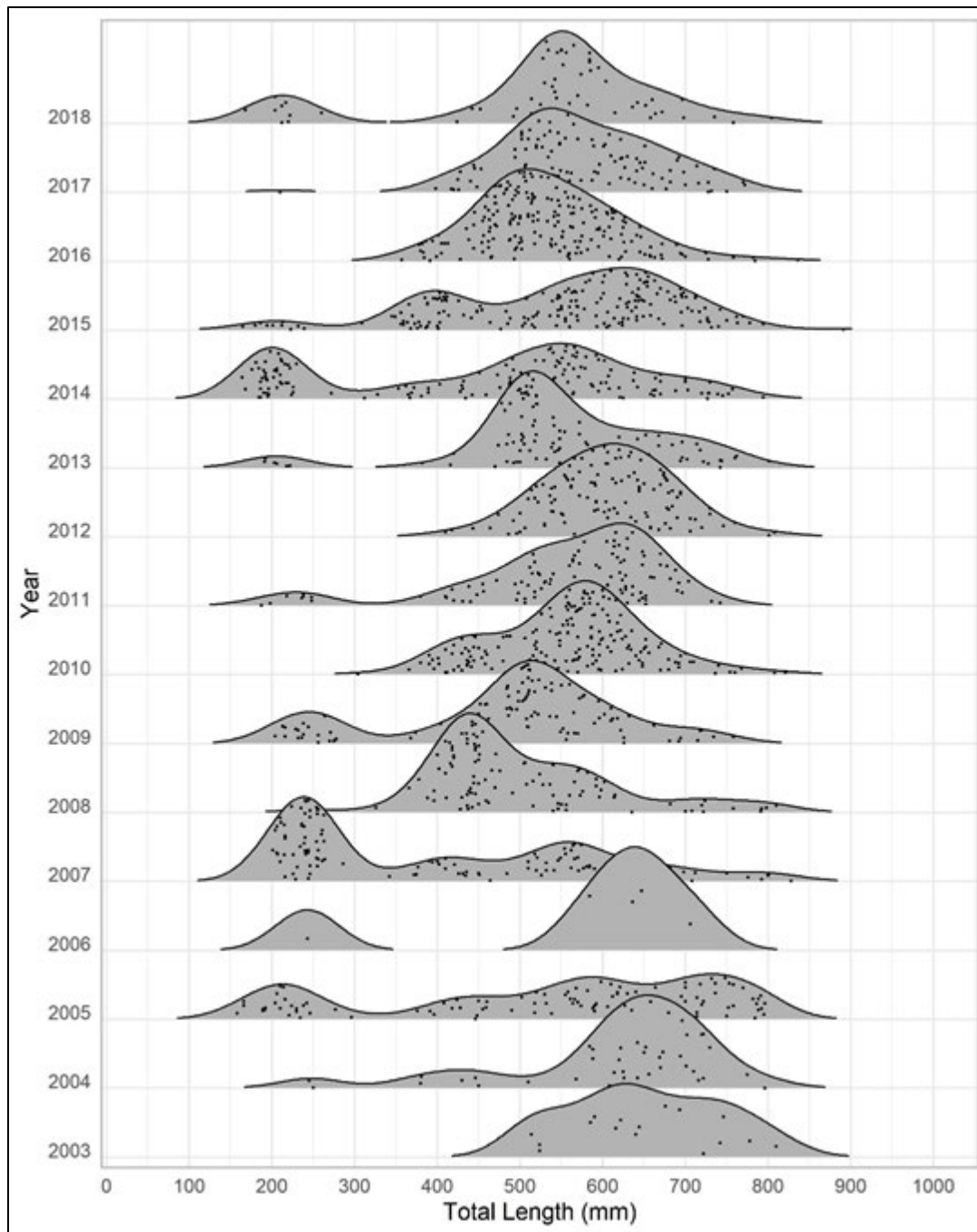
**Figure 2.28.** Relative abundance (with associated standard error) of striped bass collected during the NCWRC Neuse River electrofishing surveys, 1994–2018.



**Figure 2.29.** Striped bass length distributions for the NCWRC Neuse River electrofishing surveys, 1994–2018. Dots indicate individual length measurements.



**Figure 2.30.** Relative abundance (with associated standard error) of striped bass collected at three sample sites in the Cape Fear River, NC, 2003–2018.



**Figure 2.31.** Length distributions for striped bass collected during the NCWRC Cape Fear River electrofishing surveys, 2003–2018. Dots indicate individual length measurements.

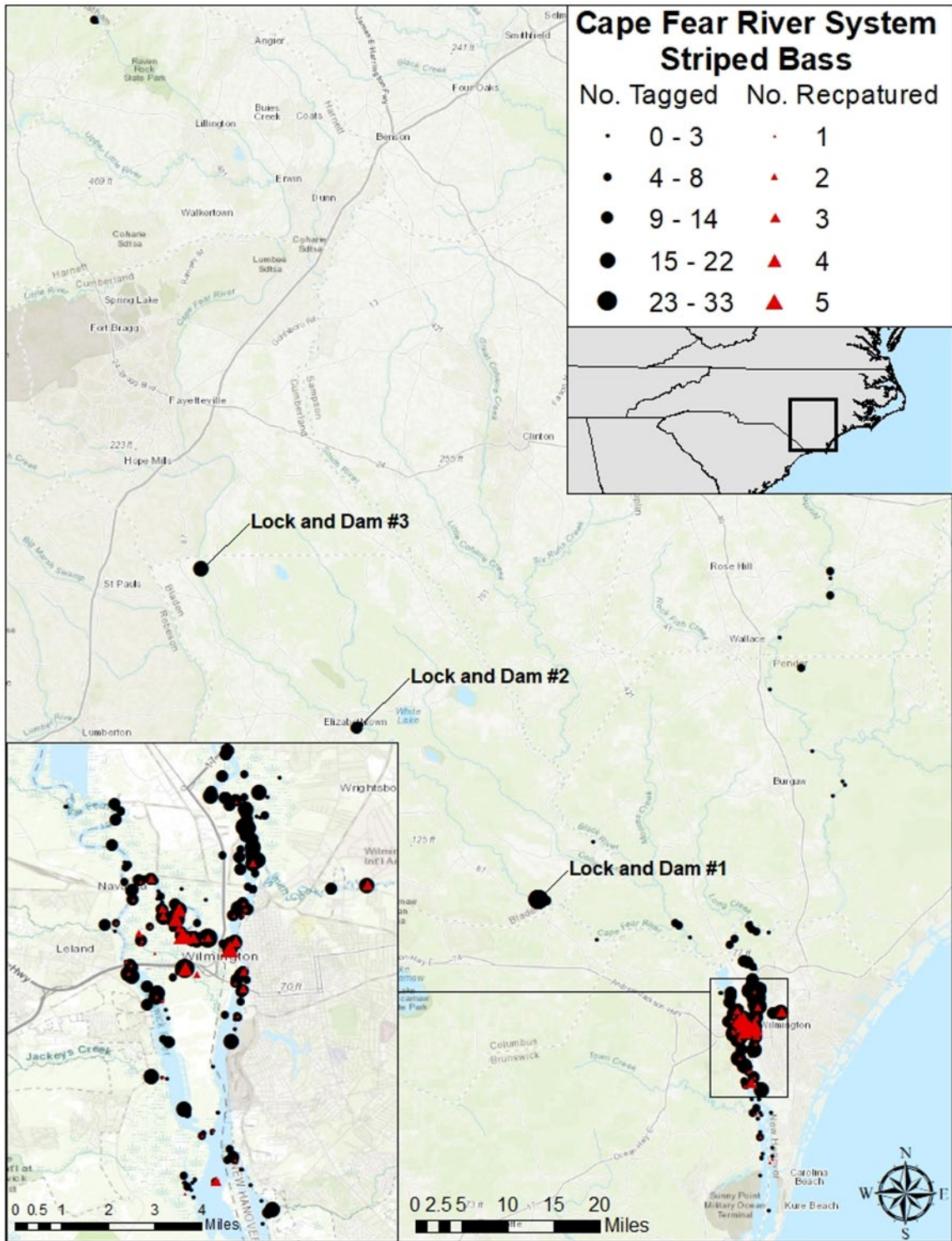
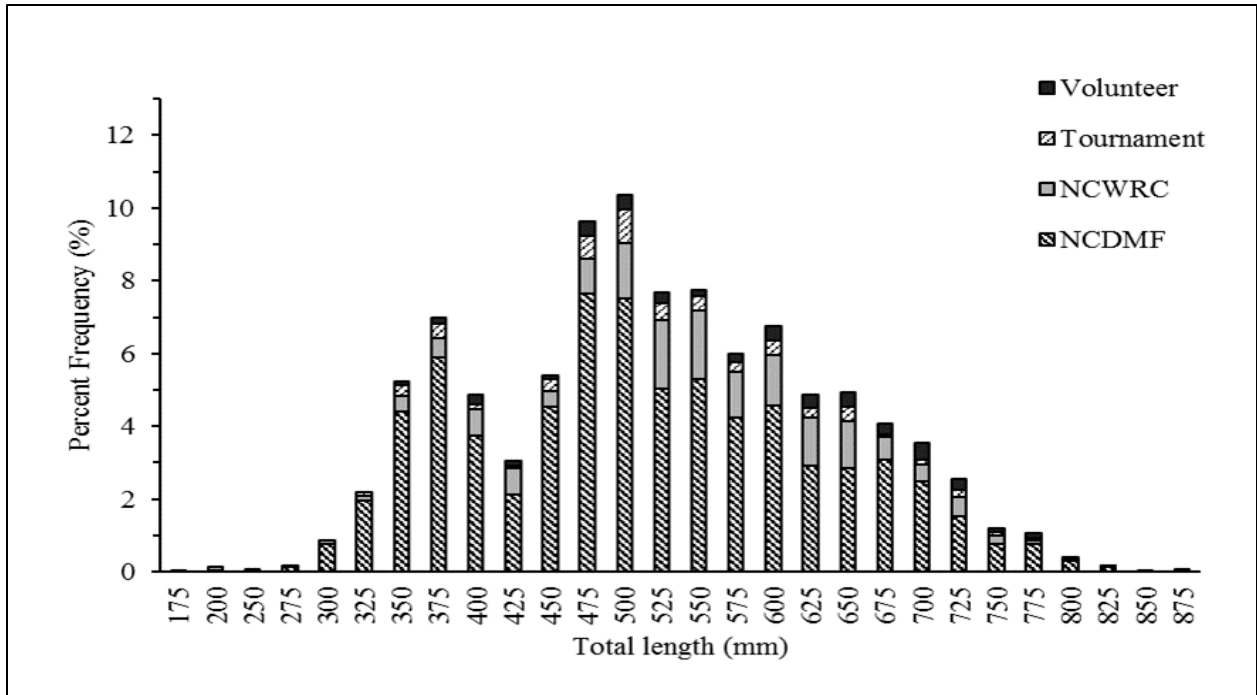
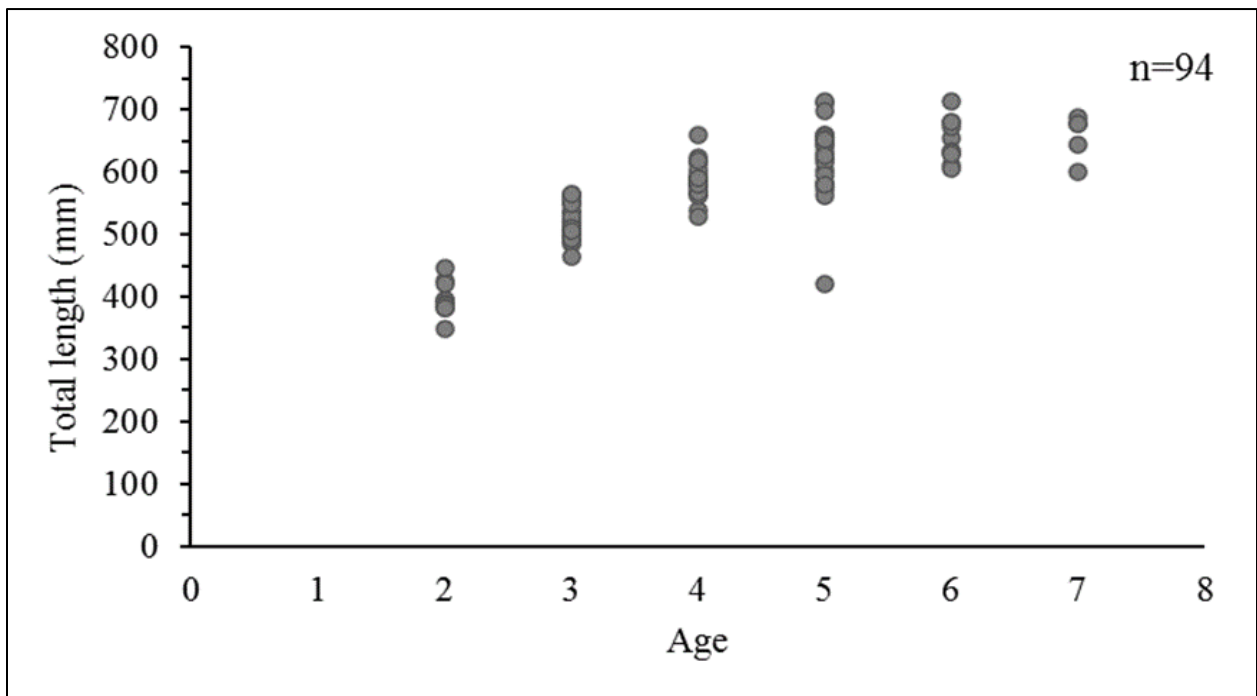


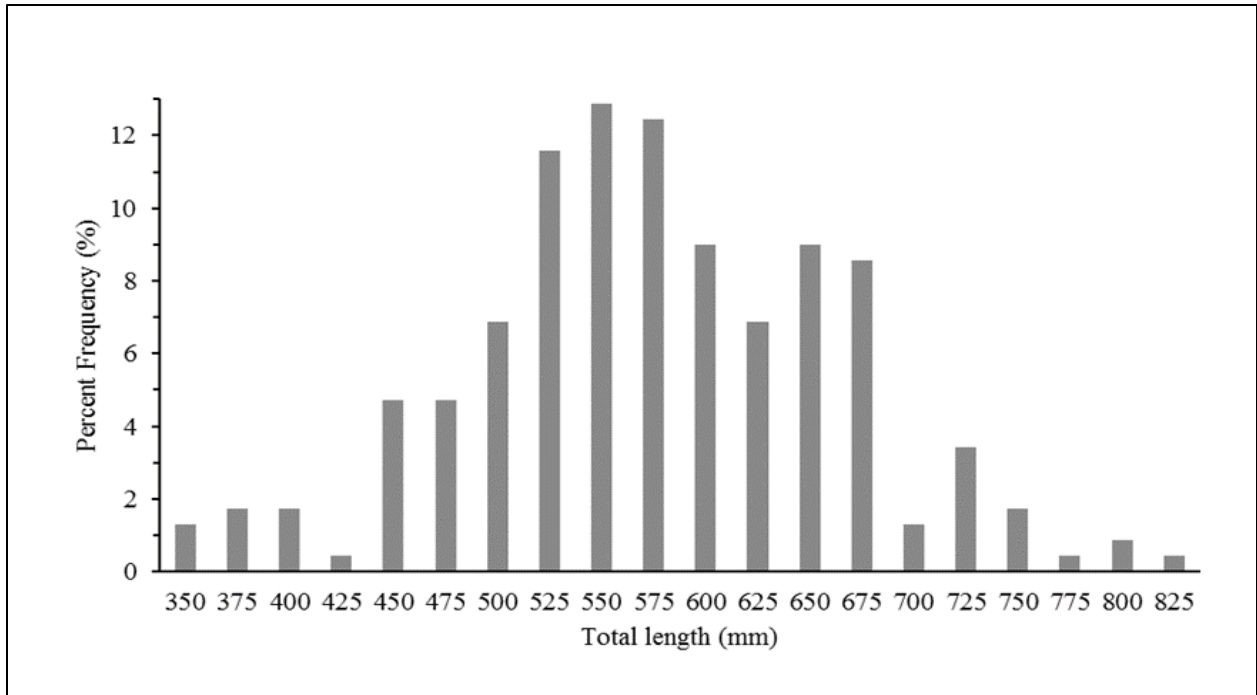
Figure 2.32. Cape Fear River striped bass tagging and recapture locations, 2012–2018.



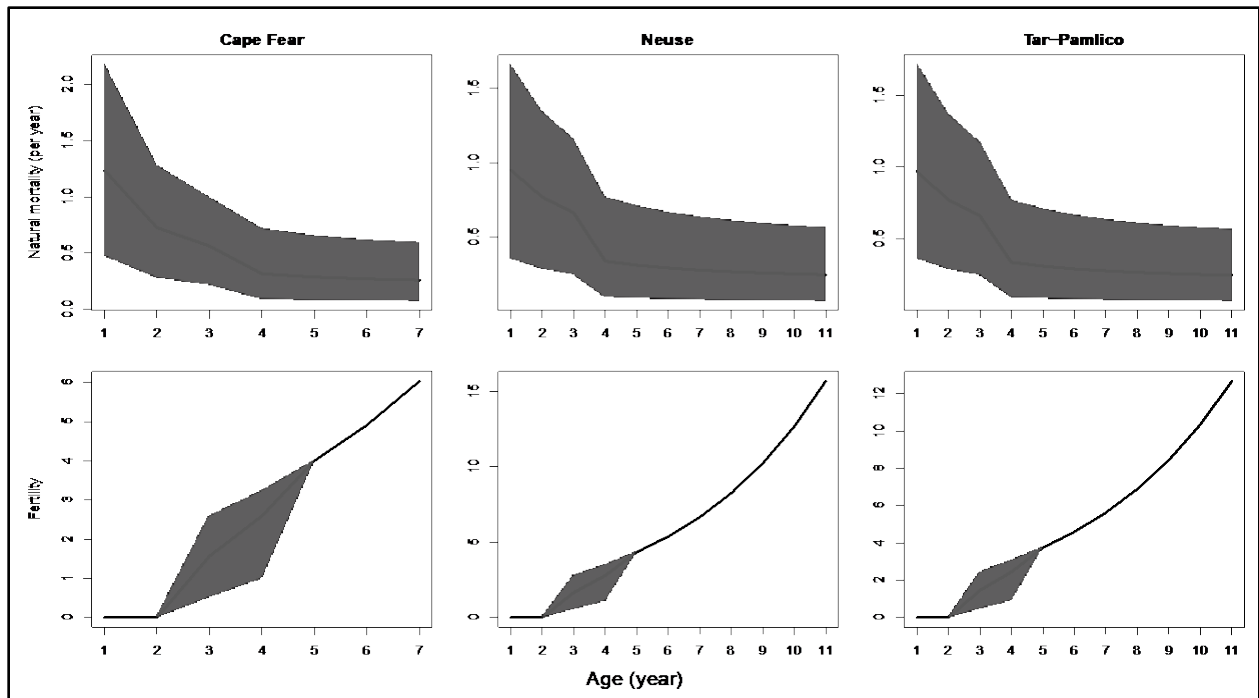
**Figure 2.33.** Length-frequency distribution of tagged striped bass included in the tagging model by tagger affiliation in the Cape Fear River, 2012–2018.



**Figure 2.34.** Genetically derived age at length of Cape Fear River striped bass, 2016–2017.

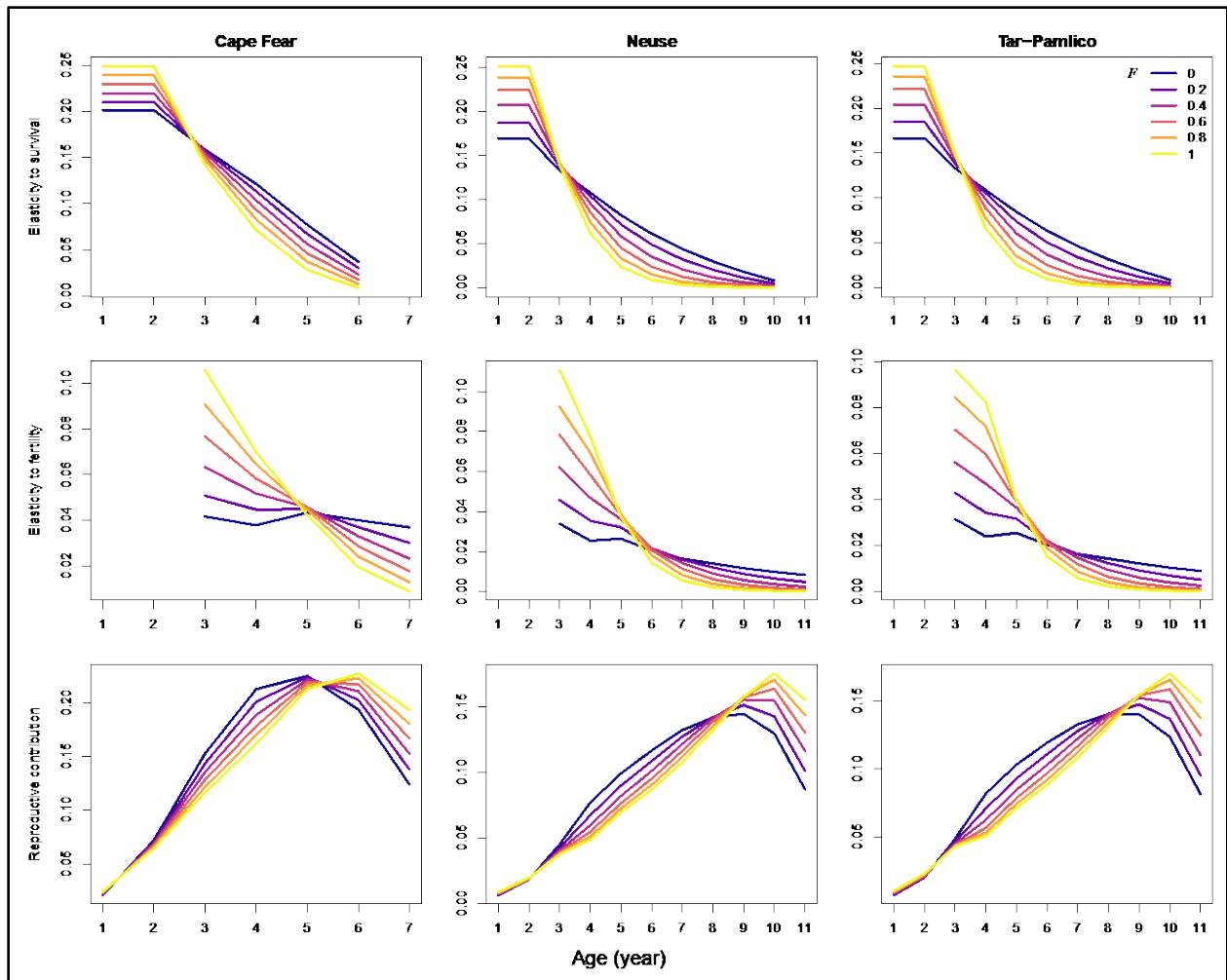


**Figure 2.35.** Length-frequency distribution of recaptured striped bass included in the tagging model by tagger affiliation in the Cape Fear River, 2012–2018.

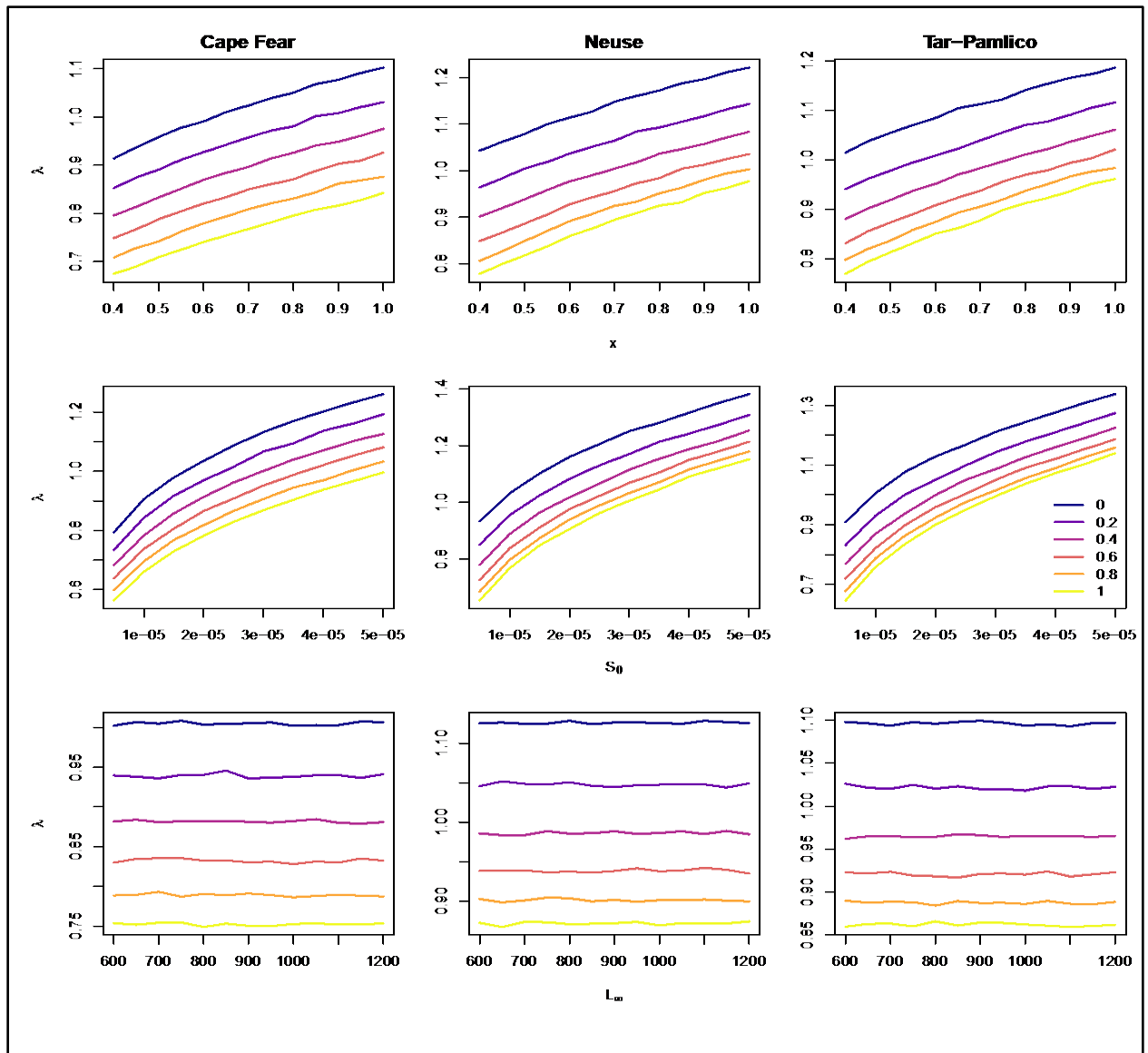


**Figure 3.1.** Age-specific natural mortality and fertility used in the matrix model. Black line is median and grey area is 95% confidence interval.

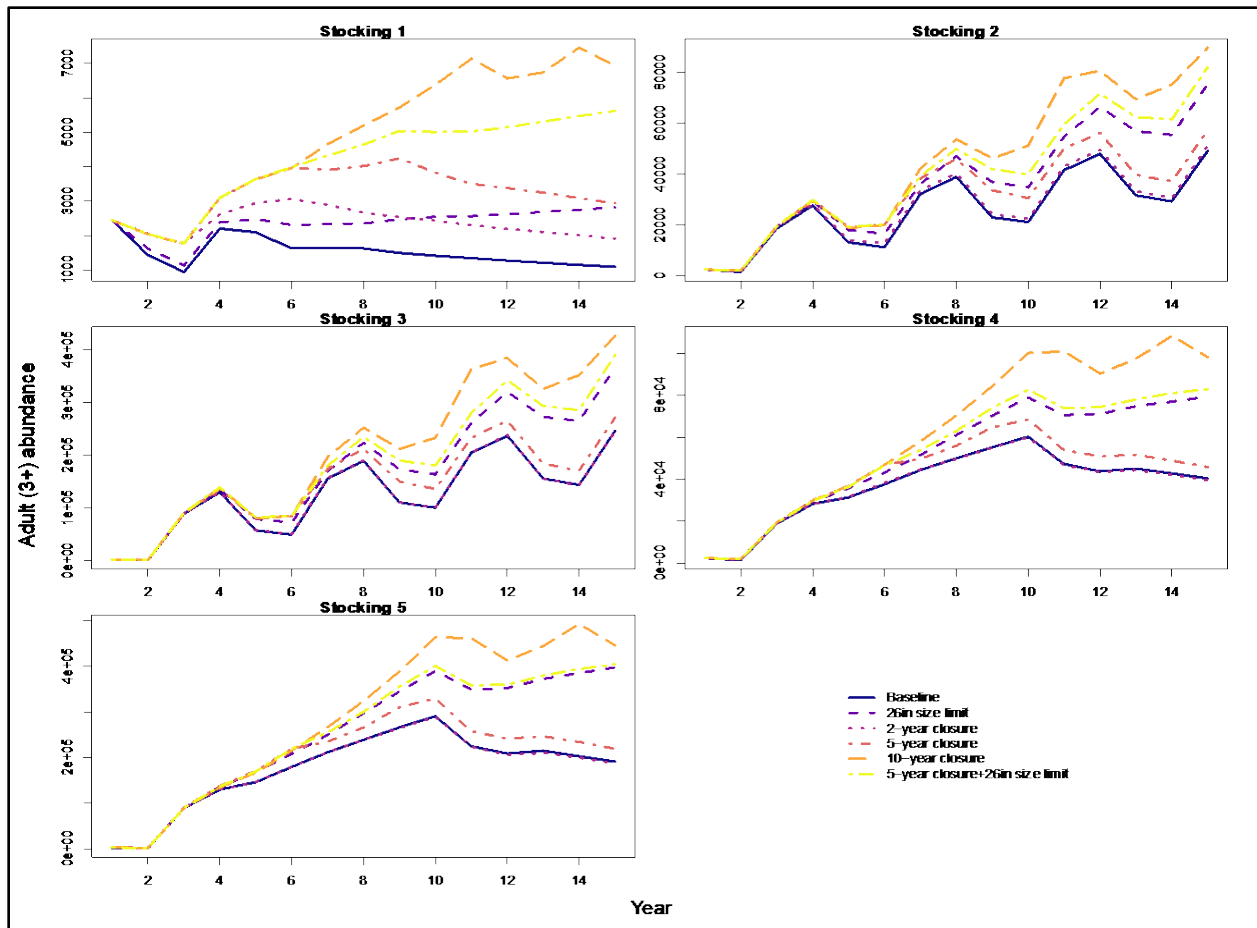




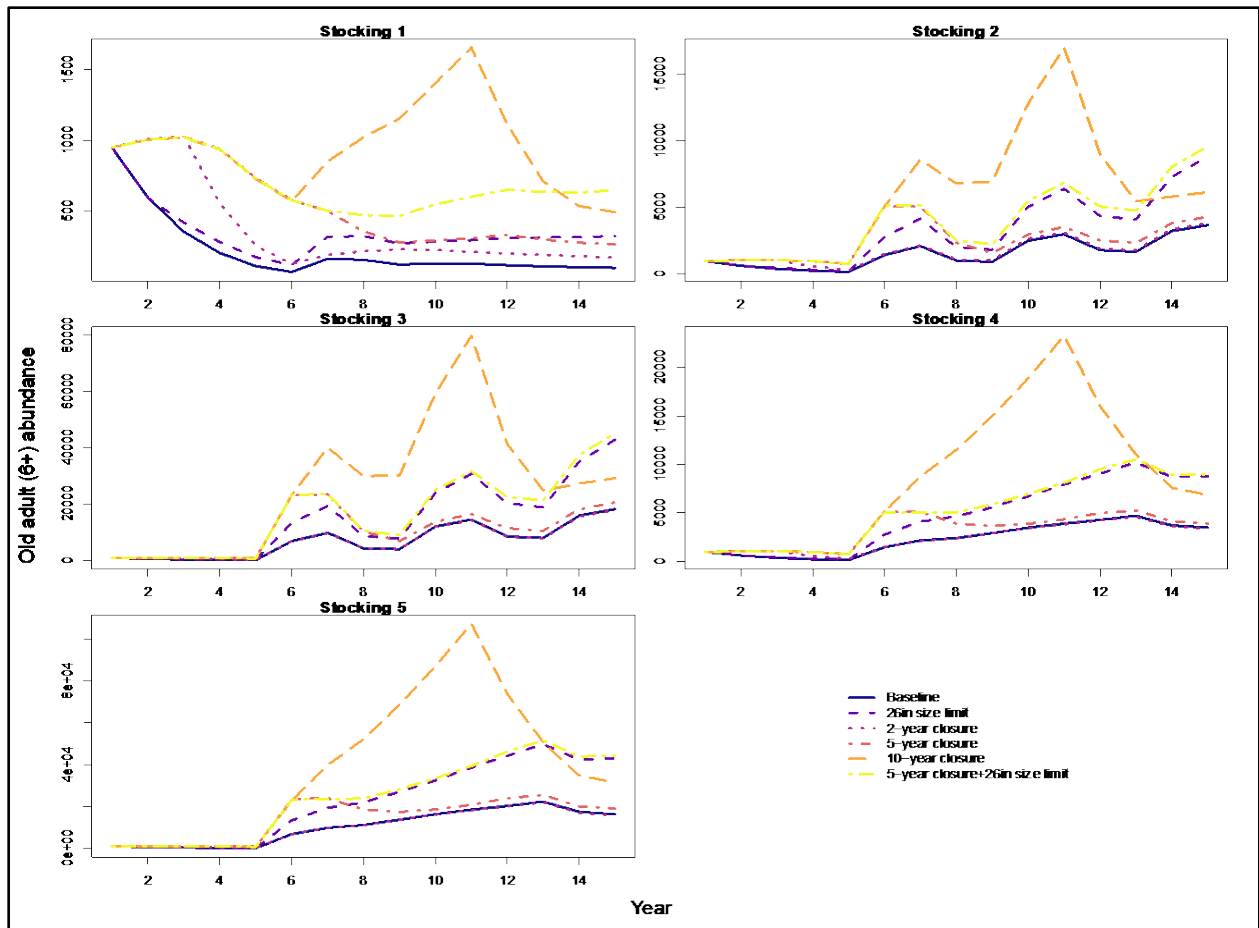
**Figure 3.2.** Elasticity of population growth rate to survival and fertility and age-specific reproduction contribution. Lines represent various fishing mortality ( $F$ ) values. Lines show the median from 10,000 iterations.



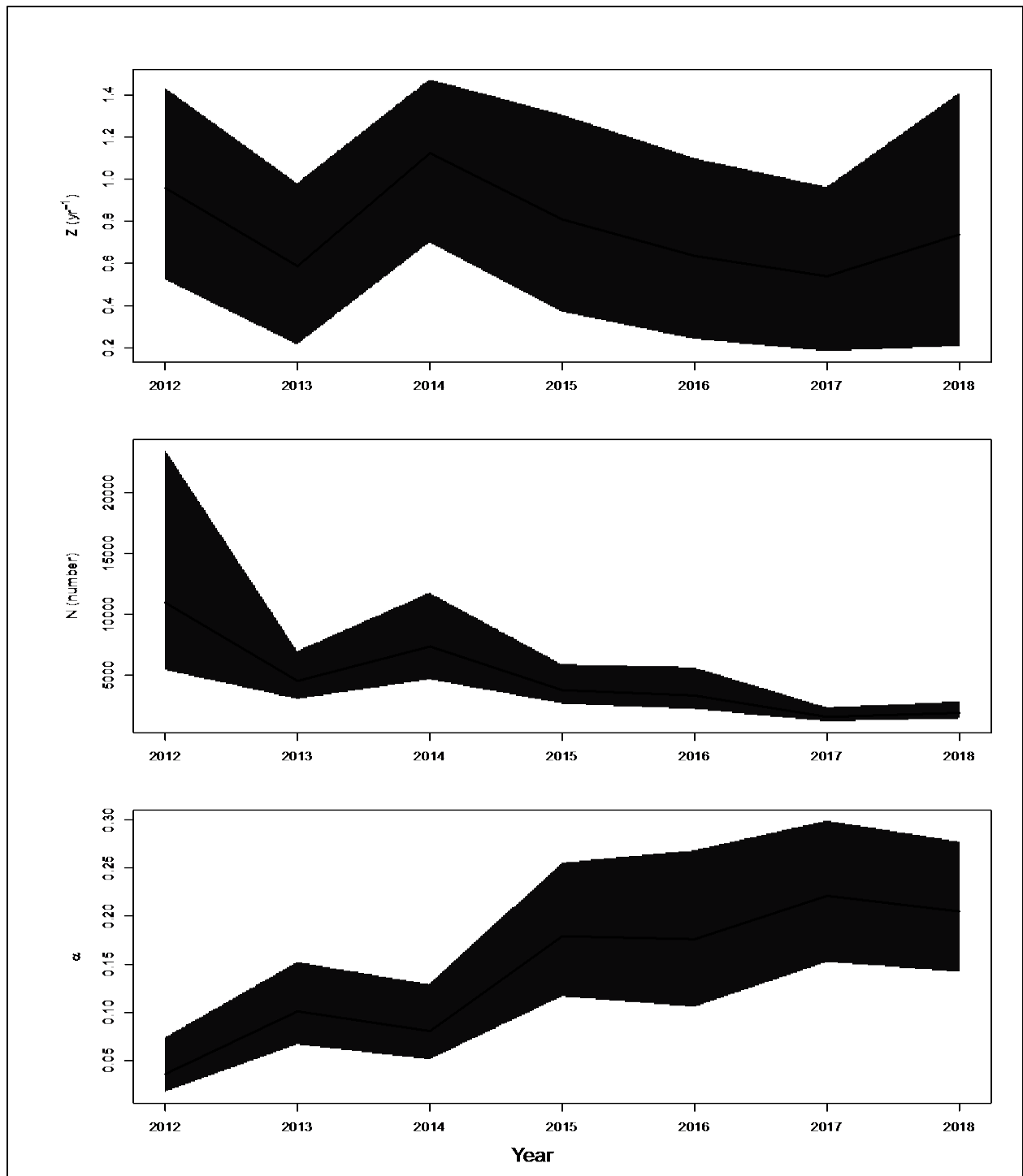
**Figure 3.3.** Sensitivity of population growth rate to viable egg proportion ( $x$ ), age-0 survival ( $S_0$ ) and the asymptotic length ( $L_\infty$ ). Lines represent various fishing mortality ( $F$ ) values. Lines show the median from 10,000 iterations.



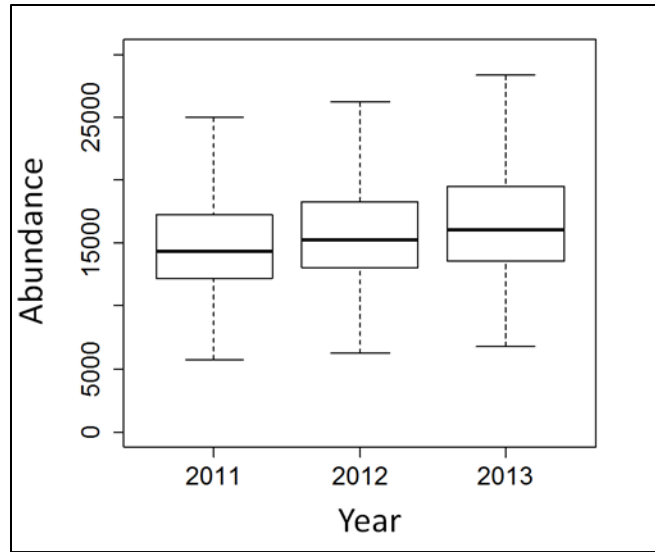
**Figure 3.4.** Abundance of adults (age 3+) projected under five stocking strategies and six fishing strategies. Stocking 1—no stocking; Stocking 2—stocking 100,000 fish per year with 2-year stocking and 2-year no stocking alternating for 15 years (8 years of stocking in total); Stocking 3—stocking 500,000 fish per year with 2-year stocking and 2-year no stocking alternating for 15 years (8 years of stocking in total); Stocking 4—stocking 100,000 fish per year with 8-year continuous stocking; Stocking 5—stocking 500,000 fish per year with 8-year continuous stocking. Lines show the median from 10,000 iterations.



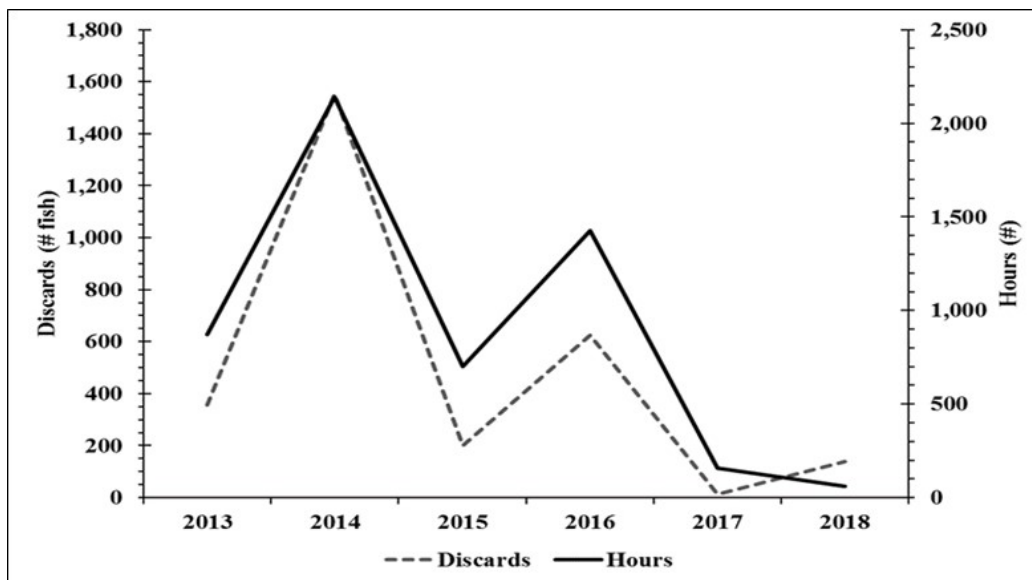
**Figure 3.5.** Abundance of old adults (age 6+) projected under five stocking strategies and six fishing strategies. Lines show the median from 10,000 iterations. See Figure 3.4 caption for explanation of the five stocking strategies.



**Figure 4.1.** Estimated instantaneous total mortality ( $Z$ ,  $\text{yr}^{-1}$ ) due to natural causes and fishing, estimated abundance ( $N$ , number) and estimated capture probability ( $\alpha$ ) from the tagging model. Line is posterior median and shaded area is 95% credible interval.



**Figure 4.2.** Posterior distributions of annual abundance estimated using a Jolly-Seber model and capture probabilities estimated by the multistate model in the Cape Fear River. The whiskers of the boxplots indicate 95% credible intervals of the estimates; boxes of the boxplots represent 50% credible intervals and the bolded lines of each boxplot represent abundance estimates. (Source: Collier et al. 2013)

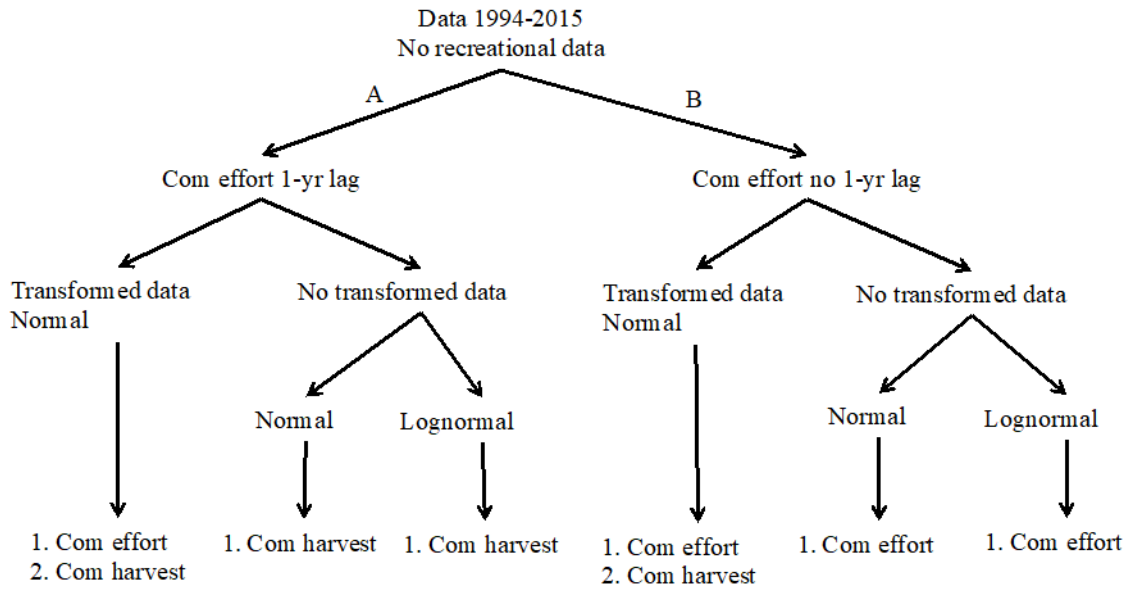


**Figure 4.3.** NCDMF recreational creel survey estimated striped bass discards (number; dotted line) and recreational fishing effort (hours; solid line) in the Cape Fear River, 2013–2018. In 2013, due to comparatively low recreational striped bass catch, American and hickory shad became the target species.

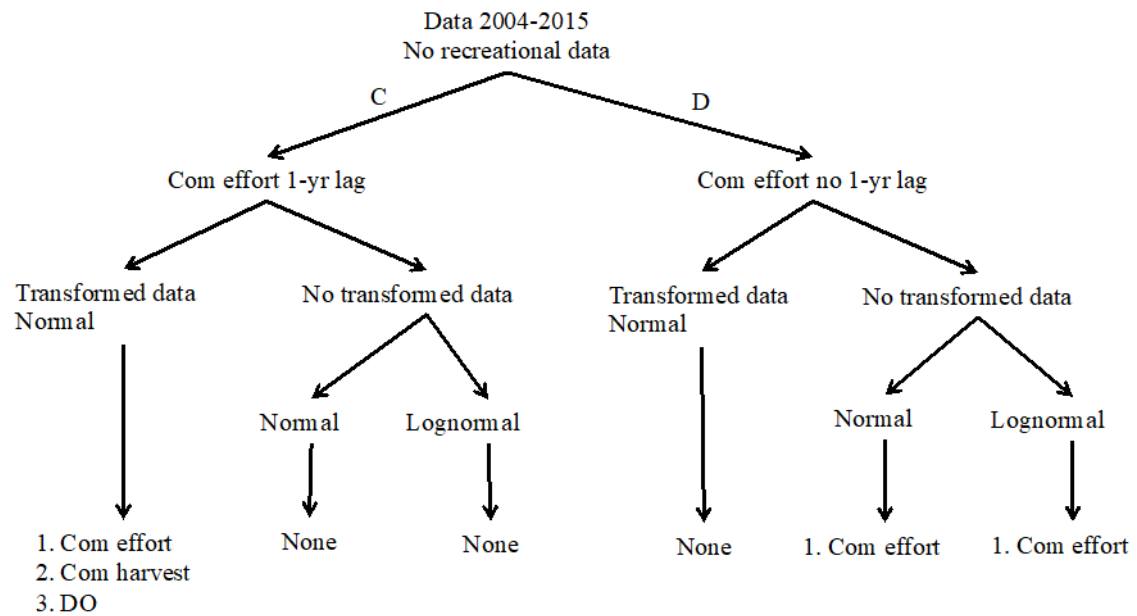


**Figure 4.4.** Dead striped bass at Battleship Park, Wilmington, NC following extensive flooding from Hurricane Florence in September 2018.

(A)

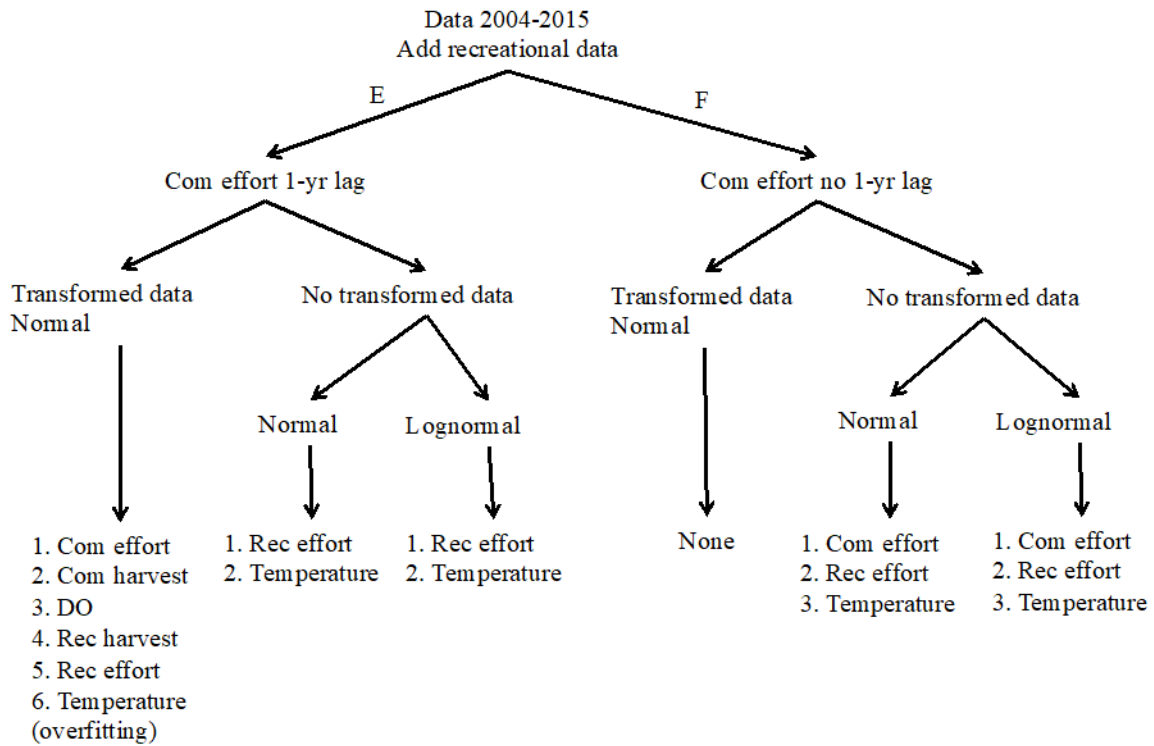


(B)

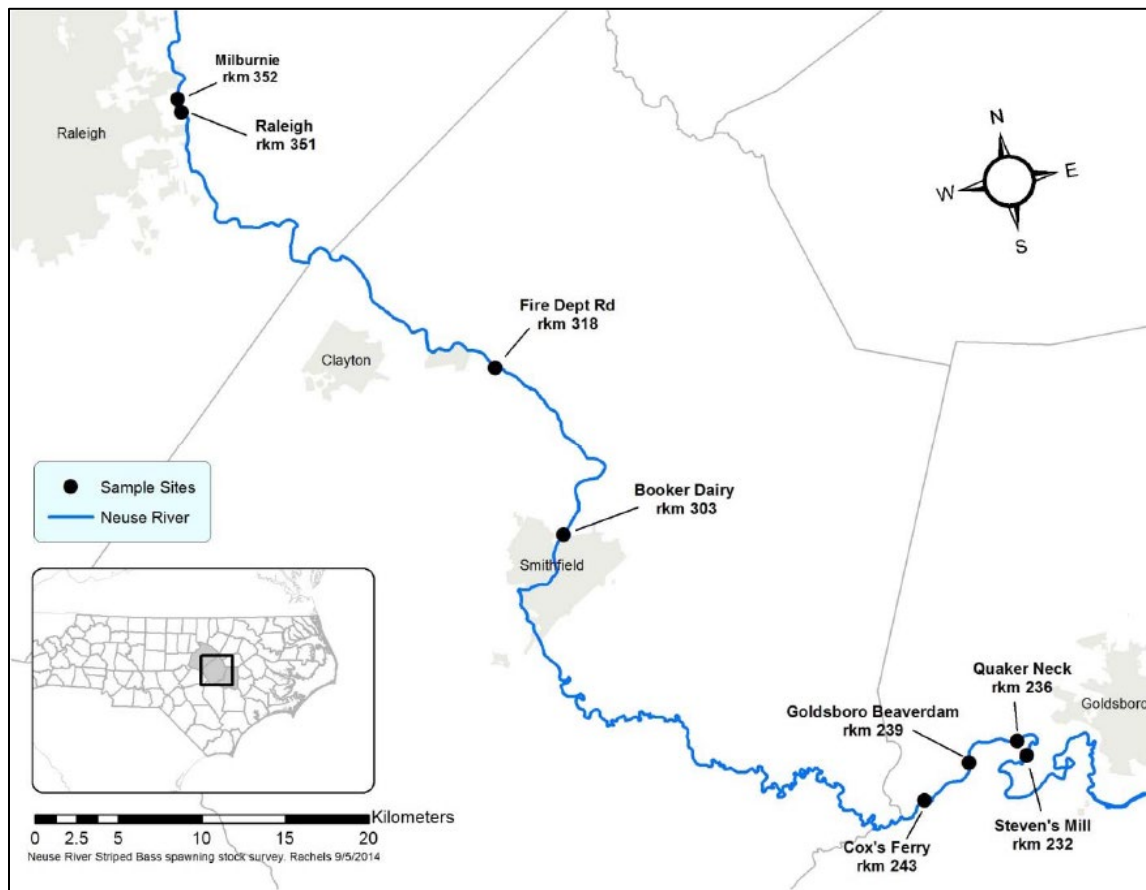




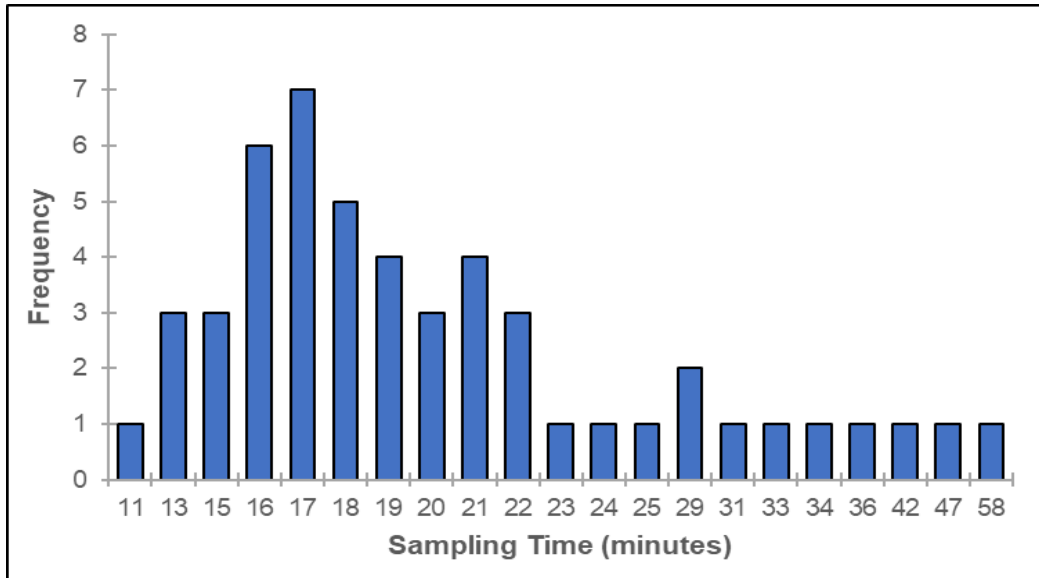
(C)



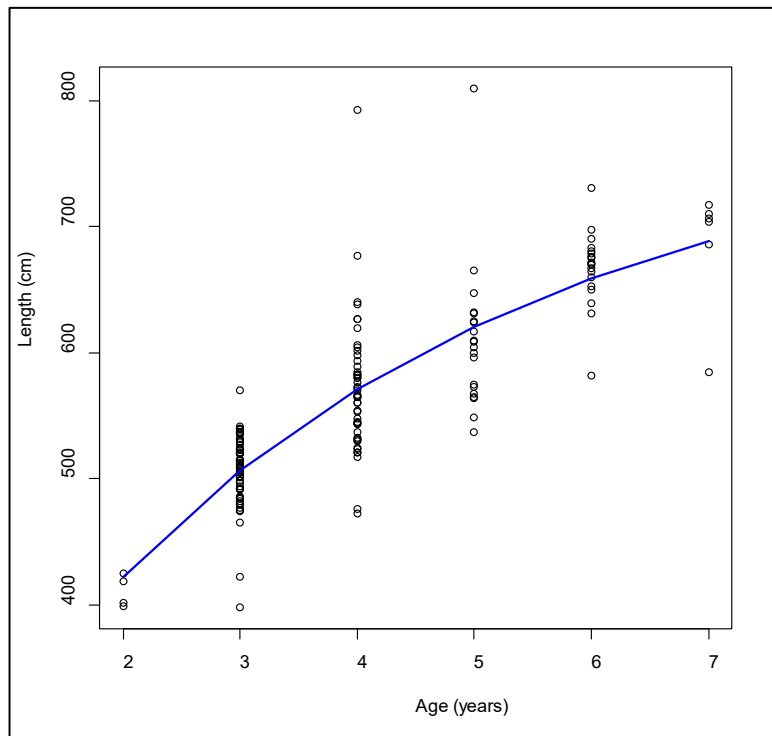
**Figure 5.1.** Important factors selected in the model when using data from (A) 1994–2015, and (B) data from 2004–2015 without considering recreational information, and (C) when using data from 2004–2015 with recreational information included. These factors are listed in the order of importance from the most important to the least important ones. See the caption of Table 1 for abbreviations of the predictor variables.



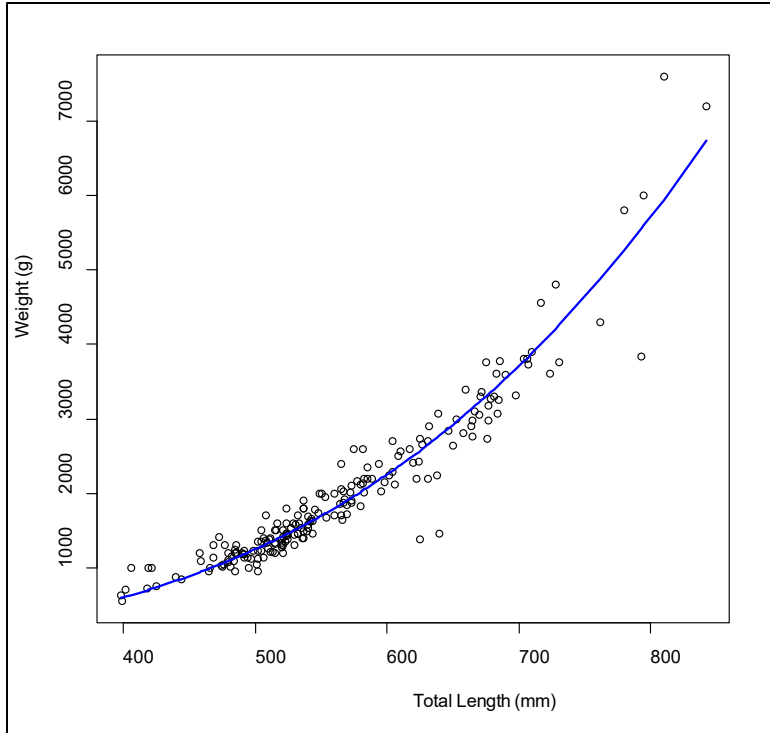
**Figure 6.1.** Sampling sites in the Neuse River for the NCWRC's Spawning Stock Survey.



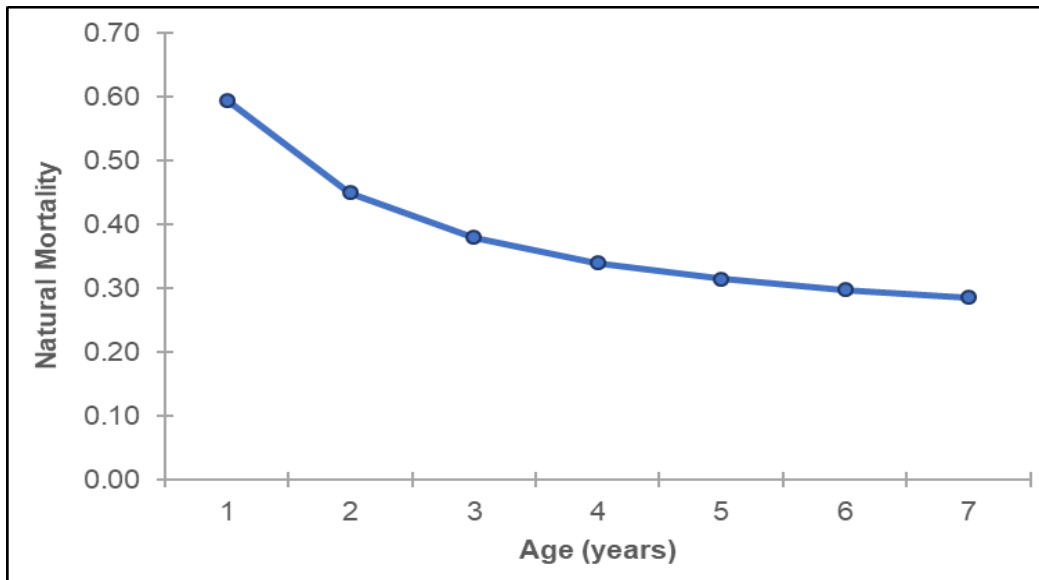
**Figure 6.2.** Range of sampling times for individual sampling trips from the NCWRC's Spawning Stock Survey in 2017.



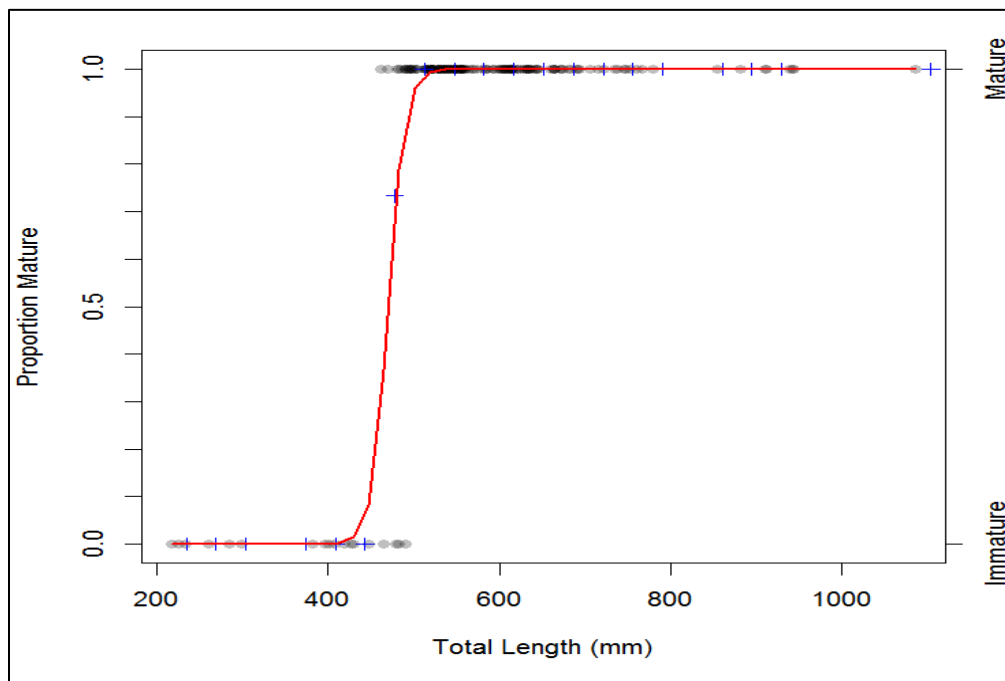
**Figure 6.3.** Observed (black circles) and predicted (blue line) values of the von Bertalanffy age-length relationship.



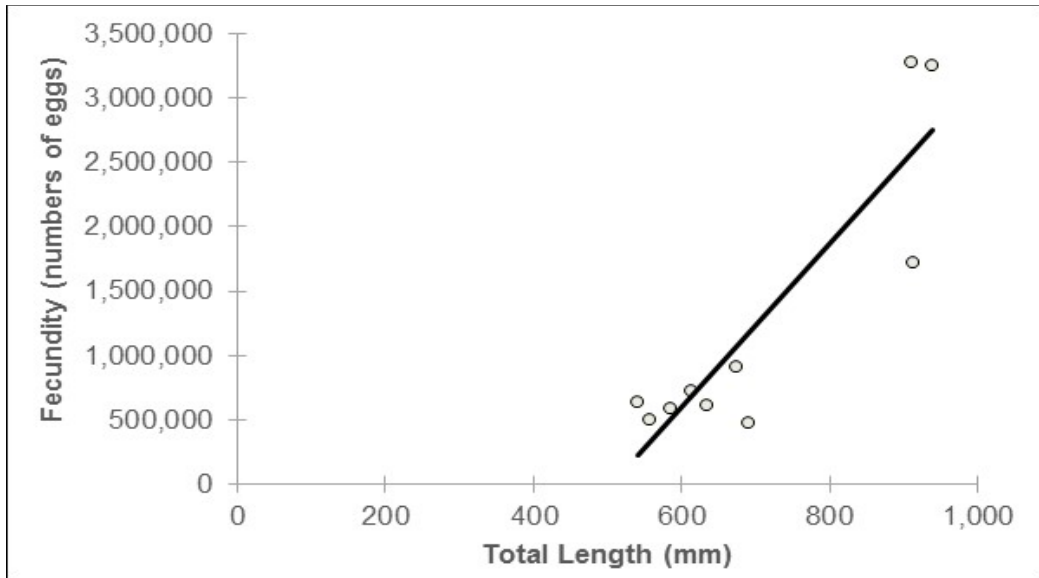
**Figure 6.4.** Observed (open black circles) and predicted (blue line) values of the length-weight relationship.



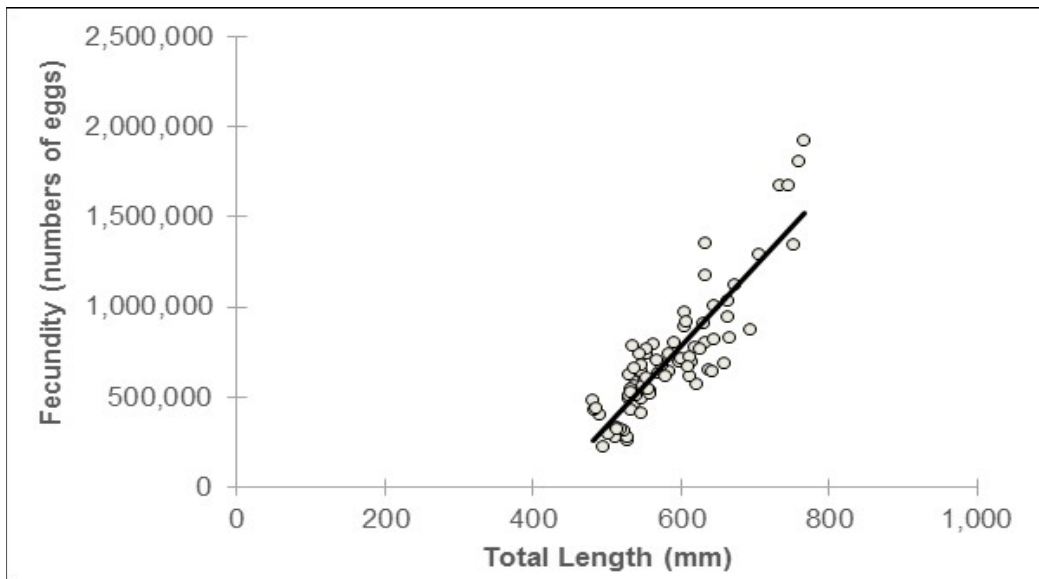
**Figure 6.5.** Estimated natural mortality at age based on Lorenzen’s (1996) approach. The values shown represent instantaneous rates.



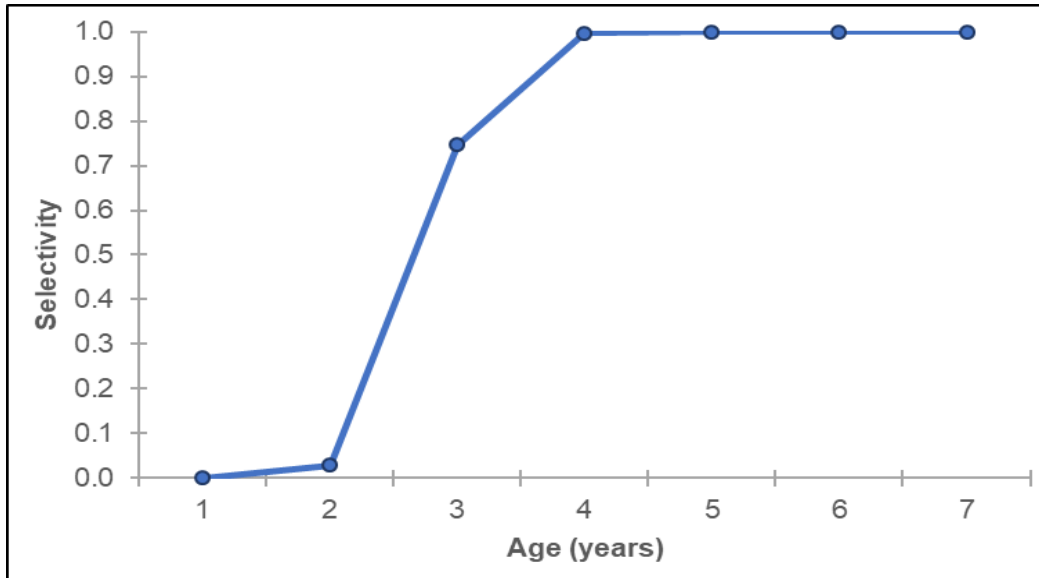
**Figure 6.6.** Observed (grey circles) and predicted (red line) values of the length-maturity relationship. The blue plus signs represent the proportion mature for selected length categories.



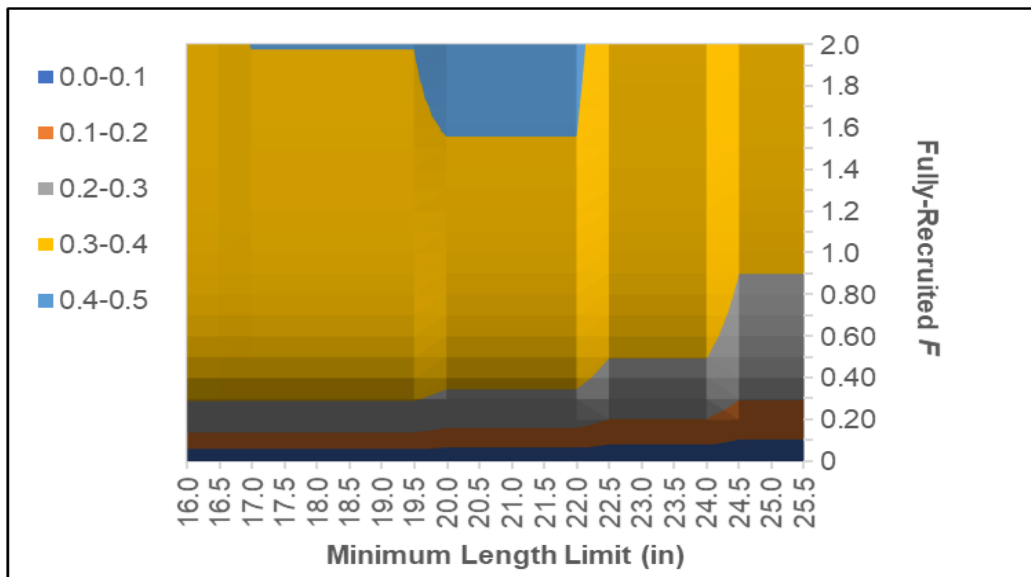
**Figure 6.7.** Observed (grey circles) and predicted (black line) values of the length-fecundity relationship for non-hatchery origin fish.



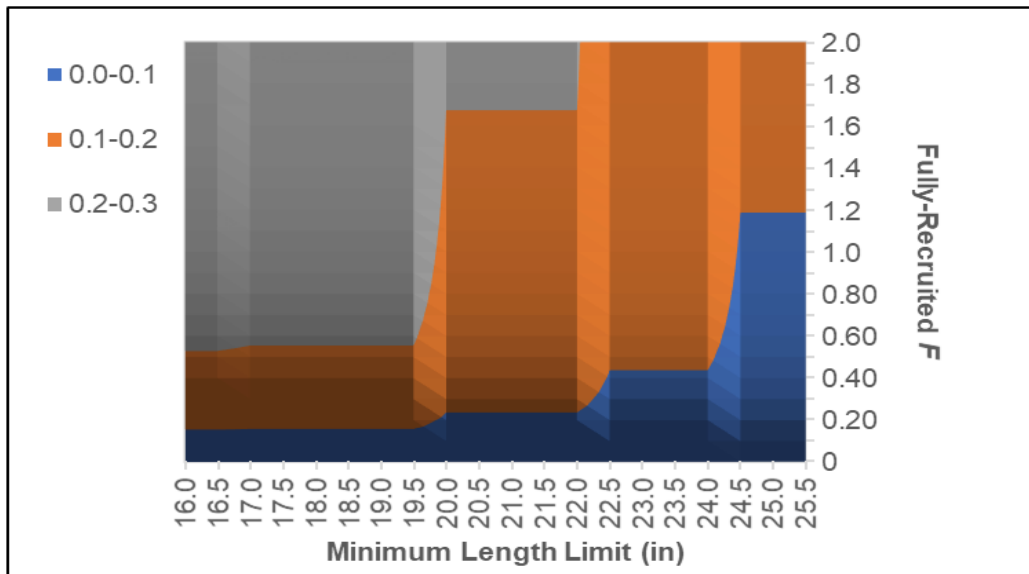
**Figure 6.8.** Observed (grey circles) and predicted (black line) values of the length-fecundity relationship for hatchery-origin fish.



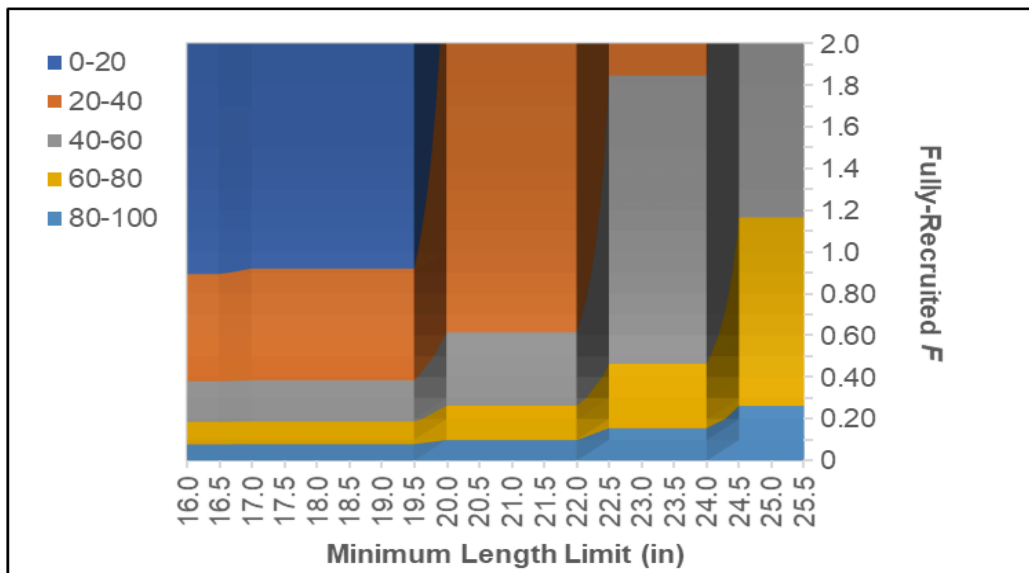
**Figure 6.9.** Selectivity at age assumed in the per-recruit analyses.



**Figure 6.10.** Yield per recruit in terms of weight (kilograms) at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.

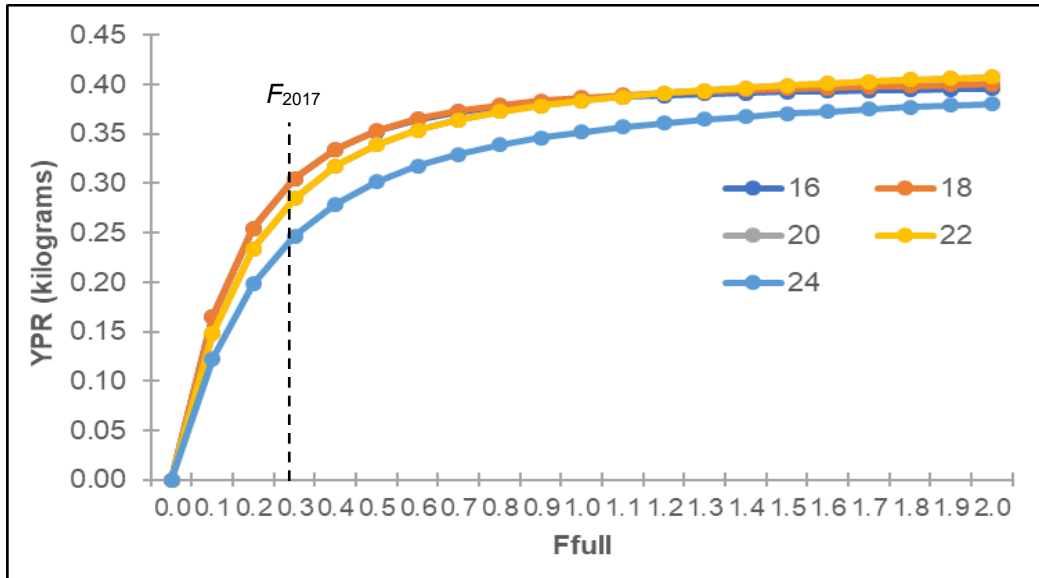


**Figure 6.11.** Yield per recruit in terms of numbers at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.

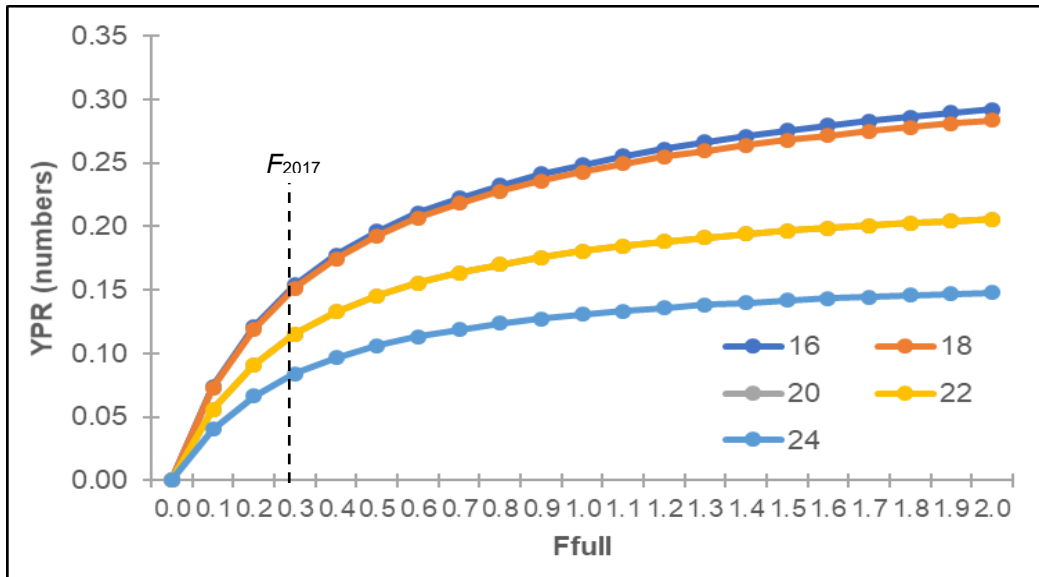


**Figure 6.12.** Spawning potential ratio (%SPR) at various combinations of fully-recruited fishing mortality ( $F$ ) and minimum length limits.

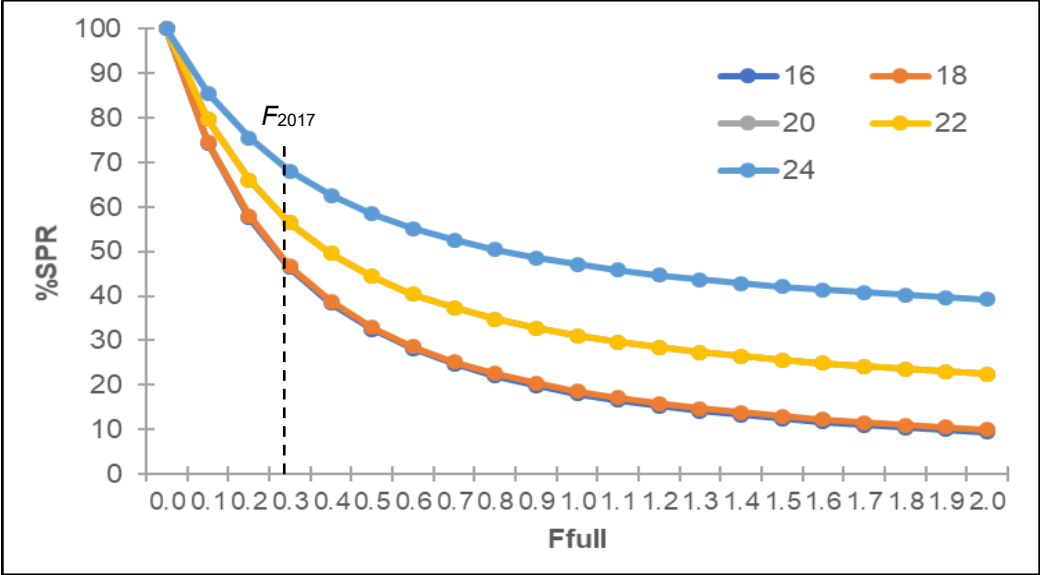




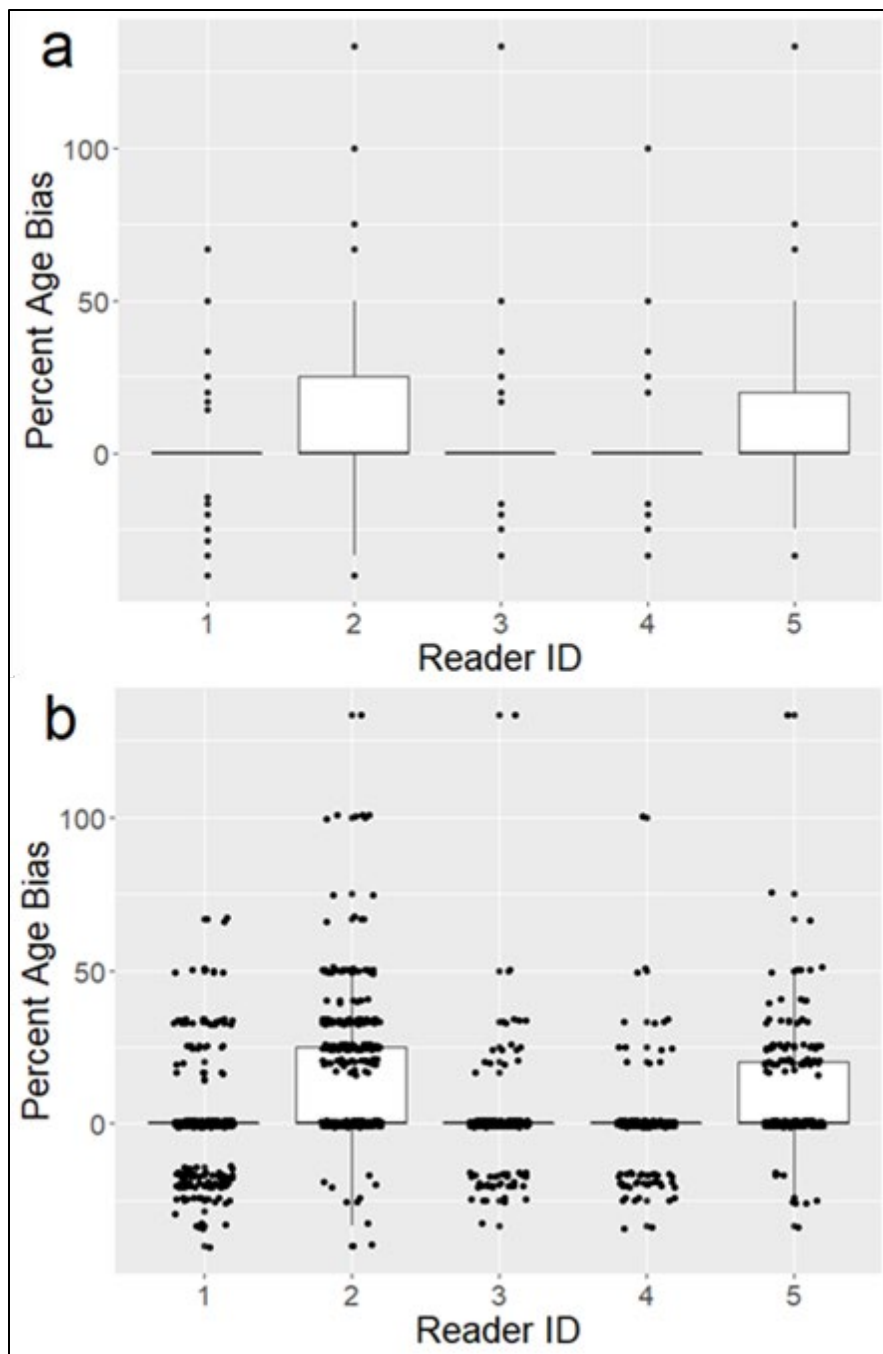
**Figure 6.13.** Yield per recruit in terms of weight (kilograms) over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.



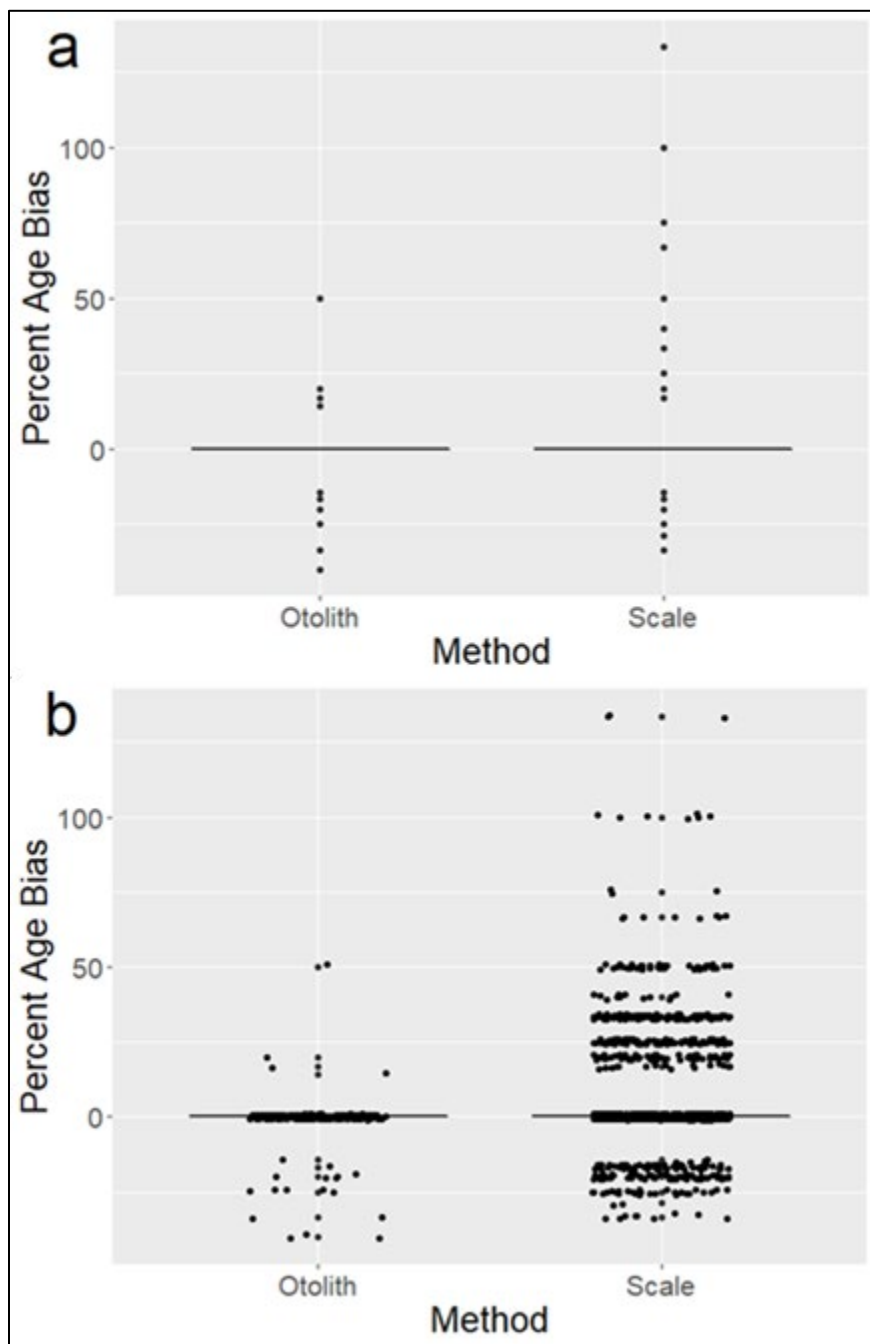
**Figure 6.14.** Yield per recruit in terms of numbers over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.



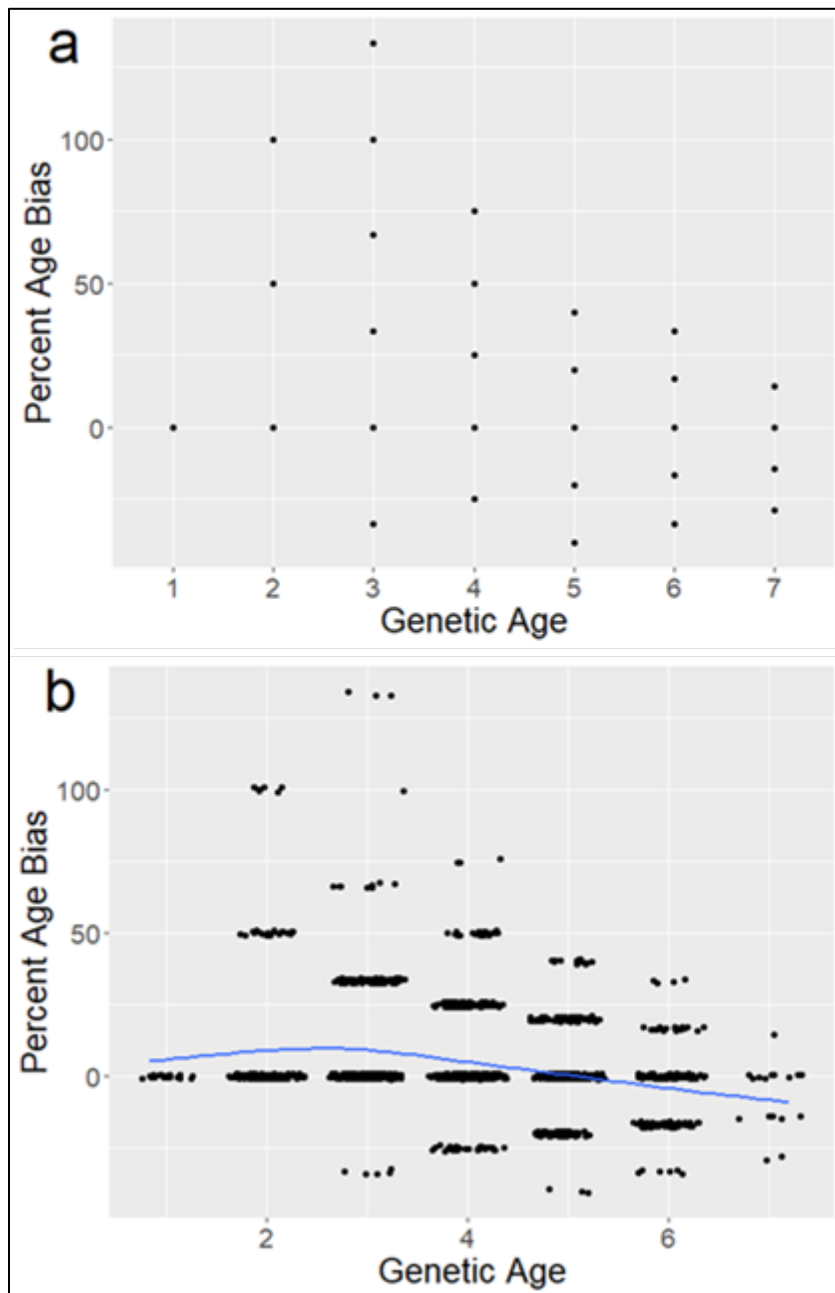
**Figure 6.15.** Spawning potential ratio (%SPR) over a range of fully-recruited fishing mortality rates ( $F_{full}$ ) for select minimum length limits.



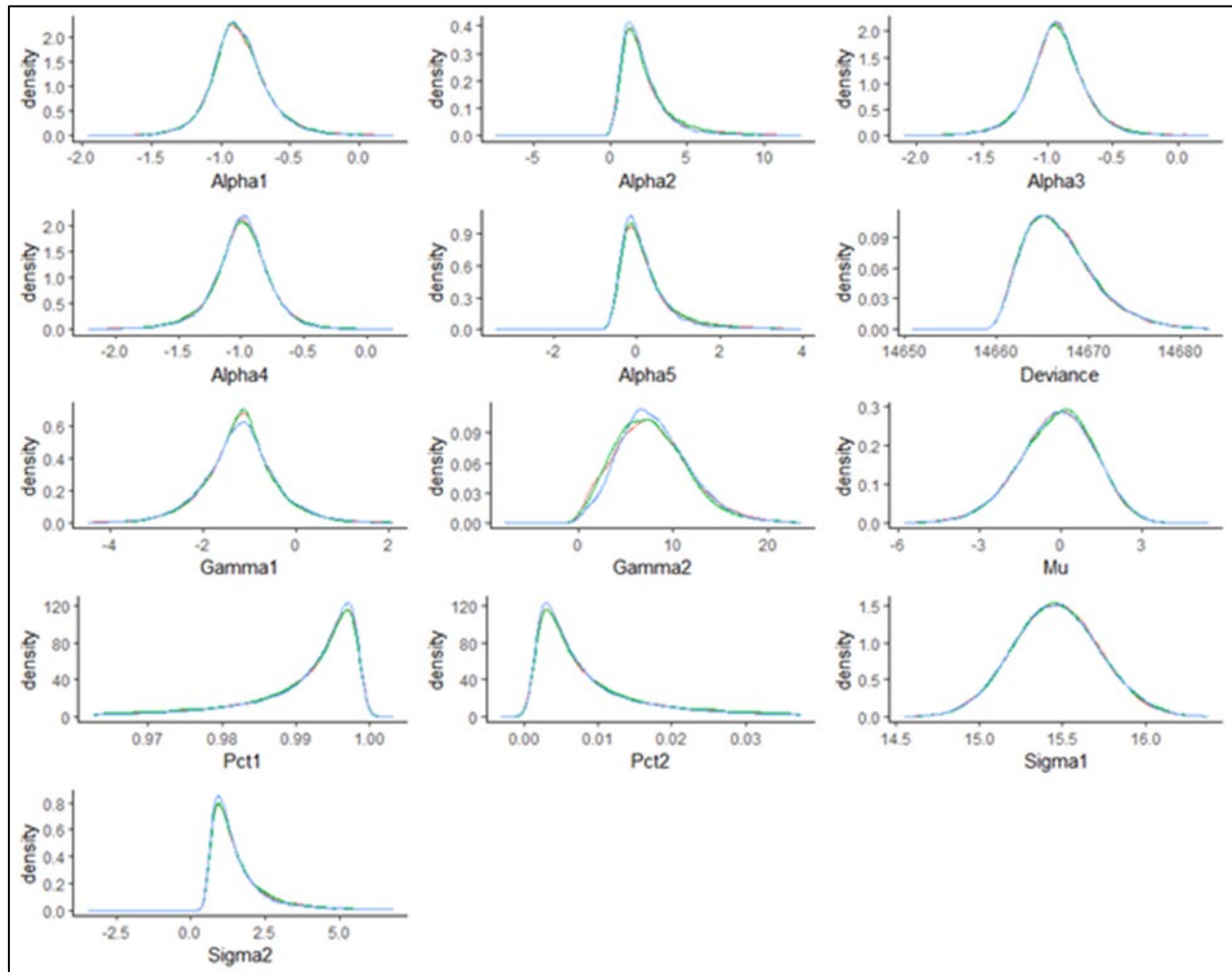
**Figure 7.1.** Boxplot of percentage age bias by reader ID. The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data.



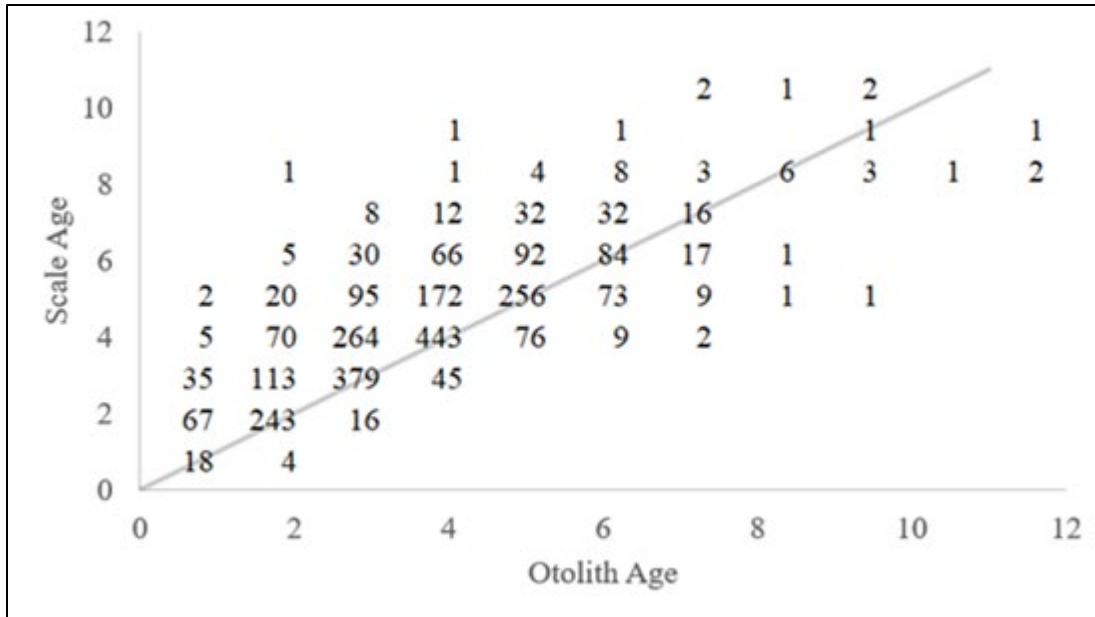
**Figure 7.2.** Boxplot of percentage age bias by ageing method. The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data.



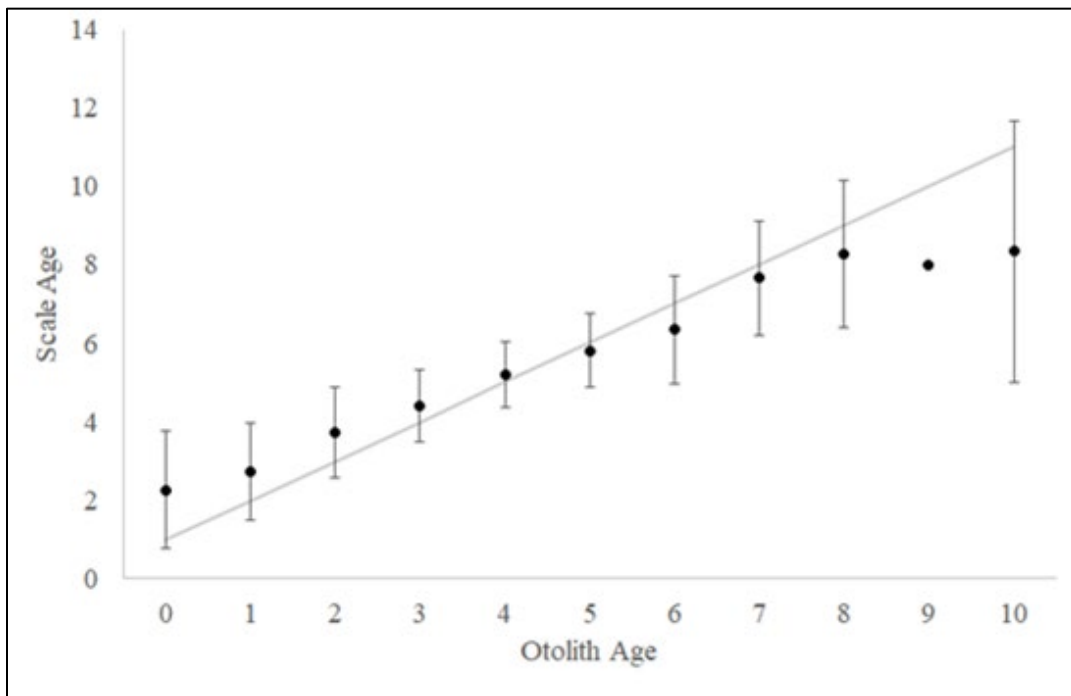
**Figure 7.3.** Percentage age bias by genetic age (from parental base tagging) with trend line (solid line). The majority of the data points overlapped each other as shown in graph a so the points were jittered (given slightly increased or decreased values) in graph b in order to provide contrasts in data points. The jittered values were not used in the analysis, only to aid in visual inspection of the data.



**Figure 7.4.** Posterior distributions for three chains of parameter estimates from Bayesian generalized linear mixed effects model. Alpha's represent reader effects, gamma's represent method effects, mu represents the overall average bias, pct1 represents percentage of error explained by random error, pct2 represents percentage of error explained by reader effects, sigma1 represents standard deviation associated with random error, sigma2 represents standard deviation associated with reader effects, and deviance is a goodness-of-fit estimate.



**Figure 7.5.** Contingency table for number of fish in each scale age for each otolith age. Numbers represent number of fish assigned scale age for a given otolith age.



**Figure 7.6.** Age-bias plot for average scale age for each otolith age with standard deviation.

## **12 APPENDIX A**

### **FORK LENGTH/ TOTAL LENGTH CONVERSION WORKING PAPER**

#### **ESTUARINE STRIPED BASS DATA WORKSHOP**

Planning Workshop

March 20, 2017

#### **NC DIVISION OF MARINE FISHERIES**

**PROGRAM 135 STRIPED BASS INDEPENDENT GILLNET SURVEY (ASMA)**

**PROGRAM 311 CAPE FEAR RIVER STRIPED BASS MARK RECAPTURE STUDY  
(CSMA – CAPE FEAR)**

**PROGRAM 366 MULTI-SPECIES TAGGING PROGRAM (CSMA – CAPE FEAR)**

**PROGRAM 930 COMPREHENSIVE LIFE HISTORY (CSMA – TAR-PAMLICO,  
NEUSE, AND CAPE FEAR)**

*Prepared by:* Todd Mathes, Marine Fisheries Biologist I, Washington, NC

#### **Analysis Overview**

Differences in striped bass length measurement types collected between and within North Carolina State agencies necessitates standardization to compare samples among systems. The 2017 estuarine striped bass stock assessment planning workshop terms of reference established total length as the standard unit of measurement for the striped bass stock assessment. To this end, simple linear regression was used to compare total length as a function of fork length to establish a conversion for instances where only fork length was recorded.

Data were provided from the divisions' biological database from various fishery independent and dependent data collection programs (Table 1). Geographic areas analyzed included: Albemarle Sound Management Area (ASMA), Central Southern Management Area (CSMA; Pamlico Sound and Tar/Pamlico, Pungo, and Neuse rivers), and CSMA (Cape Fear River).

#### **Program Objectives**

The Striped Bass Independent Gillnet Survey (P135) is used to monitor the Albemarle/Roanoke striped bass population. The principle objectives are to describe the striped bass population as to length, age, sex, and relative abundance.

The Cape Fear River Striped Bass Mark Recapture Study (P311) is a tagging study used to: 1) estimate the population size of striped bass in the Cape Fear River, 2) estimate tag loss of internal anchor tags, and 3) compare recapture rates of striped bass caught with hook and line, electrofishing, and gill net gears. Secondary objectives of the study are obtaining age samples from striped bass in the Cape Fear River and determine residency patterns of striped bass in the Cape Fear River.

The Multi-Species Tagging Program (P366) was developed to standardize protocols for coding tag data amongst various existing programs conducted by the division and designed to accommodate future tagging projects as needed regardless of species being tagged. The overall objective is to provide a multi-species tagging program with a standardized coding procedure for conventional



tags. The specific objectives are to: 1) estimate tag-retention rates, tag-reporting rates, fishing mortality by fishing sector, and migration rates for red drum, striped bass, spotted seatrout, southern flounder, and cobia 2) estimate fishing mortality by fate (harvest or release), age, and fishing sector and to provide selectivity estimates by fate, age and fishing sector for red drum, striped bass, spotted seatrout, southern flounder, and cobia, and 3) assess annual variation in fishing and natural mortalities using a tag-return model, conventional catch-at-age stock assessment model, or an integrated tag-return catch-at-age model for red drum, striped bass, spotted seatrout, southern flounder, and cobia.

The Comprehensive Life History Program (P930), created in 1985, was developed to increase the understanding of the population dynamics and life history of North Carolina fishes and to collect fish ageing structures and other biological data to develop and validate life history information.

## **Survey Design & Methods**

### Data Source

The Striped Bass Independent Gillnet Survey (P135), ongoing since October 1990, is a random stratified multi-mesh monofilament gillnet survey. Mesh sizes used in the survey consist of 2.5 through 7.0 inch stretched mesh (ISM) at ½ inch increments, and 8.0 and 10.0 ISM. The fishing year is divided into three segments: (1) a fall/winter survey period, which begins approximately 1 November and continues through 28 February, (2) a spring survey period which begins 1 March and continues through approximately 30 June, and (3) a summer survey period which starts 1 July and continues through 30 October.

The Cape Fear River Striped Bass Mark Recapture Study (P311), 2010-2014, sampled thirty-two fixed stations in addition to randomly selected stations that were sampled in January to April each year. In 2015 striped bass tagging from this program transitioned to P366 and its sampling protocols.

The Multi-Species Tagging Program (P366), implemented 1 October, 2014, is the primary program for documenting the divisions' conventional fish tagging. Red drum, striped bass, spotted seatrout, sturgeon, southern flounder, and cobia are tagged by division staff using a variety of methods. Fish are captured through division fishery independent and dependent sampling programs. A limited number of recreational hook-and-line fishermen recruited by division staff will also tag these fish species. Sampling for this program is diverse both geographically and by gear type to achieve the studies objectives.

The Comprehensive Life History Program (P930) began collecting and ageing of fish otoliths and scales in the late 1970's. Currently, regular data collection occurs for approximately 20 recreationally and commercially important North Carolina finfish species. In the past, P930 has had no specific sampling design; ageing samples have been collected opportunistically or as needed from division fishery independent sampling, commercial catches, and recreational catches, depending on the species. Otoliths and/or scales are collected monthly from American shad, Atlantic croaker, Atlantic menhaden, black drum, black sea bass, bluefish, cobia, kingfishes, mackerels, flounders, red drum, sheepshead, spotted seatrout, spot, striped bass, striped mullet, and weakfish.

## Analysis Methods

Due to the large number of observations within the ASMA data set, spanning 1990 to present, only years 2000-2016 were used for the analysis. Initial data provided were screened to remove outliers. Two methodologies were used to establish a threshold to identify outliers: (1)  $(TL-FL)/TL > 15\%$ , and (2)  $FL > TL$ . Once the outliers were identified/removed, data were further cleaned to ensure accuracy of coding. Simple linear regression was then used to compare total length as a function of fork length. Simple linear regression is a parametric statistical test predicated on assumptions of normality, and homoscedasticity (equality of variances). Linear regression tests the null hypothesis that there would be no significant prediction of total length by fork length. All data were analyzed using SAS 9.3.

Our hypotheses are as follows:

$H_0: \rho = 0$  there is no correlation between fork length and total length within our population

$H_0: \rho \neq 0$  there is a significant correlation between fork length and total length

Where  $\rho$  is our correlation coefficient (measures the strength and direction of a linear relationship between two variables)

A student's t-test is used to determine if the relationship between our independent (fork length) and dependent variables (total length) are different from zero.

$$t = r\sqrt{(n-2)/(1-r^2)}$$

where,  $r = 1/n-1 \sum (x_i - \bar{x})(y_i - \bar{y}) / s_x s_y$

## Results

Results of the analyses validates that the assumptions of normality had been met (Figure 1, Figure 3, and Figure 5), and that the amount of variability within datasets were very low (Figure 2, Figure 4, and Figure 6) demonstrating equality of variances.

## Conclusion

Regressions from all three areas exhibited essentially the same slopes and Y intercepts differed by less than 5 millimeters. Based on these results, it is appropriate to pool data from all the regions. In conclusion, when converting fork length to total length, pooled data can be used to accurately predict total length. Listed below are the formulas for converting fork length to total length, as well as a reciprocal equation in case there is an instance where total length needs to be converted to fork length.

### FL to TL Conversion Formula:

$$\text{Total Length} = 6.206909513 + (1.055954699 * \text{Fork Length})$$

### Example:

FL = 640 mm, what's the TL?

$$TL = 6.206909513 + (1.055954699 * 640) =$$

$$TL = 6.206909513 + 675.811 =$$

$$TL = 682.0179 \text{ mm}$$

$$TL = 682 \text{ mm}$$

**Reciprocal TL to FL Conversion Formula:**

$$\text{Fork Length} = (\text{Total Length} * 0.945673822) - 5.277089838$$

**Example:**

TL = 682 mm, what's the FL?

$$\text{TL} = (682 * 0.945673822) - 5.277089838 =$$

$$\text{TL} = 644.949546604 - 5.277089838 =$$

$$\text{TL} = 639.6724 \text{ mm}$$

$$\text{TL} = 640 \text{ mm}$$

**Dataset Information**

Charlton Godwin, [Charlton.Godwin@ncdenr.gov](mailto:Charlton.Godwin@ncdenr.gov),

File Location:

U:\striped bass\Stock Assessment Benchmark FMP 2017\2\_Data Workshop\Data\Life History\FL-TL Conversion

P135 dataset:

AR STB FL TL conversion.xlsx

Chris Stewart, [Chris.Stewart@ncdenr.gov](mailto:Chris.Stewart@ncdenr.gov)

File Location:

U:\striped bass\Stock Assessment Benchmark FMP 2017\2\_Data Workshop\Data\Life History\FL-TL Conversion

P311, P366, and P930 dataset:

p311&366\_cfr\_stb.sas7bdat

cfr\_stb.sas7bdat

Chris Wilson, [Chris.Wilson@ncdenr.gov](mailto:Chris.Wilson@ncdenr.gov)

File Location:

U:\striped bass\Stock Assessment Benchmark FMP 2017\2\_Data Workshop\Data\Life History\FL-TL Conversion

SAS Program:

length regression.sas

Analysis dataset:

sbass.sas7bdat

eg\_clean.sas7bdat

Todd Mathes, [Todd.Mathes@ncdenr.gov](mailto:Todd.Mathes@ncdenr.gov)

File Location:

U:\striped bass\Stock Assessment Benchmark FMP 2017\2\_Data Workshop\Data\Life History\FL-TL Conversion

P930 dataset:

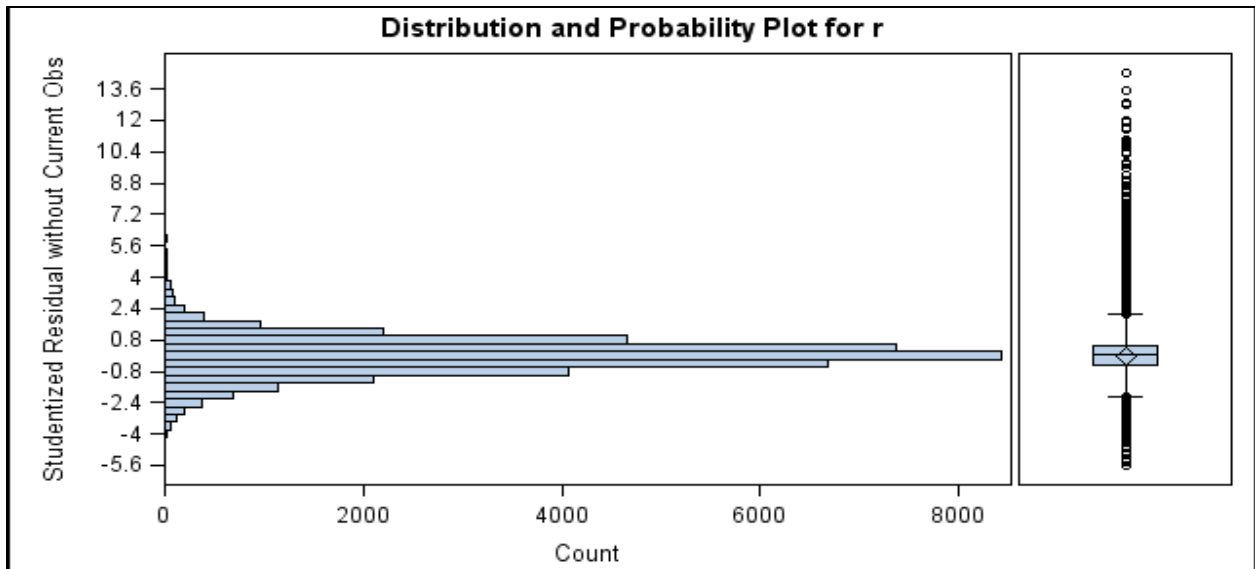
CSMA STB FL to TL conversion (4-20-17).xls

**Tables**

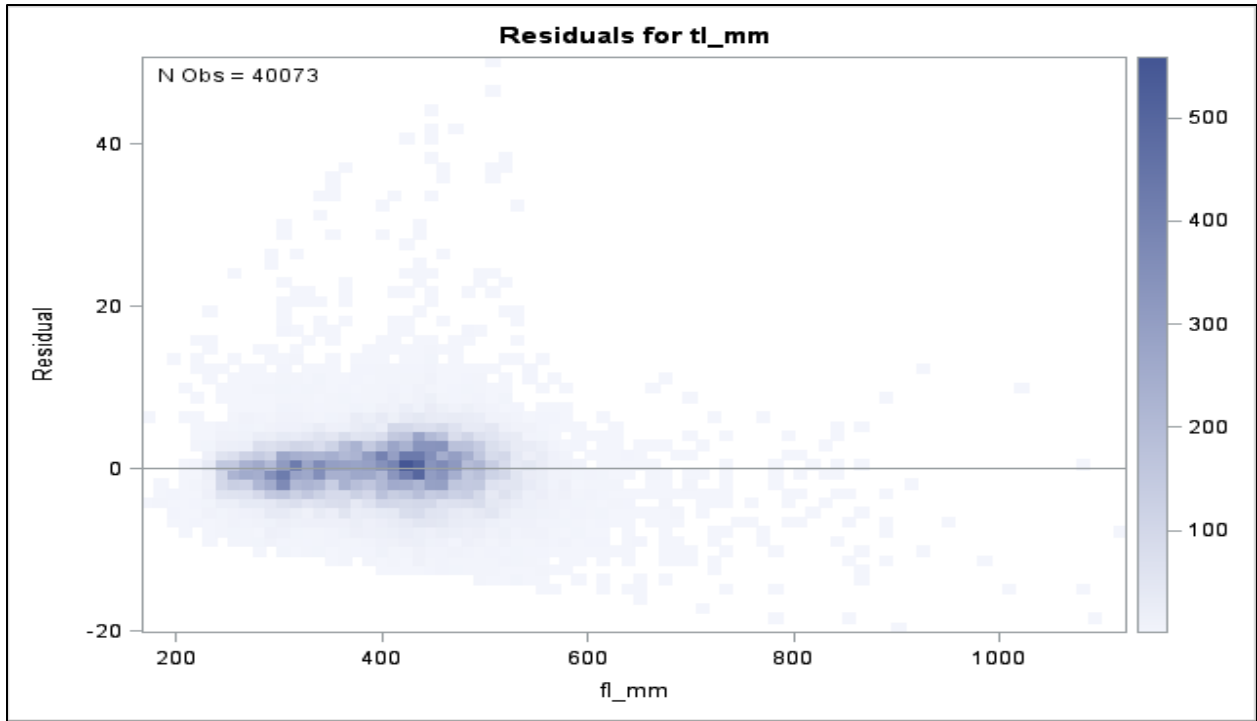
**Table 1.** FL to TL conversion data description.

Area	Program	n	Years	Data Source
ASMA	P135	40,073	2000–2016	Charlton Godwin
CSMA—Tar-Pamlico	P930	3,764	2000–2016	Todd Mathes
CSMA—Neuse	P930	2,482	2000–2016	Todd Mathes
CSMA—Cape Fear	P311, P366, P930	2,372	2011–2016	Chris Stewart

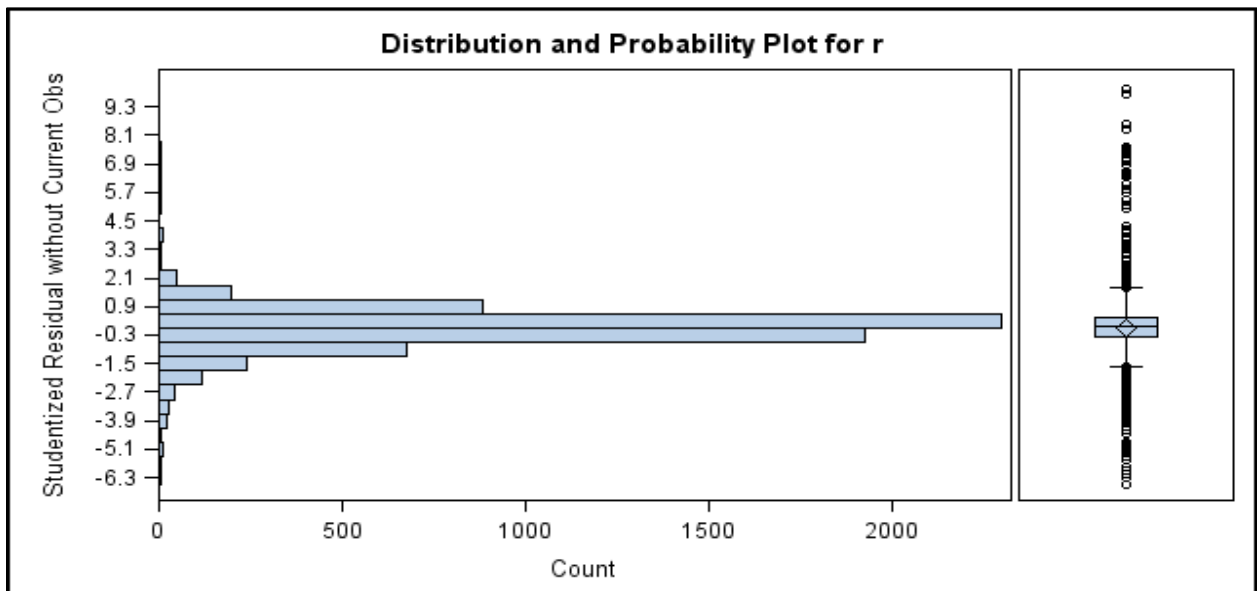
**Figures**



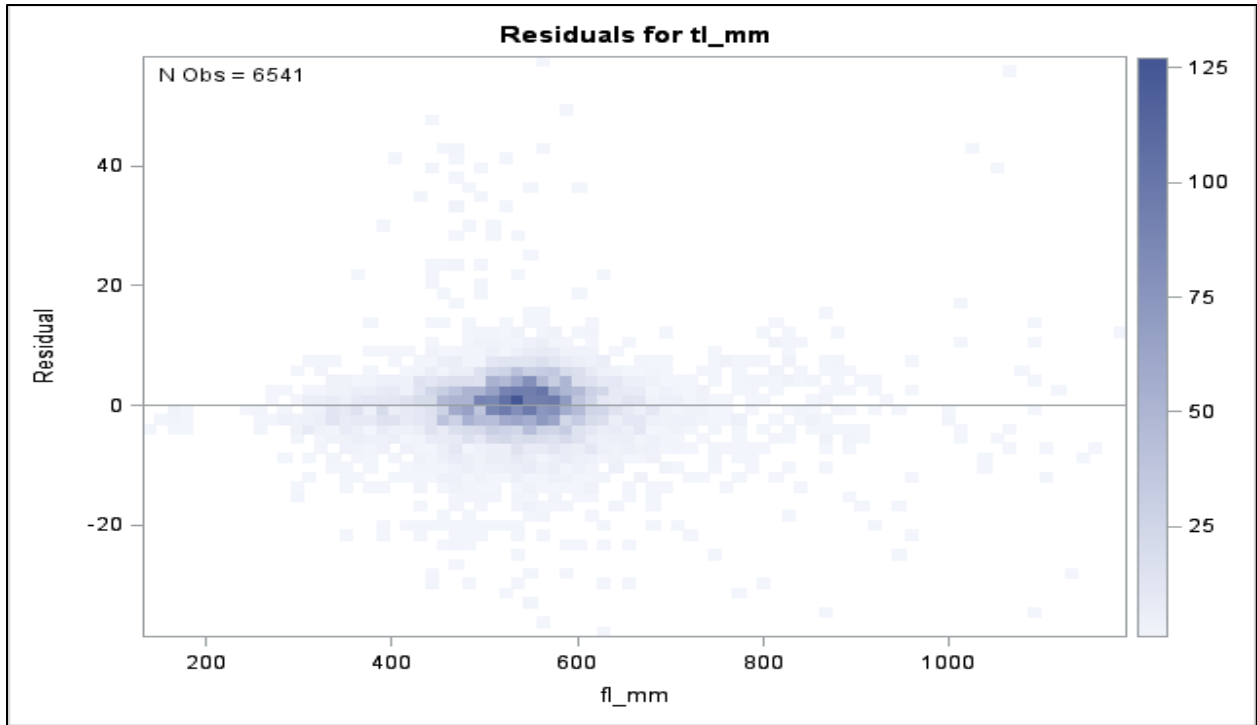
**Figure 1.** ASMA residual plot validating assumptions of normality.



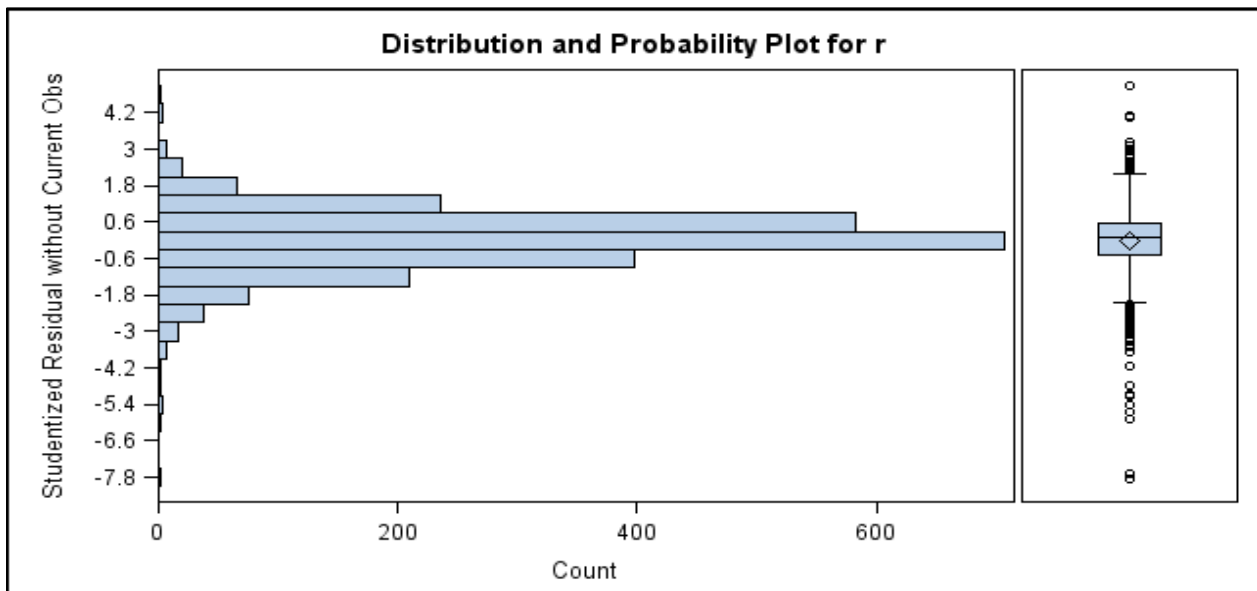
**Figure 2.** ASMA residuals demonstrate low variability associated with the best fit line.



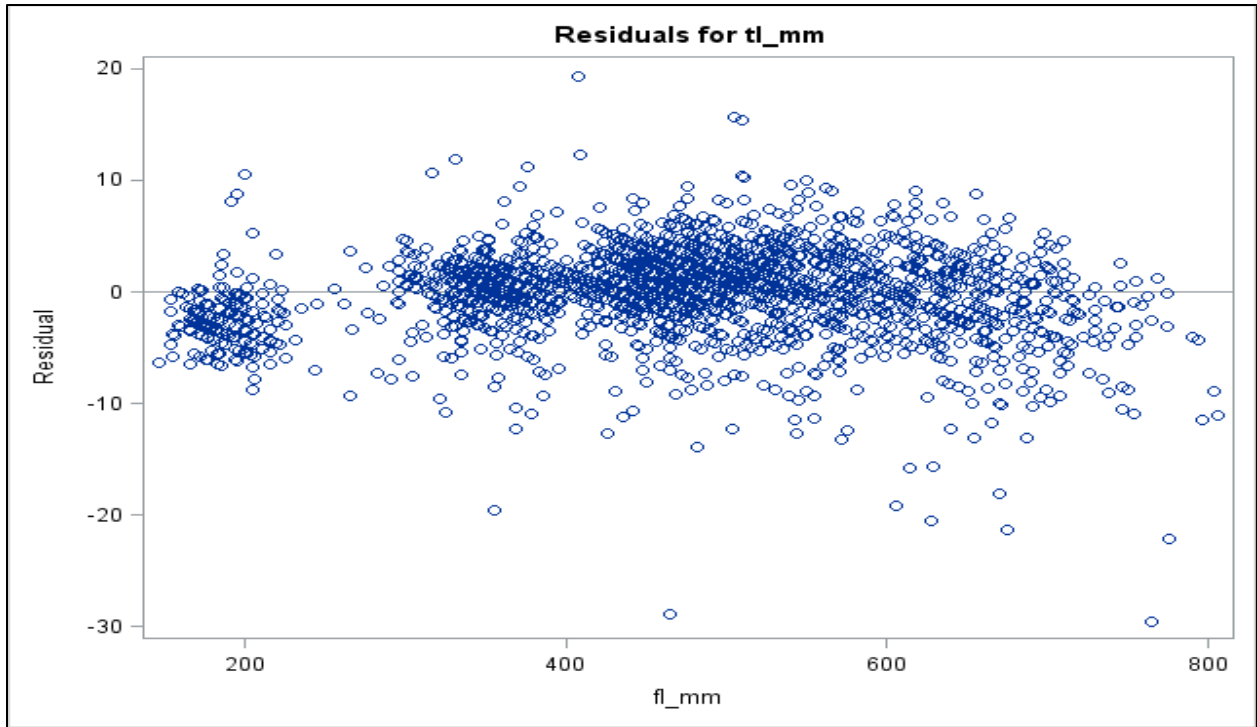
**Figure 3.** CSMA residual plot validating assumptions of normality.



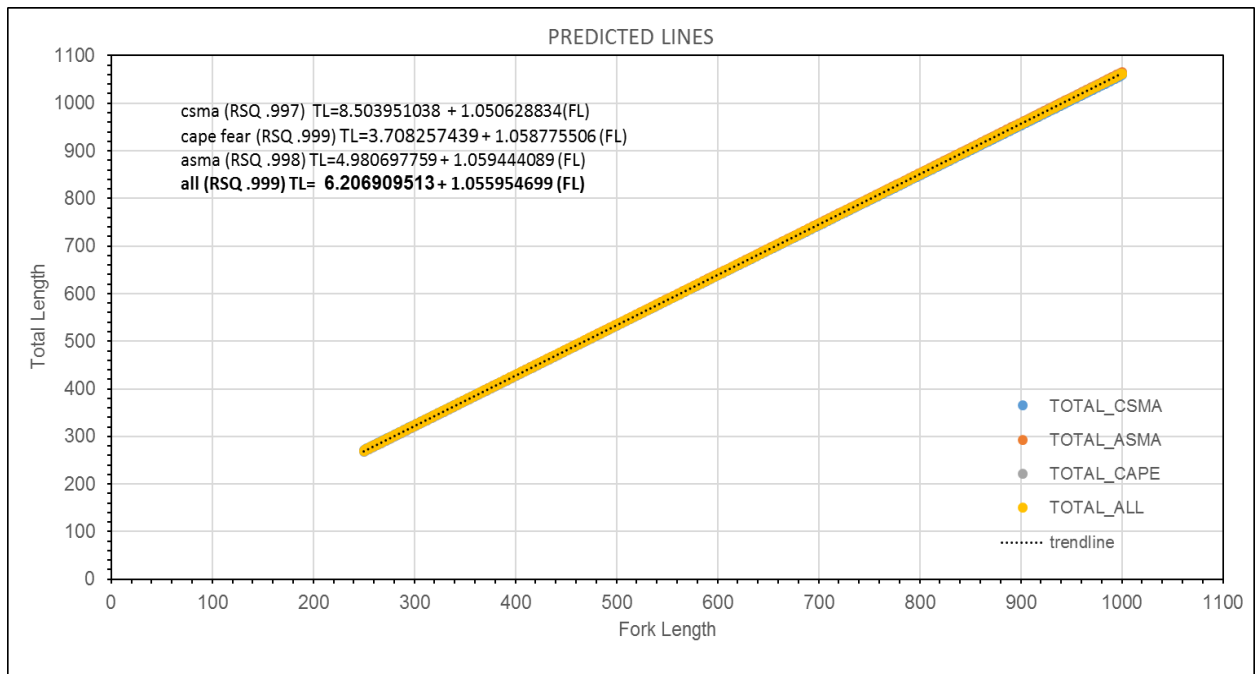
**Figure 4.** CSMA residuals demonstrate low variability associated with the best fit line.



**Figure 5.** Cape Fear residual plot validating assumptions of normality.



**Figure 6.** Cape Fear residuals demonstrate low variability associated with the best fit line (number of observations=2,372).



**Figure 7.** Regression analyses show strong relationships for CSMA, Cape Fear, ASMA, and all areas combined. The high RSQ value indicates a strong fit.

**13 APPENDIX B**



ARTICLE

## Exploring Causal Factors of Spawning Stock Mortality in a Riverine Striped Bass Population

Kyle T. Rachels\* and Benjamin R. Ricks

North Carolina Wildlife Resources Commission, 1721 Mail Service Center, Raleigh, North Carolina 27699-1700, USA

---

### Abstract

The recovery of the Atlantic Striped Bass *Morone saxatilis* stock in the 1990s is an important example of effective natural resources management. Implementation of Atlantic States Marine Fisheries Commission (ASMFC) harvest regulations reduced mortality, protected older and more fecund females, and contributed to the formation of dominant year-classes in the 1980s and 1990s. However, Striped Bass stocks south of Albemarle Sound, North Carolina, are not subject to ASMFC management plans, and many populations have failed to attain recovery goals. Catch-curve analyses indicate that the Neuse River Striped Bass population continues to experience spawning stock exploitation rates similar to those implicated in the decline of the Atlantic Migratory and Albemarle Sound/Roanoke River stocks in the 1970s. From 1994 to 2015, Striped Bass instantaneous fishing mortality ( $F$ ) in the Neuse River ranged from 0.12 to 0.84 and exceeded the overfishing threshold ( $F_{Threshold} = 0.41$ ) in 12 of 22 years. A global linear model using environmental and exploitation factors accounted for 55% of the variability in spawning stock discrete annual mortality. An information-theoretic approach was used to elucidate the best linear model for predicting discrete annual mortality. The best model included previous-year gill-net effort and same-year commercial harvest (Akaike weight = 0.64,  $R^2 = 0.50$ ). Model-averaged coefficients for gill-net effort and commercial harvest suggested total exploitation impacts that were congruent with other studies of Neuse River Striped Bass. Results indicate that reducing exploitation to target levels will require substantial reductions in gill-net effort in areas of the Neuse River where Striped Bass occur. Reducing exploitation may increase spawning stock biomass and advance the age structure of spawning females, conferring an increased likelihood of successful recruitment and production of dominant year-classes during periods of favorable environmental conditions.

---

Striped Bass *Morone saxatilis* populations sustained severe declines in abundance throughout the U.S. Atlantic coast in the 1970s after several years of record commercial harvest combined with poor recruitment (Boreman and Austin 1985; Richards and Deuel 1987). In North Carolina, Striped Bass commercial landings declined by 80% between 1973 and 1983 (Boreman and Austin 1985). Recovery efforts began with the development of the Atlantic States Marine Fisheries Commission's (ASMFC) Interstate Fisheries Management Plan

for Striped Bass (IFMP) in 1981 (Richards and Rago 1999). A centerpiece of the IFMP and its amendments was the use of harvest restrictions to curtail overexploitation. The harvest provisions of the IFMP were implemented in North Carolina beginning in 1984, along with an expansion of Striped Bass stocking programs and continued development of optimized streamflow releases from Roanoke Rapids Dam to improve spawning conditions in the Roanoke River, North Carolina (Figure 1; NCDENR 2004, 2013). Albemarle Sound/Roanoke River

---

Subject editor: Debra J. Murie, University of Florida, Gainesville

\*Corresponding author: kyle.rachels@ncwildlife.org  
Received January 11, 2018; accepted June 18, 2018

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Striped Bass were declared recovered in 1997 (NCDENR 2004).

In North Carolina, Striped Bass populations south of Albemarle Sound (Figure 1) are not subject to compliance with ASMFC management plans due to their minimal contribution to the Atlantic Migratory stock (Merriman 1941; Greene et al. 2009). These populations are collectively managed as the Central Southern Management Area (CSMA) stock under a collaborative agreement by the North Carolina Division of Marine Fisheries (NCDMF; coastal waters) and the North Carolina Wildlife Resources Commission (NCWRC; inland waters). Of the populations comprising the CSMA, Neuse River Striped Bass were among the first to receive targeted monitoring and management actions (Hammers et al. 1995).

Although Striped Bass are documented as historically utilizing all major coastal North Carolina rivers (Smith 1907), the Neuse River population was among the most studied by early ichthyologists. In the 19th century, the population was subject to the second-largest Striped Bass fishery in North Carolina after the fisheries operating on the Albemarle Sound/Roanoke River stock. Yarrow (1877) described Striped Bass in the Neuse River as “exceedingly plenty” and reported that 3,000 were sold to New Bern (Figure 1) fish houses from January to April 1873 (Yarrow 1874). By 1880, almost 16,000 Striped Bass

were harvested and shipped from New Bern to northern cities, with an additional unknown quantity consumed locally during the fishing season (McDonald 1884). Despite their former abundance, declines were evident before the end of the 19th century, leading McDonald (1884) to note that “...the supply has materially decreased...owing to overfishing and the erection of obstructions.” By 1939, only 318 kg of Striped Bass were commercially harvested in Craven County (Figure 1; Chestnut and Davis 1975).

Although fishing records during World War II are sparse, acquisition of fishing vessels and labor for the war effort likely reduced Striped Bass harvest and allowed for stock rebuilding. Fishing restrictions and labor shortages were eased toward the end of the war, leading to the harvest of 18,000 kg of Striped Bass in Craven County during 1945 (Anderson and Power 1949). However, construction of Quaker Neck Dam in 1952 prohibited access to essentially all spawning habitat (Burdick and Hightower 2006). By the mid-1960s, recreational and commercial anglers reported population declines, and a subsequent 3-year NCWRC survey collected only 12 adult fish (Miller 1975). Despite minimal harvest restrictions, commercial landings remained low throughout the latter half of the 20th century and did not exceed 4,500 kg again until 2010 (NCDMF, unpublished data). It is possible that the intensity of post-war fishing in the lower Neuse River

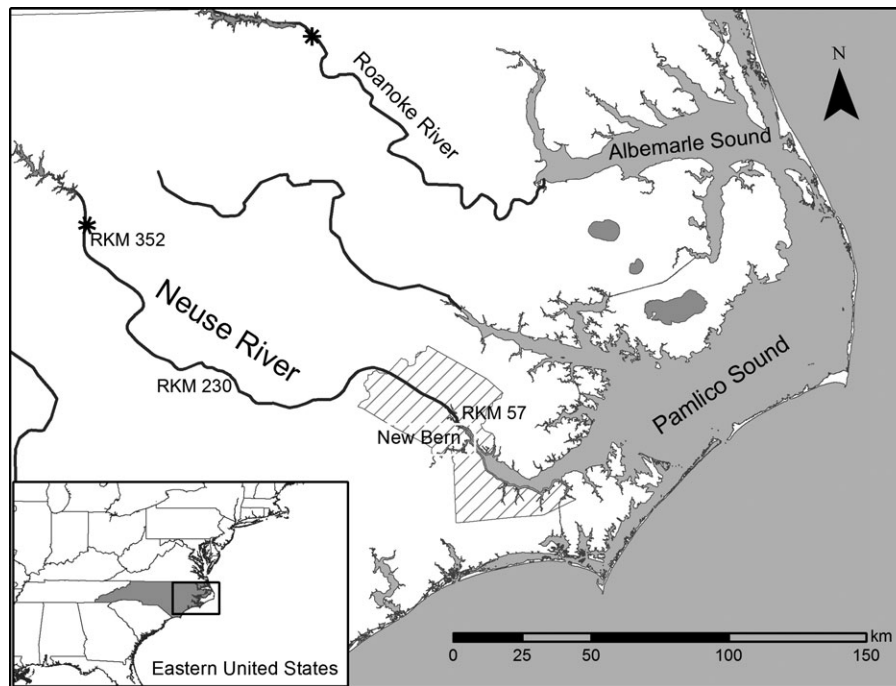


FIGURE 1. Coastal North Carolina, showing the Neuse River in relation to Pamlico Sound; RKM denotes river kilometers from the confluence of the Neuse River and Pamlico Sound. The first impediments to upstream migration (Milburnie Dam on the Neuse River; Roanoke Rapids Dam on the Roanoke River) are indicated by black asterisks. Gray diagonal lines denote Craven County.

combined with an inability to access suitable spawning habitat led to the near extirpation of the population.

Active management efforts in the Neuse River began with the implementation of an annual stocking regime in 1992 (although intermittent stocking began as early as 1931). In 1994, annual spawning ground surveys commenced, and a 11,340-kg commercial harvest quota was established for the entire CSMA stock (NCDENR 2004). The removal of Quaker Neck Dam in 1998 allowed unobstructed access to approximately 120 km of historical spawning habitat (Burdick and Hightower 2006). Finally, gill-net use was prohibited in NCWRC-managed inland waters in 2001 (NCDENR 2013).

Recovery efforts were first formalized in 2004 as part of the North Carolina Estuarine Striped Bass Management Plan (NCDENR 2004) that was jointly developed by NCDMF and NCWRC. Unweighted linearized catch-curve analyses of age structures collected on the Neuse River spawning grounds indicated that overfishing was occurring (NCDENR 2004), leading to the implementation of gill-net restrictions in 2008 (established minimum distance from shore and use of tie-downs during the closed harvest season; NCDENR 2013). A stock assessment conducted in 2010 using unweighted linearized catch curves again documented high mortality, but the assessment was deemed unsuitable for management use due to large confidence intervals around the mortality estimate. However, the need for continued conservation management measures was supported by truncated size and age distributions, low CPUE, and an absence of older fish in spawning ground samples. Albemarle Sound/Roanoke River spawning potential ratios of 45% and 40% were used to develop biological reference points for the Neuse River, resulting in an instantaneous fishing mortality rate ( $F$ ) target ( $F_{Target}$ ) of 0.33 and an overfishing threshold ( $F_{Threshold}$ ) of 0.41 (NCDENR 2013, 2014).

Electrofishing assessments on the spawning grounds indicate that size and age distributions have not expanded since the 2010 stock assessment (Rachels and Ricks 2015). Additionally, recent results utilizing parentage-based tagging (PBT) indicate that hatchery fish (Table 1) comprise at least two-thirds of the spawning stock (O'Donnell et al. 2016) and may approach 100% stocking contribution (Rachels and Ricks 2015; O'Donnell et al. 2016). The development of recommendations for catch-curve best practices (Smith et al. 2012) render former Neuse River Striped Bass stock assessments obsolete and present an opportunity to re-evaluate spawning ground age-structure data. Our objectives were two-fold: (1) to improve the precision of catch-curve mortality estimates by using current methodology and an expanded time series; and (2) to use linear modeling in an information-theoretic approach (Burnham and Anderson 2002) to elucidate factors responsible for driving the observed mortality rates.

## METHODS

*Study area.*—The Neuse River flows approximately 400 km from its origin at the confluence of the Eno and Flat rivers before discharging into Pamlico Sound, North Carolina (Figure 1). The lower 60 km constitute a wind-mixed mesohaline estuary, although salinity can range from 0‰ to 27‰ depending on precipitation and streamflow (Burkholder et al. 2006). The Neuse River estuary has been classified as “Nutrient Sensitive Waters” since 1988 (NCDENR 2006) and experienced numerous algae blooms and fish kills during the 1990s resulting from nitrogen and phosphorus inputs (Burkholder et al. 1995, 2006; Rothenberger et al. 2009).

*Mortality estimation.*—From 1994 to 2015, boat-mounted electrofishing (Smith-Root 7.5 GPP; 120 Hz; 5,000–7,000 W) was used to collect Striped Bass from the spawning grounds during annual spawning migrations (March–May). Collections primarily occurred between river kilometer (RKM) 230 of the Neuse River (measuring from its confluence with Pamlico Sound) and RKM 352. Few Striped Bass were collected above Quaker Neck Dam (RKM 230; Figure 1) before its removal in 1998.

Striped Bass were measured for TL (mm) and weighed (g), and sex was determined by applying pressure to the abdomen and observing the vent for discharge of milt or eggs. Scales for age estimation were removed from the left side of each fish between the dorsal fin and lateral line. From 1994 to 2014, 15 fish of each sex per 25-mm size-class were aged by either directly reading scales (1994–2010) or reading scale impressions on acetate slides (2011–2014). Since sampling occurred during the time of year when annuli are formed, scale age was based on (1) the actual number of annuli if an annulus was present on the scale margin; or (2) the number of annuli plus 1 if there was a considerable gap between the last annulus and the scale margin (NCWRC and NCDMF 2011). A 20% subsample of each size-class was aged by a second reader. Discrepancies between primary and secondary readers' estimates were resolved by jointly reading and reaching consensus (NCWRC and NCDMF 2011). In 2015, a partial pelvic fin clip from each fish was preserved in a 95% solution of ethyl alcohol to determine hatchery or wild origin using PBT. Hatchery-origin fish were aged using PBT, while fish of unknown origin were assigned ages with sex-specific age-length keys developed using scale-aged fish from 2010 to 2014.

The Chapman–Robson estimator was used to estimate instantaneous total mortality ( $Z$ ) for each year in the time series via the recommendations of Smith et al. (2012). As with other catch-curve methods, assumptions included the following: (1) the proportion of ages in the population is estimated without error, (2) recruitment varies without trend for all age-classes, (3) mortality is stationary through time and across age-classes, and (4) all age-classes are equally vulnerable to the sampling gear (Robson and Chapman 1961; Smith et al. 2012). Of the various catch-curve methods, the Chapman–Robson estimator is the most

TABLE 1. Number of hatchery-origin Striped Bass stocked into the Neuse River, North Carolina, and exploitation and environmental factors.

Year	Number stocked	Commercial effort (trips)	Commercial harvest (kg)	Summer dissolved oxygen (mg/L)	Summer water temperature (°C)
1994	182,990	2,531	3,760	7.1	27.5
1995	99,176	2,601	1,792	6.7	26.9
1996	200,760	3,018	3,159	6.5	28.0
1997	100,000	3,084	2,424	8.6	27.8
1998	290,925	3,209	2,511	6.3	27.9
1999	100,000	2,527	2,764	9.0	28.9
2000	229,993	3,030	2,181	6.6	27.3
2001	103,000	2,619	3,149	6.8	27.7
2002	147,654	3,317	1,869	9.5	29.1
2003	100,000	3,196	2,621	6.4	28.1
2004	268,011	2,159	3,547	7.3	28.5
2005	114,000	2,305	2,346	9.1	29.9
2006	245,935	2,777	3,216	7.7	28.1
2007	242,835	2,893	3,053	8.8	28.8
2008	313,798	1,980	2,190	9.7	29.6
2009	204,289	2,464	3,758	7.9	28.2
2010	107,142	1,583	5,092	8.0	30.1
2011	102,089	1,485	7,081	7.8	29.1
2012	140,358	1,577	1,946	6.2	27.8
2013	295,161	2,206	5,328	5.9	27.0
2014	158,730	1,603	2,801	6.7	28.2
2015	109,144	1,091	3,793	6.1	27.8

robust to violations of these assumptions (Murphy 1997; Smith et al. 2012). In accordance with Smith et al. (2012), age at full recruitment to the catch curve was the age of peak catch plus 1 year (peak-plus criterion). In addition, an overdispersion parameter  $c$  (Burnham and Anderson 2002; Smith et al. 2012) was calculated for each year to correct the SE of the mortality estimate and to assess structural fit of the Chapman–Robson estimator to the age-structure data ( $c > 4$  indicates poor model fit; Burnham and Anderson 2002). Instantaneous fishing mortality was calculated for each year by subtracting instantaneous natural mortality ( $M = 0.24$ ; Bradley 2016) from  $Z$ . Uncertainty in the mortality estimates was characterized by calculating the relative standard error (RSE;  $Z/SE$ ) and bootstrapping from the distributions of  $Z$  and  $M$  (Gamma distributed; Bolker 2008) to estimate 90% confidence intervals for  $F$ .

**Mortality modeling.**—Linear models were developed to evaluate environmental and exploitation factors that potentially influence discrete annual mortality ( $A = 1 - e^{-Z}$ ) over the time series 1994–2015, including summer dissolved oxygen, summer water temperature, gill-net effort, and commercial harvest. We hypothesized that low dissolved oxygen and warm summer temperatures may lead to increased natural mortality. Hypoxic conditions can be prevalent in the Neuse River estuary during the summer months as a result of nutrient loading and water column

stratification (Luettich et al. 2000; NCDENR 2001). These hypoxic conditions have been implicated in many of the 236 fish kills occurring between 1996 and 2015, which primarily affected Atlantic Menhaden *Brevoortia tyrannus* in the Neuse River basin (NCDENR 2001; NCDEQ 2015). Hypoxic events and resulting fish kills have also been implied as negatively affecting Striped Bass (NCDENR 2013). Water quality data were obtained from the Neuse River Estuary Modeling and Monitoring Project (ModMon; UNC 2016), which is one of the few programs that has continuously monitored water quality in the lower Neuse River since 1994. The summer (June–August) mean surface dissolved oxygen (mg/L) and summer mean surface water temperature (°C) at ModMon station 30 (RKM 57; Figure 1) were used as environmental factors. Results of an acoustic telemetry study (Bradley et al. 2018) determined that the highest densities of adult and juvenile Striped Bass occur in the vicinity of the selected ModMon station.

In addition to the suite of environmental factors, several long-term data sets were available from NCDMF to allow investigation of the effects of exploitation. Beginning in 1994, a mandatory trip ticket program was implemented to monitor commercial landings at the first point of sale. Information collected by this program includes harvest (kg) landed by species, gear type, and location (NCDENR 2013). Neuse River Striped Bass commercial

harvest was used as a direct exploitation factor (NCDMF, unpublished data). However, gill-net fisheries continue to pursue other marketable species after the Striped Bass harvest season is closed. Therefore, the annual number of gill-net trips in the Neuse River was used as a measure of gill-net effort that potentially accounts for harvest, discard, and unreported or misreported mortality (NCDMF, unpublished data). Unfortunately, measures of recreational fishing effort for Striped Bass were not available for the entire time series. A recreational creel survey has been conducted annually in the lower Neuse River since 2004, yet there is limited information for prior years (for exceptions, see Borawa 1983 and Rundle et al. 2004). Several recreational fishing surveys administered by National Oceanic and Atmospheric Administration Fisheries, including the Marine Recreational Information Program, the Marine Recreational Fisheries Statistics Survey, and the Coastal Household Telephone Survey, were investigated for potential use as a surrogate recreational fishing effort metric. However, these surveys lacked the data resolution necessary to specifically assess Neuse River recreational fisheries.

Since age-structure collections occurred in the spring (March–May), it was likely that factors occurring throughout the previous year (gill-net effort) or during the previous summer (dissolved oxygen and surface water temperature) had a greater influence on the estimated mortality rate than same-year measures. Therefore, these predictor variables were modeled using a 1-year time lag. Commercial harvest was not modeled with a time lag since the commercial Striped Bass harvest season occurs in the early spring before electrofishing collections on the spawning grounds; any effects of commercial harvest should be detected using same-year measures. Striped Bass discrete annual mortality was nonstationary; the global model was of the form

$$A'_t = \beta_0 + \sum(\theta_i X'_{i,t-1}) + \theta_C X'_{C,t} + \varepsilon_t,$$

where  $A$  = discrete annual mortality;  $\beta_0$  = intercept;  $X$  = variable  $i$ ;  $\theta_i$  = effect of variable  $X_i$ ;  $t$  = year;  $C$  = commercial harvest; and  $\varepsilon$  = an independently and identically distributed white noise vector. Note that  $A'_t$  and  $X'_{i,t}$  were first-differenced to ensure stationarity and remove serial correlation as given by

$$A'_t = A_t - A_{t-1}, \text{ and } X'_{i,t} = X_{i,t} - X_{i,t-1}.$$

In the case of four predictor variables, there are 15 main-effects models and 26 total models if we consider first-order interactions. Given our small sample size (22 observations) and the potential for “too many models” (Anderson and Burnham 2002; Burnham et al. 2011; Dochtermann and Jenkins 2011), we did not consider all-subsets regression. Instead, we constrained our analyses to 12 main-effects models (example R code provided in the

Supplement available separately online) incorporating dissolved oxygen, surface water temperature, gill-net effort, and commercial harvest using the information-theoretic framework described by Burnham and Anderson (2002). The second-order Akaike’s information criterion ( $AIC_c$ ) was computed for each model, and the difference in  $AIC_c$  value ( $\Delta_i$ ) from the model with the smallest  $AIC_c$  was used to assess the relative strength of the models. After ensuring that  $A'$  and  $X'$  differencing removed time trends ( $\beta_0 = 0$ ;  $\alpha = 0.05$ ), the intercept was removed from final models, and  $AIC_c$  and  $\Delta_i$  were recalculated. The reduced parameterization improved  $AIC_c$  for all models. Akaike weights ( $\omega_i$ ) were calculated to evaluate the relative likelihood of each model (Burnham and Anderson 2002). The relative importance of each predictor variable was assessed by decomposing global model variance using the Lindeman–Merenda–Gold (LMG) method (Grömping 2007). Model-averaged estimates of the effect of each predictor variable were calculated by multiplying the coefficients of each factor in the models in which they appeared by the  $\omega_i$  of that model (Burnham and Anderson 2002). The model-averaged effect for gill-net effort and commercial harvest was multiplied by the 1994–2015 mean number of gill-net trips and mean harvest, respectively, to estimate each factor’s long-term average effect on discrete annual mortality ( $\Delta A \equiv u$ ; discrete annual fishing mortality). Linear models were fitted using ordinary least-squares (OLS) regression with package “dynlm” in R version 3.2.5.

*Model assumptions.*—Assumptions for OLS time series regression depart in some respects from those considered in classical linear modeling. Assumptions of time series regression include a mean of zero, constant variance, and constant covariance structure through time (stationarity; Hyndman and Athanasopoulos 2014). The augmented Dickey–Fuller (ADF) test ( $\alpha = 0.05$ ; Hyndman and Athanasopoulos 2014) assumes  $H_0$  = nonstationary and was employed in the R package “stats” to assess stationarity in the mortality time series. The partial autocorrelation function (PACF; Derryberry 2014) in the “stats” package was utilized to examine the potential for autocorrelation in the spawning stock discrete annual mortality time series. Multicollinearity among the predictor variables was assessed by calculating variance inflation factors (VIFs; Fox and Weisberg 2011) in the R package “car.” Variance inflation factors are generally considered to indicate the presence of multicollinearity if any VIF exceeds 10 (see O’Brien 2007).

## RESULTS

### Mortality Estimation

The number of Striped Bass collected on the spawning grounds varied throughout the time series, ranging from

TABLE 2. Chapman–Robson mortality estimator metrics and mortality rates ( $Z$  = instantaneous total mortality rate;  $A$  = discrete annual mortality;  $F$  = instantaneous fishing mortality rate;  $\mu$  = discrete annual exploitation rate) for Neuse River Striped Bass, 1994–2015 ( $N$  = total catch;  $N_c$  = number in catch curve;  $T_c$  = age at recruitment to catch curve [peak-plus];  $c$  = overdispersion parameter; LCL = lower 90% confidence limit; UCL = upper 90% confidence limit; RSE = relative standard error).

Metric	Year of sample																					
	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015
$N$	120	221	226	143	219	292	357	155	102	403	90	125	58	172	141	373	141	176	144	341	311	239
$N_c$	36	107	71	81	148	151	111	69	67	98	48	97	21	96	24	231	71	55	67	106	129	95
$T_c$	7	5	6	5	4	5	5	5	4	6	6	4	4	4	6	4	4	5	4	5	5	5
$c$	0.57	1.60	1.10	2.23	1.64	1.97	4.17	3.90	5.53	2.20	0.12	1.81	1.57	2.52	3.19	4.96	0.13	2.41	0.85	0.42	1.51	0.90
$Z$	1.08	0.73	0.85	0.61	0.45	0.75	0.45	0.52	0.36	0.65	0.78	0.44	0.53	0.63	0.98	0.84	0.94	0.84	0.62	0.74	0.86	0.94
$SE_c$	0.19	0.09	0.11	0.10	0.05	0.09	0.09	0.13	0.10	0.10	0.12	0.06	0.15	0.10	0.37	0.13	0.12	0.18	0.08	0.07	0.10	0.10
$Z$ LCL	0.77	0.58	0.67	0.44	0.37	0.61	0.31	0.32	0.19	0.49	0.59	0.34	0.29	0.46	0.37	0.63	0.75	0.54	0.49	0.62	0.70	0.78
$Z$ UCL	1.39	0.88	1.03	0.77	0.53	0.90	0.60	0.73	0.53	0.82	0.97	0.54	0.77	0.80	1.59	1.05	1.13	1.14	0.75	0.87	1.02	1.11
RSE (%)	17	12	13	17	11	12	20	24	29	15	15	14	28	16	38	15	12	22	12	10	11	11
$A$	0.66	0.52	0.57	0.45	0.36	0.53	0.36	0.41	0.30	0.48	0.54	0.36	0.41	0.47	0.62	0.57	0.61	0.57	0.46	0.53	0.58	0.61
$A$ LCL	0.54	0.44	0.49	0.35	0.31	0.45	0.26	0.27	0.17	0.39	0.44	0.29	0.25	0.37	0.31	0.47	0.53	0.42	0.39	0.46	0.50	0.54
$A$ UCL	0.75	0.58	0.64	0.54	0.41	0.59	0.45	0.52	0.41	0.56	0.62	0.42	0.53	0.55	0.80	0.65	0.68	0.68	0.53	0.58	0.64	0.67
$F$	0.84	0.48	0.61	0.36	0.21	0.51	0.21	0.28	0.11	0.41	0.53	0.20	0.28	0.38	0.73	0.59	0.69	0.59	0.37	0.50	0.61	0.70
$F$ LCL	0.49	0.23	0.34	0.10	-0.02	0.25	-0.04	-0.01	-0.15	0.15	0.26	-0.03	0.02	0.11	0.17	0.31	0.43	0.25	0.12	0.26	0.36	0.44
$F$ UCL	1.20	0.71	0.86	0.60	0.38	0.73	0.43	0.55	0.36	0.64	0.79	0.39	0.59	0.61	1.44	0.86	0.95	0.96	0.58	0.70	0.84	0.93
$\mu$	0.51	0.34	0.41	0.27	0.17	0.36	0.17	0.22	0.10	0.30	0.37	0.16	0.22	0.28	0.47	0.40	0.45	0.40	0.28	0.35	0.41	0.45

58 fish in 2006 to 403 fish in 2003 (Table 2). Scale ages were reasonably precise, as scale readers had a high rate of agreement within 1 year of age (87–100%; NCWRC, unpublished data). Recruitment to the catch curve typically occurred at age 4 or age 5. Although the oldest Striped Bass encountered on the spawning grounds was an age-13 female collected in 2005, only 73 (1.6%) of the 4,549 fish collected during the time series were age 9 or older.

The Chapman–Robson mortality estimator generally performed well, as  $c$  was greater than 4 in only 3 of 22 years (Table 2). Mortality estimates were reasonably precise (RSE < 30%) and only exhibited a high degree of

uncertainty in 2008. Instantaneous total mortality  $Z$  varied considerably throughout the time series, ranging from 0.36 to 1.08. Mortality was generally lowest during 1997–2007 and highest during 2008–2011. Values of  $F$  ranged from 0.12 to 0.84 (Table 2; Figure 2), assuming that the  $M$  given by Bradley (2016) remained constant throughout the time series. Fishing mortality was greater than  $F_{Threshold}$  in 12 of the 22 years.

**Mortality Modeling**

*Model assumptions.*—The ADF test indicated that spawning stock discrete annual mortality was nonstationary ( $P = 0.181$ ). Therefore, all modeled variables were first-differenced (Hyndman and Athanasopoulos 2014). The PACF indicated a correlation of 0.34 between  $A_t$  and  $A_{t-1}$ , suggesting weak autocorrelation. We did not consider this level of autocorrelation sufficient to warrant modeling as a first-order autoregressive process given the small sample size and the potential for model overspecification. The VIFs ranged from 1.1 to 2.5, indicating a low likelihood of multicollinearity among predictor variables.

*Model results.*—The best linear model supported by the data contained gill-net effort and commercial harvest as predictors of discrete annual mortality (Table 3). The global model containing all predictor variables accounted for 55% of the variability in spawning stock mortality, while the best model accounted for 50%. Every model receiving at least modest support as the best model ( $\Delta_i < 7$ ) incorporated gill-net effort as a predictor variable.

Gill-net effort was the most important predictor of spawning stock mortality relative to the four predictor variables examined (Table 4; Figure 3). Commercial harvest was the second most important predictor of spawning stock mortality, while summer dissolved oxygen and

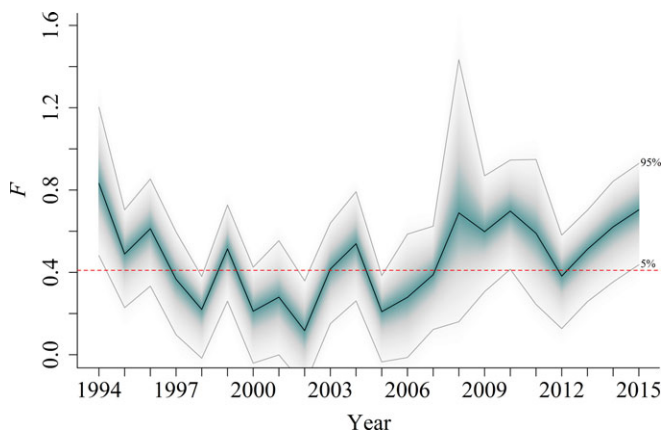


FIGURE 2. Striped Bass spawning stock fishing mortality ( $F$ ) in the Neuse River, North Carolina, during 1994–2015. The 90% confidence interval is denoted by gray lines, while the interquartile range is within a green color gradient. The dashed red line represents the overfishing threshold ( $F_{Threshold} = 0.41$ ).

TABLE 3. Linear models exploring the effect of environmental and exploitation factors on Striped Bass spawning stock discrete annual mortality, 1994–2015 (EFFORT = gill-net effort; DO = dissolved oxygen; HARV = commercial harvest; TEMP = surface water temperature). The number of estimated model parameters ( $K$ ) includes the predicting factors and an error term; final model runs did not include an intercept parameter. Akaike’s information criterion ( $AIC_c$ ), Akaike difference ( $\Delta_i$ ), Akaike weight ( $\omega_i$ ), and  $R^2$  are presented.

Model	$K$	$AIC_c$	$\Delta_i$	$\omega_i$	$R^2$
EFFORT, HARV	3	−39.95	0.00	0.64	0.50
EFFORT	2	−36.98	2.97	0.15	0.34
EFFORT, HARV, DO, TEMP	5	−34.88	5.07	0.05	0.55
EFFORT, DO	3	−34.81	5.14	0.05	0.36
EFFORT, TEMP	3	−34.60	5.36	0.04	0.35
EFFORT, DO, TEMP	4	−34.40	5.56	0.04	0.44
HARV	2	−31.68	8.27	0.01	0.14
DO	2	−30.67	9.29	0.01	0.09
HARV, DO	3	−30.38	9.57	0.01	0.20
HARV, TEMP	3	−29.83	10.12	0.00	0.10
DO, TEMP	3	−27.98	11.97	0.00	0.10
HARV, DO, TEMP	4	−27.23	12.72	0.00	0.20

surface water temperature did not substantially influence spawning stock mortality (Tables 3, 4). Multiplying the model-averaged gill-net coefficient by the mean number of gill-net trips for 1994–2015 (2,421 trips) suggests the gill-net fishery mean discrete annual exploitation rate ( $u$ ) was 0.29. Using the same procedure for commercial harvest (3,199 kg) suggests commercial harvest  $u$  is 0.08.

**DISCUSSION**

Catch-curve methodologies recommended by Smith et al. (2012) considerably reduced uncertainty in the  $Z$ -estimates compared to previous Neuse River stock assessments. The SEs of  $Z$  in our study ranged from 0.05 to

0.37, compared to 0.06–0.61 in the most recent stock assessment (Table 11 in NCDENR 2013). Similarly, RSE exceeded 30% in only 1 of the 22 years in our study, compared to 13 of the 16 years in the previous stock assessment (NCDENR 2013).

The catch-curve analysis indicates that the Neuse River Striped Bass spawning stock has been subjected to overfishing throughout much of the last two decades. The 22-year mean  $F$  in this study ( $F = 0.46$ ) is similar to the 18-year mean rate ( $F = 0.47$ ) that preceded the depletion of Albemarle Sound/Roanoke River Striped Bass in the 1970s (Hassler et al. 1981; NCDENR 2013). These high  $F$ -values also approach the level of exploitation that was deemed a major factor in the Atlantic Striped Bass stock

TABLE 4. Relative importance of predictor variables affecting Striped Bass spawning stock mortality (Lindeman–Merenda–Gold [LMG] method).

Predictor variable	Model-averaged coefficient		Relative importance (LMG)
	$\theta$	SE	
Gill-net effort	$1.21 \times 10^{-4}$	$3.54 \times 10^{-5}$	0.62
Commercial harvest	$2.37 \times 10^{-5}$	$1.00 \times 10^{-5}$	0.23
Dissolved oxygen	$-1.73 \times 10^{-2}$	$1.63 \times 10^{-2}$	0.10
Surface water temperature	$2.50 \times 10^{-2}$	$2.71 \times 10^{-2}$	0.05

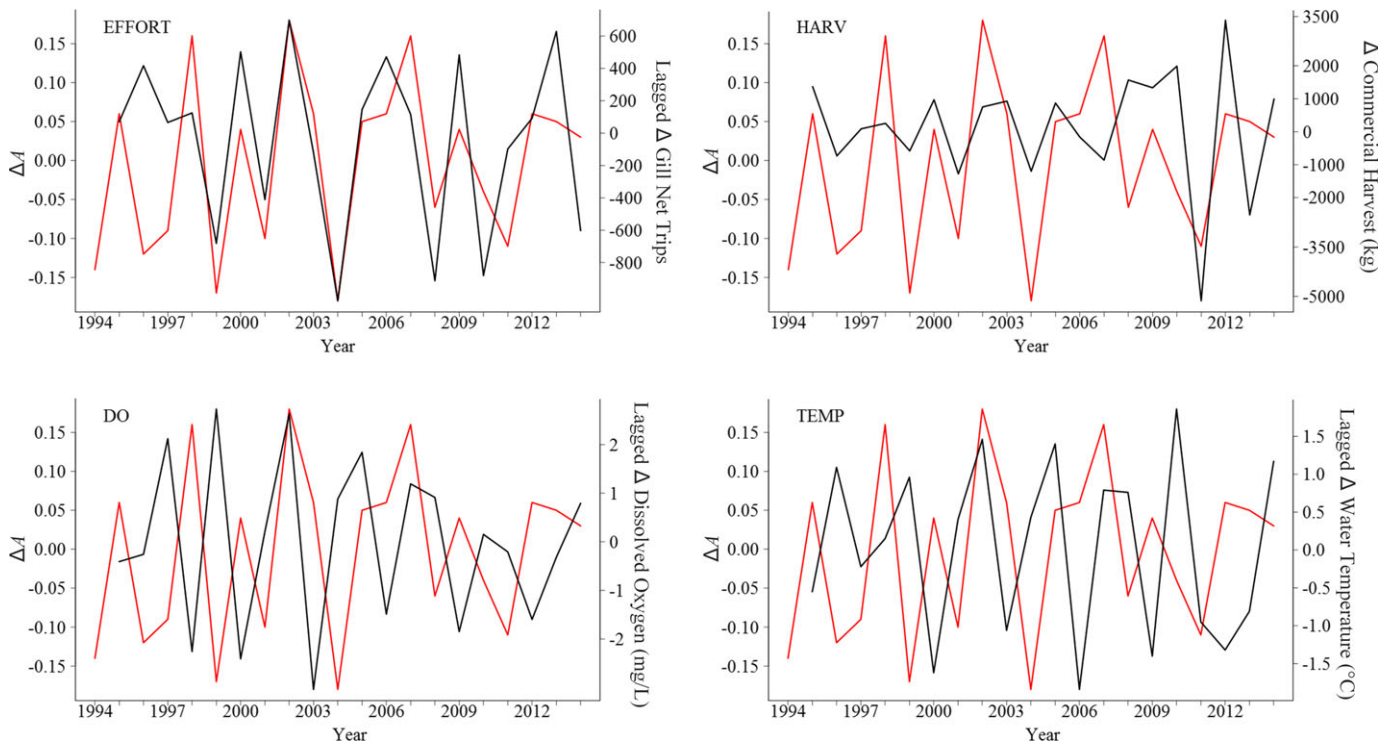


FIGURE 3. Differenced ( $\Delta$ ) Striped Bass spawning stock discrete annual mortality ( $A$ ; red) and differenced exploitation and environmental predictor variables (black) in the Neuse River, North Carolina (EFFORT = gill-net effort; DO = summer mean surface dissolved oxygen; HARV = commercial harvest; TEMP = summer mean surface water temperature). Gill-net effort, DO, and TEMP were modeled with 1-year time lags.



collapse (ASMFC 1989; Richards and Rago 1999). Mortality has not trended toward  $F_{Target}$  despite the development of two comprehensive management plans and increasingly restrictive recreational and commercial harvest regulations (see Appendix 14.5 in NCDENR 2013).

Linear modeling indicates that gill-net effort is the most important factor influencing spawning stock mortality among the exploitation and environmental factors examined. Gill-net effort accounted for substantially greater variability in spawning stock mortality than commercial harvest, and the model-averaged coefficient identified a discrete annual exploitation rate of 0.29 for gill net effort. This suggests that the commercial multispecies gill-net fishery imparts substantial mortality even when the Striped Bass harvest season is closed. The reason for this mortality is obscure, but it may be attributable to dead discard mortality; over-quota and high-grading mortality; avoidance, predation, and drop-out mortality; or unreported, misreported, and illegal harvest (ICES 1995; Gilman et al. 2013; Batsleer et al. 2015; Uhlmann and Broadhurst 2015). In particular, discard mortality should be carefully considered, as Clark and Kahn (2009) found that Striped Bass are acutely susceptible to discard mortality in multispecies gill-net fisheries. Furthermore, Striped Bass discards in the large-mesh gill-net fishery were identified as the primary source of mortality within the CSMA (NCDENR 2013). The effect of gill-net effort on discrete annual mortality as estimated by linear modeling was within 3% of the estimated effect of cryptic mortality in a cohort-based model ( $u = 0.26$ ; Table B.3 in Rachels and Ricks 2015), while the effect of commercial harvest was identical to the estimated discrete annual fishing mortality rate from commercial harvest in that study.

Contrary to exploitation factors, the environmental factors examined did not account for much variability in spawning stock mortality. Bradley et al. (2018) also failed to detect a relationship between dissolved oxygen, water temperature, and Striped Bass mortality between summer 2014 and summer 2015. Although numerous Atlantic Menhaden fish kills have occurred due to hypoxic conditions throughout the time period encompassing our research, it appears that these events have relatively little impact on Striped Bass spawning stock mortality. Campbell and Rice (2014) observed that estuarine fish can rapidly detect and avoid hypoxic areas in the Neuse River. However, they also found that habitat compression due to hypoxic conditions likely reduced growth rates in juvenile Spot *Leiostomus xanthurus* and Atlantic Croaker *Micropogonias undulatus*. Neuse River Striped Bass exhibit the fastest growth rates among coastal North Carolina Striped Bass populations (Rachels and Ricks 2015). It is likely that negative impacts of hypoxic conditions or water temperatures exceeding Striped Bass thermal optima would

manifest through reduced growth rates before mortality effects are observed. Nonetheless, the parameter coefficients for summer mean dissolved oxygen and summer mean surface water temperature indicate the potential for increased spawning stock mortality as dissolved oxygen decreases and water temperature increases. These effects were minimal—approximately 2% change in discrete annual mortality per unit change in temperature or dissolved oxygen—compared to the cumulative effects of gill-net effort and commercial harvest.

The inability to include recreational angling as an exploitation factor reduces the amount of variability in spawning stock mortality that can be accounted for in this study. The median annual recreational harvest during 2004–2015 was 2,337 kg and is similar to the median commercial harvest of 3,355 kg for the same time period (NCDMF, unpublished data). Thus, the actual commercial harvest and recreational harvest exploitation rates are similar, an observation supported by simulation studies (Rachels and Ricks 2015; Bradley 2016). It is likely that inclusion of factors that represent recreational harvest and discard would perform comparably to the results of the commercial harvest factor used in linear modeling. However, time-dynamic trends in the level of recreational fishing effort or harvest could influence its importance relative to commercial harvest in a regression analysis. In fact, recreational effort declined dramatically during 2005–2010, concurrent with increases in discrete annual mortality. The continued collection of recreational creel survey data is warranted to elucidate long-term effects of angling on Neuse River Striped Bass mortality.

Since the population is supported almost entirely by hatchery-origin fish, changes to stocking practices may affect recruitment and mortality estimation. Although the annual stocking goal is 100,000 phase-II (160–200 mm TL) Striped Bass, the actual stocking rate (Table 1) has varied (coefficient of variation = 46%) and has included phase-I fish (50 mm TL) in some years. Survival rates of phase-I and phase-II Striped Bass may be similar. Stocking practices in the nearby Cape Fear River are the same as those in the Neuse River, and phase-I and phase-II Striped Bass that were stocked at similar rates contributed almost equally to the Cape Fear River population (NCWRC, unpublished data). Additionally, the effect of variable recruitment on catch-curve mortality estimation has been extensively explored by others. Ricker (1975) determined that recruitment variation up to a factor of 5 did not prohibit catch-curve use so long as the variability was random. Similarly, Allen (1997) found that catch curves were useful for estimating mortality in populations that exhibited higher recruitment variation (55–84%) than the stocking variability observed in our study. Finally, although it does not yield insight into much of the entire time series of our data, our mortality estimates were very

similar to those reported by Bradley et al. (2018) for 2014–2015. The methodologies used in these studies (telemetry versus age structure) have different underlying assumptions, increasing confidence that mortality during the overlapping time periods was considerable.

Periodic strategists such as Striped Bass are resilient to periods of extended recruitment failure through the storage effect (Warner and Chesson 1985; Winemiller and Rose 1992). Recovery is contingent upon building spawning stock biomass by advancing the female age structure to older, more fecund fish (Secor 2000). Although regulating fishing mortality is one of the principal tools available to fisheries managers, “historical precedence is often invoked as a reason to continue unwise fishery management practices” (Richards and Rago 1999). However, the effectiveness of coordinated multi-jurisdictional management efforts in significantly reducing exploitation has been demonstrated by the restoration of the Atlantic Striped Bass stock (Field 1997; Richards and Rago 1999).

Current high exploitation rates combined with low stock abundance and a high contribution of hatchery fish to the spawning stock (Rachels and Ricks 2015; Bradley et al. 2018) suggest that the expected recovery time of Neuse River Striped Bass continues to be “both uncertain and long” (Hilborn et al. 2014). Our research indicates that fisheries managers should reduce exploitation by focusing on reductions in gill-net effort in areas of the Neuse River that are utilized by Striped Bass. Reducing spawning stock exploitation may confer an increased likelihood of recruitment during periods of favorable environmental conditions, thereby leading to improvements in population abundance and increased numbers of wild fish in the spawning stock.

## ACKNOWLEDGMENTS

We thank Joseph Hightower, Ken Pollock, and Paul Vos for their review and improvement of the statistical methods employed here. Kevin Dockendorf, Jeremy McCargo, and Chad Thomas provided guidance and positive critique throughout the development of the manuscript. We also appreciate the NCDMF and ModMon project for sharing information associated with their long-term data collection programs. This research was supported in part by the Federal Aid in Sport Fish Restoration Program (Project F-108). There is no conflict of interest declared in this article.

## REFERENCES

- Allen, M. S. 1997. Effects of variable recruitment on catch-curve analysis for crappie populations. *North American Journal of Fisheries Management* 17:202–205.
- Anderson, A. W., and E. A. Power. 1949. Fisheries statistics of the United States: 1945. U.S. Fish and Wildlife Service, Statistical Digest 18, Washington, D.C.
- Anderson, D. R., and K. P. Burnham. 2002. Avoiding pitfalls when using information-theoretic methods. *Journal of Wildlife Management* 66:912–918.
- ASMFC (Atlantic States Marine Fisheries Commission). 1989. Supplement to the ASMFC Striped Bass fisheries management plan—amendment 4. ASMFC, Fisheries Management Report 15, Washington, D.C.
- Batsleer, J., K. G. Hamon, H. M. J. van Overzee, A. D. Rijnsdorp, and J. J. Poos. 2015. High-grading and over-quota discarding in mixed fisheries. *Reviews in Fish Biology and Fisheries* 25:715–736.
- Bolker, B. M. 2008. Ecological models and data in R. Princeton University Press, Princeton, New Jersey.
- Borawa, J. C. 1983. Sport fishery harvest determination of the lower Neuse River basin area. North Carolina Wildlife Resources Commission, Coastal Fisheries Investigations, Final Report, Raleigh.
- Boreman, J., and H. M. Austin. 1985. Production and harvest of anadromous Striped Bass stocks along the Atlantic coast. *Transactions of the American Fisheries Society* 114:3–7.
- Bradley, C. E. 2016. Evaluation of juvenile and adult Striped Bass mortality and distribution, with implications for recovery efforts in the Neuse River, North Carolina. Master’s thesis. North Carolina State University, Raleigh.
- Bradley, C. E., J. A. Rice, D. D. Aday, J. E. Hightower, J. Rock, and K. L. Lincoln. 2018. Juvenile and adult Striped Bass mortality and distribution in an unrecovered coastal population. *North American Journal of Fisheries Management* 38:104–119.
- Burdick, S. M., and J. E. Hightower. 2006. Distribution of spawning activity by anadromous fishes in an Atlantic Slope drainage after removal of a low-head dam. *Transactions of the American Fisheries Society* 135:1290–1300.
- Burkholder, J., D. Dickey, C. Kinder, R. Reed, M. Mallin, M. McIver, L. Cahoon, G. Melia, C. Brownie, J. Smith, N. Deamer, J. Springer, H. Glasgow, and D. Toms. 2006. Comprehensive trend analysis of nutrients and related variables in a large eutrophic estuary. *Limnology and Oceanography* 51:463–487.
- Burkholder, J. M., H. B. Glasgow, and C. W. Hobbs. 1995. Fish kills linked to a toxic ambush-predator dinoflagellate: distribution and environmental conditions. *Marine Ecology Progress Series* 124:43–61.
- Burnham, K. P., and D. R. Anderson. 2002. Model selection and multimodel inference, 2nd edition. Springer-Verlag, New York.
- Burnham, K. P., D. R. Anderson, and K. P. Huyvaert. 2011. AIC model selection and multimodel inference in behavioral ecology: some background, observations, and comparisons. *Behavioral Ecology and Sociobiology* 65:23–35.
- Campbell, L. A., and J. A. Rice. 2014. Effects of hypoxia-induced habitat compression on growth of juvenile fish in the Neuse River estuary, North Carolina, USA. *Marine Ecology Progress Series* 49:199–213.
- Chestnut, A. F., and H. S. Davis. 1975. Synopsis of marine fisheries of North Carolina. University of North Carolina, Sea Grant Program, Publication UNC-SG-75-12, Raleigh.
- Clark, J. H., and D. M. Kahn. 2009. Amount and disposition of Striped Bass discarded in Delaware’s spring Striped Bass gill-net fishery during 2002 and 2003: effects of regulations and fishing strategies. *North American Journal of Fisheries Management* 29:576–585.
- Derryberry, D. R. 2014. Basic data analysis for time series with R. Wiley, Hoboken, New Jersey.
- Dochtermann, N. A., and S. H. Jenkins. 2011. Developing multiple hypotheses in behavioral ecology. *Behavioral Ecology and Sociobiology* 65:37–45.
- Field, J. D. 1997. Atlantic Striped Bass management: where did we go right? *Fisheries* 22(7):6–8.
- Fox, J., and S. Weisberg. 2011. An R companion to applied regression, 2nd edition. Sage Publications, Thousand Oaks, California.

- Gilman, E., P. Suuronen, M. Hall, and S. Kennelly. 2013. Causes and methods to estimate cryptic sources of fishing mortality. *Journal of Fish Biology* 83:766–803.
- Greene, K. E., J. L. Zimmerman, R. W. Laney, and J. C. Thomas-Blate. 2009. Atlantic coast diadromous fish habitat: a review of utilization, threats, recommendations for conservation, and research needs. Atlantic States Marine Fisheries Commission, Habitat Management Series Number 9, Washington, D.C.
- Grömping, U. 2007. Estimators of relative importance in linear regression based on variance decomposition. *American Statistician* 61:139–147.
- Hammers, B. E., M. B. Wynne, and A. E. Little. 1995. Striped Bass spawning stock survey for Neuse and Tar rivers, 1994–1995. North Carolina Wildlife Resources Commission, Federal Aid in Sport Fish Restoration, Project F-22, Final Report, Raleigh.
- Hassler, W. W., N. L. Hill, and J. T. Brown. 1981. The status and abundance of Striped Bass, *Morone saxatilis*, in the Roanoke River and Albemarle Sound, North Carolina, 1956–1980. North Carolina Department of Natural Resources and Community Development, Special Scientific Report 38, Morehead City.
- Hilborn, R., D. J. Hively, O. P. Jensen, and T. A. Branch. 2014. The dynamics of fish populations at low abundance and prospects for rebuilding and recovery. *ICES Journal of Marine Science* 71:2141–2151.
- Hyndman, R. J., and G. Athanasopoulos. 2014. *Forecasting: principles and practice*. OTexts, Melbourne, Australia.
- ICES (International Council for the Exploration of the Sea). 1995. Report of the study group on unaccounted mortality in fisheries. ICES, C.M. 1995/B:1, k, Copenhagen.
- Luettich, R., J. McNinch, H. Paerl, C. Peterson, J. Wells, M. Alperin, C. Martens, and J. Pinckney. 2000. Neuse River estuary modeling and monitoring project stage 1: hydrography and circulation, water column nutrients and productivity, sedimentary processes and benthic-pelagic coupling, and benthic ecology. Water Resources Research Institute, Report 325-B, Raleigh, North Carolina.
- McDonald, M. 1884. The rivers and sounds of North Carolina. Pages 625–637 in G. B. Goode, editor. *The fisheries and fishery industries of the United States*, section V. U.S. Commission of Fish and Fisheries, Washington, D.C.
- Merriman, D. 1941. Studies on the Striped Bass (*Roccus saxatilis*) of the Atlantic coast. *Fishery Bulletin of the Fish and Wildlife Service* 50:1–77.
- Miller, W. G. 1975. Biology of Striped Bass in the Tar-Pamlico and Neuse rivers. North Carolina Wildlife Resources Commission, Federal Aid in Fish Restoration, Project F-21-7, Final Report, Raleigh.
- Murphy, M. D. 1997. Bias in the Chapman-Robson and least-squares estimators of mortality rates for steady-state populations. U.S. National Marine Fisheries Service Fishery Bulletin 95:863–868.
- NCDENR (North Carolina Department of Environment and Natural Resources). 2001. North Carolina Division of Water Quality annual report of fish kill events 2001. NCDENR, Raleigh.
- NCDENR (North Carolina Department of Environment and Natural Resources). 2004. North Carolina estuarine Striped Bass fishery management plan. NCDENR, Morehead City.
- NCDENR (North Carolina Department of Environment and Natural Resources). 2006. Basinwide assessment report—Neuse River basin. NCDENR, Raleigh.
- NCDENR (North Carolina Department of Environment and Natural Resources). 2013. Amendment 1 to the North Carolina estuarine Striped Bass fishery management plan. NCDENR, Morehead City.
- NCDENR (North Carolina Department of Environment and Natural Resources). 2014. November 2014 revision to amendment 1 to the North Carolina estuarine Striped Bass fishery management plan. NCDENR, Morehead City.
- NCDEQ (North Carolina Department of Environmental Quality). 2015. North Carolina Division of Water Resources annual report of fish kill events 2015. NCDEQ, Raleigh.
- NCWRC (North Carolina Wildlife Resources Commission) and NCDMF (North Carolina Division of Marine Fisheries). 2011. Estuarine Striped Bass in North Carolina: scale aging methods. NCWRC, Federal Aid in Sport Fish Restoration, Project F-22, Final Report, Raleigh.
- O'Brien, R. M. 2007. A caution regarding rules of thumb for variance inflation factors. *Quality and Quantity* 41:673–690.
- O'Donnell, T., D. Farrae, and T. Darden. 2016. 2015 Striped Bass genotyping and parentage analyses. North Carolina Wildlife Resources Commission, Federal Aid in Sport Fish Restoration, Project F-108, Final Report, Raleigh.
- Rachels, K. T., and B. R. Ricks. 2015. Neuse River Striped Bass monitoring programs, population dynamics, and recovery strategies. North Carolina Wildlife Resources Commission, Federal Aid in Sport Fish Restoration, Project F-108, Final Report, Raleigh.
- Richards, R. A., and D. G. Deuel. 1987. Atlantic Striped Bass: stock status and the recreational fishery. *Marine Fisheries Review* 49:58–66.
- Richards, R. A., and P. J. Rago. 1999. A case history of effective fishery management: Chesapeake Bay Striped Bass. *North American Journal of Fisheries Management* 19:356–375.
- Ricker, W. E. 1975. Computation and interpretation of biological statistics of fish populations. *Fisheries Research Board of Canada Bulletin* 191.
- Robson, D. S., and D. G. Chapman. 1961. Catch curves and mortality rates. *Transactions of the American Fisheries Society* 90:181–189.
- Rothenberger, M. B., J. M. Burkholder, and C. Brownie. 2009. Long-term effects of changing land use practices on surface water quality in a coastal river and lagoonal estuary. *Environmental Management* 44:505–523.
- Rundle, K. R., C. T. Waters, and R. D. Barwick. 2004. Neuse River basin sport fishery creel survey, 2002–2003. North Carolina Wildlife Resources Commission, Federal Aid in Sport Fish Restoration, Project F-22, Final Report, Raleigh.
- Secor, D. H. 2000. Spawning in the nick of time? Effect of adult demographics on spawning behavior and recruitment in Chesapeake Bay Striped Bass. *ICES Journal of Marine Science* 57:403–411.
- Smith, H. M. 1907. *The fishes of North Carolina*. E. M. Uzzell State Printers and Binders, Raleigh, North Carolina.
- Smith, M. W., A. Y. Then, C. Wor, G. Ralph, K. H. Pollock, and J. M. Hoenig. 2012. Recommendations for catch-curve analysis. *North American Journal of Fisheries Management* 32:956–967.
- Uhlmann, S. S., and M. K. Broadhurst. 2015. Mitigating unaccounted fishing mortality from gillnets and traps. *Fish and Fisheries* 16:183–229.
- UNC (University of North Carolina). 2016. Neuse River Estuarine Modeling and Monitoring Project [database]. UNC, Chapel Hill. Available: <http://www.unc.edu/ims/neuse/modmon/data.php>. (July 2018).
- Warner, R. R., and P. L. Chesson. 1985. Coexistence mediated by recruitment fluctuations: a field guide to the storage effect. *American Naturalist* 125:769–787.
- Winemiller, K. O., and K. A. Rose. 1992. Patterns of life-history diversification in North American fishes: implications for population regulation. *Canadian Journal of Fisheries and Aquatic Sciences* 49:2196–2218.
- Yarrow, H. C. 1874. Report of a reconnaissance of the shad-rivers south of the Potomac. Pages 396–402 in S. F. Baird, editor. *Report of the Commissioner for 1872 and 1873*. U.S. Commission of Fish and Fisheries, Washington, D.C.
- Yarrow, H. C. 1877. Notes on the natural history of Fort Macon, N.C., and vicinity (number 3). *Proceedings of the Academy of Natural Sciences of Philadelphia* 29:203–218.

## SUPPORTING INFORMATION

Additional supplemental material may be found online in the Supporting Information section at the end of the article.



ROY COOPER  
*Governor*

MICHAEL S. REGAN  
*Secretary*

STEPHEN W. MURPHEY  
*Director*

Oct. 30, 2020

## MEMORANDUM

**TO:** N.C. Marine Fisheries Commission

**FROM:** Laura Lee, Lead Stock Assessment Scientist  
Charlton Godwin, Biologist Supervisor, Estuarine Striped Bass FMP Co-Lead  
Fisheries Management Section

**SUBJECT:** Updates on 2020 Benchmark Albemarle-Roanoke Striped Bass Stock  
Assessment, Adaptive Management under Amendment 1 of the Estuarine  
Striped Bass FMP, and Amendment 2 Development

---

### Issue

Review of the N.C. Estuarine Striped Bass Fishery Management Plan (FMP) is underway. To begin review of Amendment 2 to the FMP the division and Wildlife Resources Commission (WRC) staff conducted a stock assessment of Albemarle-Roanoke (A-R) striped bass. This memo provides a summary of: 1) results for the 2020 benchmark A-R striped bass stock assessment, 2) information on adaptive management contained in Amendment 1 and the 2020 Revision to the N.C. Estuarine Striped Bass FMP, and 3) progress towards developing Amendment 2.

### Findings

- The benchmark A-R stock assessment passed an external peer review process in June of 2020. Results showed the A-R striped bass stock is overfished and overfishing is occurring in the terminal year (2017) of the assessment.
- Adaptive management contained in Amendment 1 to the N.C. Estuarine Striped Bass FMP states: “*should the target F [fishing mortality] be exceeded, then restrictive measures will be imposed to reduce F to the target level*”.
- Target F has been exceeded triggering the Revision to Amendment 1 implementing restrictive measures to reduce F to the target.
- Beginning in January 2021 a reduction in the A-R striped bass total allowable landings (TAL) to 51,126 lb will be implemented and will remain in place until the adoption of Amendment 2.
- The division is planning an update to the stock assessment in 2023 with data through 2022 to reassess stock conditions.
- Adequate river flows and sufficient spawning stock biomass (SSB) are both needed for successful spawning. Even at high levels of SSB, if river flows during the spawning season are not within recommended ranges successful spawning will not occur, which in turn leads to population decline regardless of the amount of fishing mortality (*F*).

### Action Needed

For informational purposes only, **no action is needed at this time.**

## Overview

### Results from the 2020 benchmark Albemarle-Roanoke Striped Bass Stock Assessment

Results from the 2020 benchmark A-R stock assessment indicate the stock is overfished and overfishing is occurring. Benchmark assessments involve a full analysis and review of the stock, including consideration of data inputs, new or improved assessment models, and recalculation of the Biological Reference Points (BRPs). The BRPs for this assessment are listed below in Table 1, along with the estimates of fishing mortality and spawning stock biomass from the terminal year of the assessment. The estimate of fishing mortality ( $F$ ) in the terminal year of the assessment (2017) was 0.27, which is above the  $F$  Threshold of 0.18. The estimate of SSB was 78,576 lb, which is below the SSB Threshold of 267,390 lb.

Table 1. Biological reference points (BRPs) and the 2017 estimate of fishing mortality ( $F$ ) and spawning stock biomass (SSB) from the 2020 Albemarle-Roanoke striped bass benchmark assessment.

Biological Reference Points (lb)		Terminal Year (2017) Estimate
$F$ Target	0.13	$F = 0.27$
$F$ Threshold	0.18	
SSB Target	350,371 lb	SSB = 78,576 lb
SSB Threshold	267,390 lb	

### Adaptive Management actions required under Amendment 1 to the N.C. Estuarine Striped Bass FMP to lower fishing mortality to the target

Implementing a new lower harvest level accomplishes the adaptive management directive in Amendment 1 to the North Carolina Estuarine Striped Bass FMP and maintains compliance with ASMFC's Addendum IV to Amendment 6 to the Interstate FMP for Atlantic Striped Bass. This same directive resulted in the November 2014 Revision to Amendment 1 that reduced the TAL from 550,000 lb to 275,000 lb based on projections starting from the terminal year (2013) of the then most recent assessment.

The Revision to Amendment 1 will be implemented a lower TAL effective January 2021 to reduce  $F$  to the  $F$  target. A 57% reduction in total removals relative to 2017 total removals is needed to bring  $F$  back to the  $F$  target and discards are already accounted for in the landings calculation. Total removals in 2017 included 119,244 lb of harvest and 23,795 lb of discards. Table 2 shows the TAL required to bring  $F$  back to the target in one year. During 2021 and 2022, harvest for the Albemarle-Roanoke fisheries will be monitored and controlled to keep harvest below the 51,216 lb TAL.

Table 2. The total allowable landings (TAL) necessary to reduce fishing mortality back to the target. Implementation date January 1, 2021.

<b>Total Allowable Landings</b>	<b>51,216 lb</b>
<b>Fleet</b>	<b>TAL (lb)</b>
ASMA commercial	25,608 lb
ASMA recreational	12,804 lb
RRMA recreational	12,804 lb

### River flow and population decline

One important factor to note, which was pointed out by the peer reviewers, is it appears there are other reasons for the decline in SSB other than just removals due to fishing. Poor recruitment (the number of age-0 fish coming into the population each year) could be the main reason for the population decline. Flow on the Roanoke River during egg and larval development during the month of May plays an important role in successful year-class production. Flows that are low to moderate are favorable for abundant year-class production while very high flows are almost always detrimental to year-class production because larvae flow out of the banks of the river into the floodplain where survival is low. After many years of above-average recruitment (1993-2000), the population experienced several poor and missing year classes (2003, 2004, 2009, 2013), and four recent poor year classes (2017, 2018, 2019 and again in 2020). It is apparent from the results of fishery-independent monitoring and results from the stock assessment that even in years when SSB is well above the SSB target, if flows are not within the recommended range, successful spawning will not occur. In short, adequate flows and sufficient SSB are both needed for successful spawning. It is also evident from previous stock performance and results from the most recent assessment, that SSB levels can increase dramatically with just a couple of years of successful spawning events .

### Amendment 2 to the N.C. Estuarine Striped Bass FMP

The November 2020 Revision to Amendment 1 to the N.C. Estuarine Striped Bass FMP is separate from and will not impede timing of the development of Amendment 2. Development of Amendment 2 is ongoing. The scoping period was Nov. 2-15, 2020 and public meetings were held Nov. 5 and 9, 2020. Results of the scoping period, the draft Goals and Objectives of Amendment 2, and a request for any additional management strategies to be considered will be brought before the MFC at its February 2021 business meeting. Under the provisions of the Fisheries Reform Act, Amendment 2 management strategies for the Albemarle-Roanoke stock will be adopted to address the overfished/overfishing condition. Adaptive management measures are also likely to be included in Amendment 2 to provide needed flexibility to account for changing stock conditions. The division is planning an update to the stock assessment in 2023 with data through 2022 to reassess stock conditions and update the TAL.

### ***For more information, please refer to the full documents included in the briefing materials:***

- Assessment of the Albemarle Sound-Roanoke River Striped Bass (*Morone saxatilis*) in North Carolina, 1991-2017
- November 2020 Revision to Amendment 1 to the N.C. Estuarine Striped Bass Fishery Management Plan
- Scoping document for Amendment 2 to the N.C. Estuarine Striped Bass Fishery Management Plan

**Assessment of the Albemarle Sound-Roanoke River  
Striped Bass (*Morone saxatilis*) in North Carolina, 1991–2017**

L.M. Lee, T.D. Tears, Y. Li, S. Darsee, and C. Godwin (editors)

August 2020

This document may be cited as:

Lee, L.M., T.D. Tears, Y. Li, S. Darsee, and C. Godwin (editors). 2020. Assessment of the Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*) in North Carolina, 1991–2017. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2020-01, Morehead City, North Carolina. 171 p.



## ACKNOWLEDGEMENTS

Members of the North Carolina Division of Marine Fisheries (NCDMF) Striped Bass Plan Development Team and their counterparts at the North Carolina Wildlife Resources Commission (NCWRC) were invaluable in providing assistance for the development of this stock assessment. Plan Development Team members from the NCDMF are Charlton Godwin (co-lead), Todd Mathes (co-lead), Katy West (mentor), Drew Cathey, Sean Darsee, David Dietz, Joe Facendola, Daniel Ipock, Laura Lee, Yan Li, Brian Long, Lee Paramore, Jason Peters, Jason Rock, Scott Smith, Chris Stewart, Thom Tears, Amanda Tong, Curt Weychert, and Chris Wilson. Members from the NCWRC are Jessica Baumann, Courtney Buckley, Kelsey Lincoln, Jeremy McCargo, Katy Potoka, Kyle Rachels, Ben Ricks, Kirk Rundle, Christopher Smith, and Chad Thomas. Thanks also to Kathy Rawls, NCDMF Fisheries Management Section Chief, and Catherine Blum, NCDMF Fishery Management Plan and Rulemaking Coordinator.

We are appreciative of Richard Methot (NOAA Fisheries) for his help in development of the Stock Synthesis stock assessment model.

We are especially grateful to the external peer reviewers for offering their time and effort to review this striped bass stock assessment: Jeff Kipp at the Atlantic States Marine Fisheries Commission, Dr. Michael Allen at the University of Florida, and Dr. Rod Bradford at the Department of Fisheries and Oceans Canada.

## EXECUTIVE SUMMARY

The North Carolina Fisheries Reform Act requires that fishery management plans be developed for the state's commercially and recreationally important species to achieve sustainable levels of harvest. Stock assessments are the primary tools used by managers to assist in determining the status of stocks and developing appropriate management measures to ensure the long-term viability of stocks.

The Albemarle Sound-Roanoke River (A-R) striped bass stock is managed jointly by the North Carolina Division of Marine Fisheries (NCDMF), the North Carolina Wildlife Resources Commission (NCWRC), and the South Atlantic Fisheries Coordination Office (SAFCO) of the U.S. Fish and Wildlife Service (USFWS) under guidelines established in the Atlantic States Marine Fisheries Commission (ASMFC) Interstate Fishery Management Plan (FMP) for Atlantic Striped Bass and the North Carolina Estuarine Striped Bass FMP. The Albemarle Sound Management Area (ASMA) includes Albemarle Sound and all of its joint and inland water tributaries, (except for the Roanoke, Middle, Eastmost, and Cashie rivers), Currituck Sound, Roanoke and Croatan sounds and all of their joint and inland water tributaries, including Oregon Inlet, north of a line from Roanoke Marshes Point to the north point of Eagle Nest Bay. The Roanoke River Management Area (RRMA) includes the Roanoke River and its joint and inland water tributaries, including Middle, Eastmost, and Cashie rivers, up to the Roanoke Rapids Lake Dam.

A forward-projecting statistical catch-at-age model was applied to data characterizing landings/harvest, discards, fisheries-independent indices, and biological data collected from the 1991 through 2017 time period. Both observed recruitment and model-predicted recruitment have been relatively low and declining in recent years. Fisheries-dependent and fisheries-independent data indicate a truncation of both length and age structure in recent years.

Reference point thresholds for the A-R striped bass stock were based on 35% spawner potential ratio (SPR). The estimated threshold for female spawning stock biomass (SSB;  $SSB_{\text{Threshold}}$  or  $SSB_{35\%}$ ) was 121 metric tons. Terminal year (2017) female SSB was 35.6 metric tons, which is less than the threshold value and suggests the stock is currently overfished ( $SSB_{2017} < SSB_{\text{Threshold}}$ ). The female SSB target ( $SSB_{\text{Target}}$  or  $SSB_{45\%}$ ) was 159 metric tons. The assessment model estimated a value of 0.18 for the threshold fishing mortality ( $F_{\text{Threshold}}$  or  $F_{35\%}$ ). The estimated value of fishing mortality in the terminal year (2017) of the model was 0.27, which is greater than the threshold value and suggests that overfishing is currently occurring in the stock ( $F_{2017} > F_{\text{Threshold}}$ ). The fishing mortality target ( $F_{\text{Target}}$  or  $F_{45\%}$ ) was estimated at a value of 0.13.

An independent, external peer review of this stock assessment approved the stock assessment for use in management for at least the next five years.

## TABLE OF CONTENTS

ACKNOWLEDGEMENTS .....	iii
EXECUTIVE SUMMARY .....	iv
LIST OF TABLES .....	vi
LIST OF FIGURES .....	viii
1 INTRODUCTION .....	14
1.1 The Resource .....	14
1.2 Life History .....	14
1.3 Habitat .....	19
1.4 Description of Fisheries .....	20
1.5 Fisheries Management .....	22
1.6 Assessment History .....	24
2 DATA .....	26
2.1 Fisheries-Dependent .....	26
2.2 Fisheries-Independent .....	34
3 ASSESSMENT .....	40
3.1 Method—Stock Synthesis .....	40
3.2 Discussion of Results .....	48
4 STATUS DETERMINATION CRITERIA .....	49
5 SUITABILITY FOR MANAGEMENT .....	50
6 RESEARCH RECOMMENDATIONS .....	51
7 LITERATURE CITED .....	53
8 TABLES .....	62
9 FIGURES .....	90
10 APPENDIX .....	166

## LIST OF TABLES

Table 1.1.	Parameter estimates and associated standard errors (in parentheses) of the von Bertalanffy age-length growth curve by sex. The function was fit to total length in centimeters. ....	62
Table 1.2.	Parameter estimates and associated standard errors (in parentheses) of the length-weight function by sex. The function was fit to total length in centimeters and weight in kilograms. ....	62
Table 1.3.	Percent maturity of female striped bass as estimated by Boyd (2011). ....	62
Table 1.4.	Age-constant estimates of natural mortality derived from life history characteristics. ....	63
Table 1.5.	Estimates of natural mortality at age by sex based on the method of Lorenzen (1996). ....	63
Table 1.6.	Changes in the total allowable landings (TAL) in metric tons and pounds (in parentheses) for the ASMA-RRMA, 1991–2017. ....	64
Table 1.7.	Striped bass commercial landings and discards and recreational harvest and discards from the ASMA-RRMA, 1991–2017. ....	65
Table 2.1.	Annual estimates of commercial gill-net discards (numbers of fish), 1991–2017. Note that values prior to 2012 were estimated using a hindcasting approach. ....	66
Table 2.2.	Annual estimates of recreational harvest and dead discards (numbers of fish) for the ASMA, 1991–2017. ....	67
Table 2.3.	Annual estimates of recreational harvest and dead discards (numbers of fish) for the RRMA, 1991–2017. Note that discard values prior to 1995 were estimated using a hindcasting approach. ....	68
Table 3.1.	Annual estimates of commercial landings and recreational harvest that were input into the SS model, 1991–2017. Values assumed for the coefficients of variation (CVs) are also provided. ....	69
Table 3.2.	Annual estimates of dead discards that were input into the SS model, 1991–2017. Values assumed for the coefficients of variation (CVs) are also provided. ....	70
Table 3.3.	GLM-standardized indices of relative abundance derived from fisheries-independent surveys that were input into the SS model, 1991–2017. The empirically-derived standard errors (SEs) are also provided. ....	71
Table 3.4.	Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds. ....	72
Table 3.4.	<i>(continued)</i> Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds. ....	73
Table 3.4.	<i>(continued)</i> Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds. ....	74
Table 3.4.	<i>(continued)</i> Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds. ....	75

Table 3.4.	( <i>continued</i> ) Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds. ....	76
Table 3.5.	Results of the base run compared to the results of 50 jitter trials in which initial parameter values were jittered by 10%. A single asterisk (*) indicates that the Hessian matrix did not invert. Two asterisks (**) indicate that the convergence level was greater than 1. ....	77
Table 3.5.	( <i>continued</i> ) Results of the base run compared to the results of 50 jitter trials in which initial parameter values were jittered by 10%. A single asterisk (*) indicates that the Hessian matrix did not invert. Two asterisks (**) indicate that the convergence level was greater than 1. ....	78
Table 3.6.	Results of the runs test for temporal patterns and results of the Shapiro-Wilk test for normality applied to the standardized residuals of the fits to the fisheries-independent survey indices from the base run of the assessment model. <i>P</i> -values were considered significant at $\alpha = 0.05$ . ....	79
Table 3.7.	Annual estimates of recruitment (thousands of fish), female spawning stock biomass (SSB; metric tons), and spawner potential ratio (SPR) and associated standard deviations (SDs) from the base run of the stock assessment model, 1991–2017. ....	80
Table 3.8.	Predicted population numbers (numbers of fish) at age at the beginning of the year from the base run of the stock assessment model, 1991–2017. ....	81
Table 3.9.	Predicted population numbers (numbers of fish) at age at mid-year from the base run of the stock assessment model, 1991–2017. ....	82
Table 3.10.	Predicted landings at age (numbers of fish) for the ARcomm fleet from the base run of the stock assessment model, 1991–2017. ....	83
Table 3.11.	Predicted dead discards at age (numbers of fish) for the ARcomm fleet from the base run of the stock assessment model, 1991–2017. ....	84
Table 3.12.	Predicted harvest at age (numbers of fish) for the ASrec fleet from the base run of the stock assessment model, 1991–2017. ....	85
Table 3.13.	Predicted dead discards at age (numbers of fish) for the ASrec fleet from the base run of the stock assessment model, 1991–2017. ....	86
Table 3.14.	Predicted harvest at age (numbers of fish) for the RRrec fleet from the base run of the stock assessment model, 1991–2017. ....	87
Table 3.15.	Predicted dead discards at age (numbers of fish) for the RRrec fleet from the base run of the stock assessment model, 1991–2017. ....	88
Table 3.16.	Annual estimates of fishing mortality (numbers-weighted, ages 3–5) and associated standard deviations (SDs) from the base run of the stock assessment model, 1991–2017. ....	89

## LIST OF FIGURES

Figure 1.1.	Boundary lines defining the Albemarle Sound Management Area, Central-Southern Management Area, and the Roanoke River Management Area. ....	90
Figure 1.2.	Fit of the age-length function to available age data for female striped bass.....	91
Figure 1.3.	Fit of the age-length function to available age data for male striped bass.....	91
Figure 1.4.	Fit of the length-weight function to available biological data for female striped bass.....	92
Figure 1.5.	Fit of the length-weight function to available biological data for male striped bass.....	92
Figure 1.6.	Estimates of natural mortality at age based on the method of Lorenzen (1996). ...	93
Figure 1.7.	Annual total landings/harvest in metric tons of striped bass from the ASMA and RRMA commercial and recreational sectors combined compared to the TAL, 1991–2017.....	93
Figure 2.1.	Annual commercial landings of striped bass in the ASMA-RRMA, 1962–2017. ....	94
Figure 2.2.	Annual length frequencies of striped bass commercial landings, 1982–2005.....	95
Figure 2.3.	Annual length frequencies of striped bass commercial landings, 2006–2017.....	96
Figure 2.4.	Annual age frequencies of striped bass commercial landings, 1982–2005. The age-15 bin represents a plus group.....	97
Figure 2.5.	Annual age frequencies of striped bass commercial landings, 2006–2017. The age-15 bin represents a plus group.....	98
Figure 2.6.	Management areas used in development of GLM for commercial gill-net discards. ....	99
Figure 2.7.	Ratio of commercial (A) live and (B) dead discards to commercial landings, 2012–2017.....	100
Figure 2.8.	Annual estimates of commercial gill-net discards, 1991–2017. Note that values prior to 2012 were estimated using a hindcasting approach. ....	101
Figure 2.9.	Annual length frequencies of striped bass commercial gill-net discards, 2004–2017.....	102
Figure 2.10.	Sampling zones and access sites of the striped bass recreational creel survey in the ASMA. ....	103
Figure 2.11.	Annual estimates of recreational harvest for the Albemarle Sound, 1991–2017..	104
Figure 2.12.	Annual estimates of recreational dead discards for the Albemarle Sound, 1991–2017.....	104
Figure 2.13.	Annual length frequencies of striped bass recreational harvest in the Albemarle Sound, 1996–2017. ....	105
Figure 2.14.	Annual length frequencies of striped bass recreational discards in the Albemarle Sound, 1997–2017. ....	106
Figure 2.15.	Map of angler creel survey interview locations in the RRMA, NC. The dashed line indicates the demarcation point between the upper and lower zones. Zone 1 access areas include (GA) Gaston (US HWY 48), (WE) Weldon, and (EF) Scotland Neck (Edwards Ferry US HWY 258). Zone 2 access areas include (HA) Hamilton, (WI) Williamston, (JA) Jamesville, (PL) Plymouth, (45) US HWY 45, (CC) Conaby Creek, and (SS) Sans Souci (Cashie River).....	107
Figure 2.16.	Ratio of recreational dead discards to recreational harvest in the Roanoke River, 1995–2017.....	108

Figure 2.17. Annual estimates of recreational harvest for the Roanoke River, 1982–2017. ....	108
Figure 2.18. Annual estimates of recreational dead discards for the Roanoke River, 1982–2017. Note that discard values prior to 1995 were estimated using a hindcasting approach. ....	109
Figure 2.19. Annual length frequencies of striped bass recreational harvest in the Roanoke River, 1994–2017. ....	110
Figure 2.20. Annual length frequencies of striped bass recreational discards in the Roanoke River, 2005–2017. ....	111
Figure 2.21. Map of NCDMF Juvenile Abundance Survey (Program 100) sampling sites. ....	112
Figure 2.22. Nominal and GLM-standardized indices of relative age-0 abundance derived from the Juvenile Abundance Survey (P100), 1991–2017. ....	113
Figure 2.23. Map of sampling grids and zones for the NCDMF Independent Gill-Net Survey (Program 135). ....	113
Figure 2.24. Nominal and GLM-standardized indices of relative abundance derived from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2016. ....	114
Figure 2.25. Nominal and GLM-standardized indices of relative abundance derived from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1992–2017. ....	114
Figure 2.26. Annual length frequencies of striped bass sampled from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. ....	115
Figure 2.27. Annual length frequencies of striped bass sampled from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. ....	116
Figure 2.28. Annual age frequencies of striped bass sampled from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. The age-15 bin represents a plus group. ....	117
Figure 2.29. Annual age frequencies of striped bass sampled from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. The age-15 bin represents a plus group. ....	118
Figure 2.30. Striped Bass spawning grounds on the Roanoke River, near the vicinity of Weldon, North Carolina. Black boxes represent relative locations of river strata. The gray star indicates location of rapids near the Weldon boating access area; flows less than 7,000 cfs restrict access to the strata above this location. ....	119
Figure 2.31. Nominal and GLM-standardized indices of relative abundance derived from the NCWRC Roanoke River Electrofishing Survey, 1994–2017. ....	120
Figure 2.32. Annual length frequencies of striped bass sampled from the NCWRC Roanoke River Electrofishing Survey, 1991–2017. ....	121
Figure 2.33. Annual age frequencies of striped bass sampled from the NCWRC Roanoke River Electrofishing Survey, 1991–2017. The age-15 bin represents a plus group. ....	122
Figure 3.1. Annual (A) ARcomm landings, (B) ASrec harvest, and (C) RRrec harvest values that were input into the SS model, 1991–2017. ....	123
Figure 3.2. Annual (A) ARcomm, (B) ASrec, and (C) RRrec dead discards that were input into the SS model, 1991–2017. ....	124

Figure 3.3.	GLM-standardized indices of abundance derived from the (A) P100juv, (B) P135fw, (C) P135spr, and (D) RRef surveys that were input into the SS model, 1991–2017.....	125
Figure 3.4.	Summary of the data sources and types used in the stock assessment model for striped bass.....	126
Figure 3.5.	Negative log-likelihood values produced from the 50 jitter trials in which initial parameter values were jittered by 10%. The solid black circle is the value from the base run.....	126
Figure 3.6.	Predicted (A) female SSB and (B) $F$ (numbers-weighted, ages 3–5) from the converged jitter trials (run 46 removed) in which initial parameter values were jittered by 10%, 1991–2017.....	127
Figure 3.7.	Observed and predicted (A) ARcomm landings, (B) ASrec harvest, and (C) RRrec harvest from the base run of the stock assessment model, 1991–2017.....	128
Figure 3.8.	Observed and predicted (A) ARcomm, (B) ASrec, and (C) RRrec dead discards from the base run of the stock assessment model, 1991–2017.....	129
Figure 3.9.	Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P100juv survey from the base run of the stock assessment model, 1991–2017.....	130
Figure 3.10.	Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P135fw survey from the base run of the stock assessment model, 1991–2017.....	131
Figure 3.11.	Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P135spr survey from the base run of the stock assessment model, 1992–2017.....	132
Figure 3.12.	Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the RRef survey from the base run of the stock assessment model, 1994–2017.....	133
Figure 3.13.	Observed and predicted length compositions for each data source from the base run of the stock assessment model aggregated across time. $N_{adj}$ represents the input effective sample size (number of trips sampled) and $N_{eff}$ represents the model estimate of effective sample size.....	134
Figure 3.14.	Observed and predicted length compositions for the ARcomm landings from the base run of the stock assessment model, 1991–2006. $N_{adj}$ represents the input effective sample size (number of trips sampled) and $N_{eff}$ represents the model estimate of effective sample size.....	135
Figure 3.15.	Observed and predicted length compositions for the ARcomm landings from the base run of the stock assessment model, 2007–2017. $N_{adj}$ represents the input effective sample size (number of trips sampled) and $N_{eff}$ represents the model estimate of effective sample size.....	136
Figure 3.16.	Observed and predicted length compositions for the ARcomm discards from the base run of the stock assessment model, 2004–2017. $N_{adj}$ represents the input effective sample size (number of trips sampled) and $N_{eff}$ represents the model estimate of effective sample size.....	137
Figure 3.17.	Observed and predicted length compositions for the ASrec harvest from the base run of the stock assessment model, 1996–2011. $N_{adj}$ represents the input	



	effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	138
Figure 3.18.	Observed and predicted length compositions for the ASrec harvest from the base run of the stock assessment model, 2012–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	139
Figure 3.19.	Observed and predicted length compositions for the ASrec discards from the base run of the stock assessment model, 1997–2012. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	140
Figure 3.20.	Observed and predicted length compositions for the ASrec discards from the base run of the stock assessment model, 2013–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	141
Figure 3.21.	Observed and predicted length compositions for the RRrec harvest from the base run of the stock assessment model, 1999–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	142
Figure 3.22.	Observed and predicted length compositions for the RRrec discards from the base run of the stock assessment model, 2005–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	143
Figure 3.23.	Observed and predicted length compositions for the P135fw survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	144
Figure 3.24.	Observed and predicted length compositions for the P135fw survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	145
Figure 3.25.	Observed and predicted length compositions for the P135spr survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	146
Figure 3.26.	Observed and predicted length compositions for the P135spr survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	147
Figure 3.27.	Observed and predicted length compositions for the RRef survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	148
Figure 3.28.	Observed and predicted length compositions for the RRef survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size. ....	149

Figure 3.29. Pearson residuals (red: female; blue: male) from the fit of the base model run to the ARcomm landings length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	150
Figure 3.30. Pearson residuals from the fit of the base model run to the ARcomm discards length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	150
Figure 3.31. Pearson residuals from the fit of the base model run to the ASrec harvest length composition data, 1996–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	151
Figure 3.32. Pearson residuals from the fit of the base model run to the ASrec discard length composition data, 1997–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	151
Figure 3.33. Pearson residuals (red: female; blue: male) from the fit of the base model run to the RRrec harvest length composition data, 1999–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	152
Figure 3.34. Pearson residuals from the fit of the base model run to the RRrec discard length composition data, 2005–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	152
Figure 3.35. Pearson residuals (red: female; blue: male) from the fit of the base model run to the P135fw survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	153
Figure 3.36. Pearson residuals (red: female; blue: male) from the fit of the base model run to the P135spr survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	153
Figure 3.37. Pearson residuals (red: female; blue: male) from the fit of the base model run to the RRef survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected). .....	154
Figure 3.38. Comparison of empirical and model-predicted age-length growth curves for (A) female and (B) male striped bass from the base run of the stock assessment model.....	155
Figure 3.39. Predicted length-based selectivity for the fleets from the base run of the stock assessment model.....	156
Figure 3.40. Predicted length-based selectivity for the P135fw and P135spr surveys from the base run of the stock assessment model.....	156
Figure 3.41. Predicted length-based selectivity for the RRef survey from the base run of the stock assessment model. ....	157

Figure 3.42. Predicted recruitment of age-0 fish from the base run of the stock assessment model, 1991–2017. Dotted lines represent $\pm 2$ standard deviations of the predicted values. ....	157
Figure 3.43. Predicted recruitment deviations from the base run of the stock assessment model, 1991–2017. Dotted lines represent $\pm 2$ standard deviations of the predicted values. ....	158
Figure 3.44. Predicted female spawning stock biomass from the base run of the stock assessment model, 1991–2017. Dotted lines represent $\pm 2$ standard deviations of the predicted values. ....	158
Figure 3.45. Predicted Beverton-Holt stock-recruitment relationship from the base run of the stock assessment model with labels on first (1991), last (2017), and years with (log) deviations $> 0.5$ . ....	159
Figure 3.46. Predicted spawner potential ratio (SPR) from the base run of the stock assessment model, 1991–2017. Dotted lines represent $\pm 2$ standard deviations of the predicted values. ....	159
Figure 3.47. Predicted fishing mortality (numbers-weighted, ages 3–5) from the base run of the stock assessment model, 1991–2017. Dotted lines represent $\pm 2$ standard deviations of the predicted values. ....	160
Figure 3.48. Sensitivity of model-predicted (A) female spawning stock biomass (SSB) and (B) fishing mortality rates (numbers-weighted, ages 3–5) to removal of different fisheries-independent survey indices from the base run of the stock assessment model, 1991–2017. ....	161
Figure 3.49. Sensitivity of model-predicted (A) female spawning stock biomass (SSB) and (B) fishing mortality rates (numbers-weighted, ages 3–5) to the assumption about natural mortality, 1991–2017. ....	162
Figure 3.50. Predicted recruitment from the sensitivity runs in which the assumption about natural mortality was changed, 1991–2017. ....	163
Figure 4.1. Estimated fishing mortality (numbers-weighted, ages 3–5) compared to fishing mortality target ( $F_{45\%}=0.18$ ) and threshold ( $F_{35\%}=0.13$ ). Error bars represent $\pm$ two standard errors. ....	164
Figure 4.2. Estimated female spawning stock biomass compared to spawning stock biomass target ( $SSB_{45\%}=159$ mt) and threshold ( $SSB_{35\%}=121$ mt). Error bars represent $\pm$ two standard errors. ....	164
Figure 5.1. Update of the nominal and GLM-standardized indices of relative age-0 abundance derived from the Juvenile Abundance Survey (P100), 1991–2019. ....	165

# **1 INTRODUCTION**

## **1.1 The Resource**

The common and scientific names for the species are striped bass, *Morone saxatilis* (Artedi et al. 1792). In North Carolina it is also known as striper, rockfish, or rock. Striped bass naturally occur in fresh, brackish, and marine waters along the western Atlantic coast from Canada to Florida, and through the U.S. coast of the Gulf of Mexico. Striped bass are anadromous, conducting annual spawning migrations in the spring of each year up to the fall line in freshwater tributaries. In addition, after spawning portions of the stocks from the Albemarle Sound-Roanoke River, Chesapeake Bay, Delaware Bay, and the Hudson River migrate along the Atlantic coast north in the summer and south in the winter. The stocks from the Chesapeake Bay constitute the majority of this migrating population. Due to these facts, striped bass have been the focus of fisheries from North Carolina to New England for several centuries and have played an integral role in the development of numerous coastal communities (ASMFC 1998). Striped bass regulations in the United States date to colonial times; in 1639 the Massachusetts Bay colony passed a law that prohibited striped bass from being used as fertilizer to promote fishery commerce with Europe (Hutchinson, T. [1764] 1936; McFarland 1911).

## **1.2 Life History**

### **1.2.1 Stock Definitions**

There are two geographic management units and four striped bass stocks inhabiting the estuarine and inland waters of North Carolina. The northern management unit is comprised of two harvest management areas: the Albemarle Sound Management Area (ASMA) and the Roanoke River Management Area (RRMA; Figure 1.1). The striped bass stock in the two harvest management areas is referred to as the Albemarle-Roanoke (A-R) stock, and its spawning grounds are located in the Roanoke River in the vicinity of Weldon, NC. The ASMA includes the Albemarle Sound and all its tributaries, (except for the Roanoke, Middle, East-most, and Cashie rivers), Currituck, Roanoke and Croatan sounds and all their tributaries, including Oregon Inlet, north of a line from Roanoke Marshes Point across to the north point of Eagle Nest Bay in Dare county. The RRMA includes the Roanoke River and its tributaries, including Middle, East-most, and Cashie rivers, up to the Roanoke Rapids Lake Dam. Management of recreational and commercial striped bass regulations within the ASMA is the responsibility of the NCDMF. Within the RRMA, commercial regulations are the responsibility of the NCDMF while recreational regulations are the responsibility of the North Carolina Wildlife Resources Commission (NCWRC). The A-R stock is also included in the management unit of Amendment 6 to the Atlantic States Marine Fisheries Commission (ASMFC) Interstate Fishery Management Plan (FMP) for Atlantic Striped Bass (ASMFC 2003).

### **1.2.2 Movements & Migration**

Numerous tagging studies have been conducted on striped bass in North Carolina and along the Atlantic Coast since the 1930s. Several older studies suggest the A-R stock is at least partially migratory, with primarily older adults participating in offshore migrations. Tag-recapture studies (Merriman 1941; Vladykov and Wallace 1952; Davis and Sykes 1960; Chapoton and Sykes 1961; Nichols and Cheek 1966; Holland and Yelverton 1973; Street et al. 1975; Hassler et al. 1981; Boreman and Lewis 1987; Benton, unpublished) indicated that a small amount of offshore

migration occurs; however, these studies occurred when the stock was experiencing very high exploitation rates and the age structure was truncated. Most of the fish tagged during these early studies were young and male. Recent research on the A-R stock demonstrates that as A-R striped bass get older they migrate out of the ASMA into North Carolina's near shore ocean waters, and then as they continue to age they participate in summertime coastal migrations to northern areas including Chesapeake Bay, Delaware Bay, Hudson Bay, and coastal areas of New Jersey, New York, Rhode Island, and Massachusetts (Callihan et al. 2014). The probability of a six-year-old striped bass (average size 584 mm or 23 inches total length, TL) migrating out of the ASMA is 7.5%. This probability increases with age, and by age 11 (average size 940 mm or 37 inches TL) the probability of migrating outside North Carolina's waters is 72.5%. (Callihan et al. 2014). Callihan et al. (2014) also found that when the total A-R stock abundance is higher there is a greater likelihood that smaller striped bass utilize habitat in the Pungo, Tar-Pamlico, and Neuse rivers and northwestern Pamlico Sound.

### **1.2.3 Age & Size**

Striped bass have been aged using scales for more than 70 years (Merriman 1941). Scales of striped bass collected in North Carolina show annulus formation taking place between late April through May in the Albemarle Sound and Roanoke River (Trent and Hassler 1968; Humphries and Kornegay 1985). Annuli form on scales of striped bass caught in Virginia between April and June during the spawning season (Grant 1974).

Age data have been a fundamental part of assessing A-R striped bass since the first A-R assessment (Gibson 1995). The oldest observed striped bass in the A-R stock to date (in 2017) was 23 years old from the 1994 year class. The fish was originally collected and tagged on the spawning grounds during the 2007 season by the NCWRC, aged to 13 years old and was then recaptured by an angler on June 10, 2017 near Sandy Hook, New Jersey. The fish was 40 inches long and weighed 35 pounds when originally tagged. Historically, Smith (1907) reported several striped bass captured in pound nets in Edenton in 1891 that weighed 125 pounds each. Worth (1904) reported the largest female striped bass taken at Weldon that year for strip spawning weighed 70 pounds. The oldest striped bass observed in the data used for this assessment was 17 years old.

### **1.2.4 Growth**

As a relatively long-lived species, striped bass can attain a moderately large size. Females grow to a considerably larger size than males; striped bass over 30 pounds are almost exclusively female (Bigelow and Schroeder 1953; NCDMF and NCWRC, unpublished data).

Growth rates for the A-R stock are rapid during the first three years of life and then decrease to a slower rate as the fish reach sexual maturity (Olsen and Rulifson 1991). Growth occurs between April and October. Striped bass stop feeding for a brief period just before and during spawning but feeding continues during the upriver spawning migration and begins again soon after spawning (Trent and Hassler 1966). From November through March growth is negligible.

Available annual age data (scales) were fit with the von Bertalanffy age-length model to estimate growth parameters for both female and male striped bass. This model was weighted by the number of data points and applied to fractional ages. Unsexed age-0 fish were included in the fits for both the males and females. Estimated parameters of the age-length model are shown in Table 1.1. Fits to the available data performed well for both females (Figure 1.2) and males (Figure 1.3).

Parameters of the length-weight relationship were also estimated in this study. The relation of total length in centimeters to weight in kilograms was modeled for males and females separately. Parameter estimates of the length-weight model are shown in Table 1.2. Predicted weight at length performed well based on both the female (Figure 1.4) and male (Figure 1.5) striped bass data.

### **1.2.5 Reproduction**

Striped bass spawn in freshwater or nearly freshwater portions of North Carolina's coastal rivers from late March to June depending on water temperatures (Hill et al. 1989). Peak spawning activity occurs when water temperatures reach 16.7°–19.4°C (62.0°–67.0°F) on the Roanoke River (Rulifson 1990, 1991). Spawning behavior is characterized by brief peaks of surface activity when a mature female is surrounded by up to 50 males as eggs are broadcast into the surrounding water, and males release sperm, termed “rock fights” by locals (Worth 1904; Setzler et al. 1980). Spawning by a given female is probably completed within a few hours (Lewis and Bonner 1966).

#### **1.2.5.1 Eggs**

Mature eggs are 1.0–1.5 mm (0.039 to 0.059 inch) in diameter when spawned and remain viable for about one hour before fertilization (Stevens 1966). Fertilized eggs are spherical, non-adhesive, semi-buoyant, and nearly transparent. The incubation period at peak spawning temperatures ranges from 42 to 55 hours. At 20.0°C (68.0°F), fertilized eggs need to drift downstream with currents to hatch into larvae. If the egg sinks to the bottom, its chances of hatching are reduced because the sediments reduce oxygen exchange between the egg and the surrounding water. Hassler et al. (1981) found that eggs hatch in 38 hours. After hatching, larvae are carried by the current to the downstream nursery areas located in the western Albemarle Sound (see section 1.3.3; Hassler et al. 1981).

#### **1.2.5.2 Larvae**

Larval development is dependent upon water temperature and is usually regarded as having three stages: (1) yolk-sac larvae are 5–8 mm (0.20 to 0.31 inch) in total length (TL) and depend on yolk material as an energy source for 7 to 14 days; (2) fin-fold larvae (8–12 mm; 0.31–0.47 inch TL) having fully developed mouth parts and persist about 10 to 13 days; and (3) post fin-fold larvae attain lengths up to 30 mm (1.18 inches) TL in 20 to 30 days (Hill et al. 1989). Researchers of North Carolina stocks of striped bass (primarily the A-R stock) divide larval development into yolk-sac and post yolk-sac larvae (Hill et al. 1989; Rulifson 1990). Growth occurs generally within the same rates described above depending upon temperature. At temperatures  $\geq 20^{\circ}\text{C}$  (68°F) larvae develop into juveniles in approximately 42 days (Hassler et al. 1981).

#### **1.2.5.3 Juveniles**

Most striped bass enter the juvenile stage at about 30 mm (1.18 inches) TL; the fins are then fully formed, and the external morphology of the young is like the adults. Juveniles are often found in schools and associate with clean sandy bottoms (Hill et al. 1989). Juveniles spend the first year of life in western Albemarle Sound and lower Chowan River nursery areas (Hassler et al. 1981). There is evidence of density-dependent habitat utilization; when large year classes are produced juveniles are collected in early June as far away from the western Albemarle Sound as the lower Alligator River (63 water miles) and Stumpy Point, Pamlico Sound (75 water miles; NCDMF, unpublished data).

#### 1.2.5.4 Maturation & Fecundity

Early research conducted on the A-R stock indicated that females began reaching sexual maturity in approximately three years, at sizes of about 45.7 cm (18 inches) TL (Trent and Hassler 1968; Harris and Burns 1983; Harris et al. 1984). In the most recent maturation study conducted on a recovered stock with expanded age structure, Boyd (2011) found that 29% of A-R females reached sexual maturity by age 3, while 97% were mature by age 4, and 100% were mature at age 5 (Table 1.3). In general, there is a strong positive correlation between the length, weight, and age of a female striped bass and the number of eggs produced. Boyd (2011) estimated fecundity ranging from 176,873 eggs for an age-3 fish to 3,163,130 eggs for an age-16 fish.

#### 1.2.6 Mortality

##### 1.2.6.1 Natural Mortality

Striped bass are a long-lived species with a maximum age of at least 31 years (Atlantic coastal stock) based on otoliths (Secor 2000), suggesting overall natural mortality is relatively low. Previous assessments have assumed a constant natural mortality ( $M$ ) of 0.15 across all ages, consistent with Hoenig's (1983) regression on maximum age (ASMFC 2009; NCDMF 2010).

Harris and Hightower (2017) estimated annual total instantaneous natural mortality for striped bass using both an integrated model and a multi-state only model based on VEMCO acoustic, Passive Integrated Transponder, and traditional external anchor tagging data. The integrated model produced a study-wide natural mortality rate of 0.70 while the multi-state only model produced an estimate of 0.74 (average of 0.72 over the two methods). The estimates apply to striped bass ranging in length from 45.8 cm to 89.9 cm (18 inches to 35 inches, approximately 3 to 9 years old).

There are a number of methods available to estimate natural mortality based on life history characteristics. These include approaches based on parameters of the von Bertalanffy age-length relationship (Alverson and Carney 1975; Ralston 1987; Jensen 1996; Cubillos 2003) as well as approaches based on maximum age (Alverson and Carney 1975; Hoenig 1983; Hewitt and Hoenig 2005; Then et al. 2015). Several of these methods were applied to A-R striped bass to produce estimates of age-constant natural mortality for females and males. Values for the life history parameters required by some of these approaches were those estimated in this stock assessment (see section 1.2.4). For approaches that depend on maximum age, a maximum age of 17 was assumed for females and a maximum age of 15 was assumed for males. These maximum ages are based on the maximum ages observed in the available data within the ASMA and RRMA over the assessment time series (1991–2017). Life history-based empirical estimates of age-constant natural mortality ranged from 0.099 to 0.37 for females and from 0.090 to 0.44 for males (Table 1.4).

Natural mortality of long-lived fish species is commonly considered to decline with age, as larger fish escape predation. Several approaches are available to derive estimates of age-varying natural mortality (e.g., Lorenzen 1996, 2005). Here, the Lorenzen (1996) approach was used to produce estimates of  $M$  at age. As expected, estimates of  $M$  decrease with increasing age (Table 1.5; Figure 1.6).

### **1.2.6.2 Discard Mortality**

Discards from the commercial gill-net fishery are broken into two categories, live and dead discards as recorded by the observer. Live discards are multiplied by a discard mortality rate, which for gill-net fisheries is estimated at 43% (ASMFC 2007).

Nelson (1998) estimated short-term mortality for striped bass caught and released by recreational anglers in the Roanoke River, North Carolina as 6.4%. Nelson found that water temperature and hooking location were important factors affecting catch-and-release mortality, consistent with previous studies (Harrell 1988; Diodati 1991).

### **1.2.7 Food & Feeding Habits**

Several food habit studies have been conducted for juvenile and adult striped bass since 1955 in the Roanoke River and Albemarle Sound. Studies of juvenile striped bass diets in Albemarle Sound found zooplankton and mysid shrimp as primary prey items in the summer, with small fish (most likely bay anchovies) entering the diet later in the season (Rulifson and Bass 1991; Cooper et al. 1998). Adults feed extensively on blueback herring and alewives in the river during the spawning migration (Trent and Hassler 1968). Manooch (1973) conducted a seasonal food habit study in Albemarle Sound and found primarily fish in the Clupeidae (Atlantic menhaden, blueback herring, alewife, and gizzard shad) and Engraulidae (anchovies) families dominated the diet in the summer and fall. Atlantic menhaden (54%) was the most frequently eaten species and comprised a relatively large percentage of the volume (50%). In the winter and spring months, invertebrates occurred more frequently in the diet (primarily amphipods during the winter and blue crabs in the spring). Similarly, Rudershausen et al. (2005) found a diverse array of fish in the diets of age-1 striped bass whereas the diets of age-2 and age-3+ striped bass were primarily comprised of menhaden in 2002 and 2003 in the Albemarle Sound. Tuomikoski et al. (2008) investigated age-1 striped bass diets in Albemarle Sound where American shad comprised most of their diet in 2002, but yellow perch dominated the diet in 2003. The 2003 year class for yellow perch was one of the highest on record in NCDMF sampling programs, so the high occurrence of yellow perch in striped bass stomachs may not be typical (NCDMF 2010). However, it also supports other research that striped bass exhibit an opportunistic feeding behavior (Rulifson et al. 1982).

From the fall of 1995 through the spring of 2001, stomach contents from 1,796 striped bass collected from the NCDMF Striped Bass Independent Gill-Net Survey were analyzed. Unidentifiable fish parts were the dominant stomach content from western Albemarle Sound samples (35.9%), followed by river herring (33.2%) and Atlantic menhaden (16.5%). The dominance of river herring during the spawning migration supports results reported by Trent and Hassler (1968) and Manooch (1973). Blue crab accounted for 0.2% of the total stomach contents from the western sound. In eastern Albemarle Sound samples, unidentifiable fish parts accounted for 34.0%, followed by Atlantic menhaden (31.5%), Atlantic croaker (12.1%), anchovy spp. (11.1%) and spot (6.5%). Blue crab comprised 2.1% of the stomach contents from the eastern sound.

From the fall of 2001 through the spring 2010, the NCDMF analyzed 4,448 striped bass stomachs having food contents. In western Albemarle Sound samples unidentifiable fish parts accounted for 61.2% of stomach contents, followed by Atlantic menhaden (23.1%), anchovy spp. (4.0%), invertebrates (3.0%), Atlantic croaker (2.5%), and river herring (2.0%). Blue crab accounted for less than 1.0% of stomach contents in western sound samples. It is interesting to note the decline in the prevalence of river herring in striped bass diets in the western sound since 2001. In eastern



Albemarle Sound samples, unidentifiable fish parts accounted for 41.2% of the stomach contents, followed by Atlantic menhaden (40.8%), anchovy spp. (6.4%), spot (6.4%), and Atlantic croaker (2.9%). Blue crab accounted for less than 1.0% of stomach contents in the eastern sound samples as well.

From 2011 through 2017, the NCDMF analyzed 1,918 striped bass stomachs having contents. In western Albemarle Sound samples, unidentifiable fish parts accounted for 35.9% of stomach contents, followed by Atlantic menhaden (12.6%), Atlantic croaker (10.0%), and Clupeidae species (1.8%). Blue crab accounted for less than 1.0% of stomach contents in western sound samples. In eastern Albemarle Sound samples, unidentifiable fish parts accounted for 19.3% of the stomach contents, followed by Atlantic menhaden (2.4%) and invertebrates (1.7%). Blue crab accounted for less than 1.0% of stomach contents in the eastern sound samples.

### **1.3 Habitat**

#### **1.3.1 Overview**

Habitat loss has contributed to the decline in anadromous fish stocks throughout the world (Limburg and Waldman 2009). Striped bass use a variety of habitats as described in the life history section with variations in habitat preference due to location, season, and ontogenetic stage. Although primarily estuarine, striped bass use habitats throughout estuaries and the coastal ocean. Striped bass are found in most habitats identified by the North Carolina Coastal Habitat Protection Plan (CHPP) including: water column, wetlands, submerged aquatic vegetation (SAV), soft bottom, hard bottom, and shell bottom (NCDEQ 2016). Each habitat is part of a larger habitat mosaic, which plays a vital role in the overall productivity and health of the coastal ecosystem. Although striped bass are found in all of these habitats, usage varies by habitat. Additionally, these habitats provide the appropriate physicochemical and biological conditions necessary to maintain and enhance the striped bass population. Therefore, the protection of each habitat type is critical to the sustainability of the striped bass stock.

#### **1.3.2 Spawning Habitat**

The main spawning habitat for A-R striped bass is in the Roanoke River in the vicinity of Weldon, NC, around river mile (RM) 130. This is the location of the first set of rapids at the fall line transition between the Coastal Plain and the Piedmont. Historic accounts indicate major spawning activity centered at Weldon (Worth 1904), but striped bass were known to migrate up the mainstem Roanoke River to Clarksville, VA (RM 200; Moseley et al. 1877) and possibly as far as Leesville, VA (RM 290; NMFS and USFWS 2016). Striped bass spawning migrations have been impeded since construction of the initial dam on the mainstem of the Roanoke River at Roanoke Rapids, NC (RM 137) around 1900 (NMFS and USFWS 2016). The dam was approximately 12-feet high (Hightower et al. 1996) and impeded striped bass migrations especially during low flow years. Completion of the John H. Kerr Dam, 42 river miles upstream of Roanoke Rapids Dam, by the U.S. Army Corps of Engineers in 1953 completely blocked access to upriver habitats, and construction of the current Roanoke Rapids Dam by Virginia Electric and Power Company in 1955 and Gaston Dam in 1964 eliminated striped bass usage of the 42 river miles below Kerr Dam (NMFS and USFWS 2016). Spawning activity now ranges from RM 78 to RM 137 with most of the activity occurring between RM 120 and RM 137, still centered around Weldon.

### **1.3.3 Nursery & Juvenile Habitat**

Juveniles are found in schools; the location of the schools varies considerably with the age of the fish and apparently prefer clean sandy bottoms but have been found over gravel beaches, rock bottoms, and soft mud (Hill et al. 1989). The Roanoke River delta area does not seem to be an important nursery area for YOY striped bass. They appear to spend the first year of life (age-0) growing in and around the western Albemarle Sound and lower Chowan River (Hassler et al. 1981).

As they enter their second and third year, striped bass are found throughout Albemarle Sound and its tributaries. The presence of age-1 and -2 striped bass in the Albemarle Sound Independent Gill-Net Survey confirms this, as well as reports of discarded undersized fish from the striped bass recreational creel survey conducted throughout the Albemarle Sound and its tributaries (NCDMF, unpublished data).

### **1.3.4 Adult Habitat**

Analysis of tagging data indicate younger, smaller adult A-R striped bass (from 35.0–60.0 cm TL) remain in inshore estuarine habitats, while older, larger adults (>60.0 cm TL) are much more likely to emigrate to ocean habitats after spawning; (Callihan et al. 2014). Further, smaller adults show evidence of density-dependent movements and habitat utilization, as the likelihood of recapture outside the ASMA in adjacent systems (i.e., northwestern Pamlico Sound, Tar-Pamlico, Pungo, and Neuse rivers, lower Chesapeake Bay, and the Blackwater and Nottoway rivers in Virginia) increases during periods of higher stock abundance (Callihan et al. 2014).

### **1.3.5 Habitat Issues & Concerns**

Numerous documents have been devoted entirely to habitat issues and concerns, including the North Carolina Coastal Habitat Protection Plan (Street et al. 2005; NCDEQ 2016) and ASMFC's "Atlantic Coast Diadromous Fish Habitat: A review of Utilization, Threats, Recommendations for Conservation, and Research Needs" (Greene et al. 2009). Many contaminants are known to adversely affect striped bass at numerous life stages and can be detrimental to eggs and larvae (Buckler et al. 1987; Hall et al. 1993; Ostrach et al. 2008). Adequate river flows during the spawning season are also needed to keep eggs suspended for proper development (N.C. Striped Bass Study Management Board 1991).

Hassler et al. (1981) indicated that adequate river flow during the pre-spawn and post-spawn periods was the most important factor contributing to survival of fish larvae and the subsequent production of strong or poor year classes.

## **1.4 Description of Fisheries**

Since 2015, the current total allowable landings (TAL) has been set at 124.7 metric tons (275,000 lb) and is split evenly between the commercial and recreational fisheries in the ASMA and RRMA (Table 1.6). In the ASMA, the commercial fishery has a TAL of 62.37 metric tons (137,500 lb) while the ASMA and RRMA recreational fisheries each have a TAL of 31.18 metric tons (68,750 lb). The TAL has changed throughout the previous two decades in response to changes in stock abundance and has ranged from a low of 71.12 metric tons (156,800 lb) in the early 1990s to 249.5 metric tons (550,000 lb) from 2003 to 2014.

### **1.4.1 Commercial Fishery**

Striped bass are landed commercially in the ASMA primarily with anchored gill nets and to a lesser degree by pound nets. Insignificant landings occur in fyke nets and crab pots. Since 1991, landings in the commercial fishery have ranged from a low of 31.03 metric tons (68,409 lb) in 2013 to a high of 124.2 metric tons (273,814 lb) in 2004 (Table 1.7). Total catch has shown an overall decline since 2004.

#### **1.4.1.1 Historical**

The Albemarle Sound area commercial striped bass fishery has been documented in numerous reports for over 100 years. Worth (1884) suggests an industry origin of 1872. During the early 1880s, a large fishery developed on Roanoke Island catching striped bass in the spring and fall (Taylor and White 1992). Gears included haul seines, drag nets, purse seines, fish traps, and gill nets. In 1869, pound nets were first used in the Albemarle Sound and became a more prominent aspect of the fishery in the early 1900s (Taylor and White 1992). The commercial fishery for striped bass has principally occurred from November through April in the Albemarle Sound, whereas, Roanoke River commercial effort was concentrated during the spring spawning run. During the summer months, landings from all areas were much lower (Hassler et al. 1981). Anchored and drift gill nets were the most productive gear types in the spring spawning run portion of the Roanoke River fishery. In 1981, anchored gill nets were prohibited in the Roanoke River, and the mesh size of drift gill nets was restricted, resulting in sharply curtailed landings during the spawning run (Hassler and Taylor 1984). Bow and dip netting was a productive method of harvesting spawning fish in the Roanoke River until it was prohibited in 1981. Prior to this rule, fishermen using bow nets in the upper Roanoke River could retain 25 striped bass per day when taken incidentally during shad and river herring fishing. A local law allowing the commercial sale of striped bass in Halifax and Northampton counties was enacted by the North Carolina General Assembly and created a prominent commercial fishery for striped bass in its principal spawning area (Hassler et al. 1981). This law was repealed in 1981 and commercial fishing for striped bass was eliminated in the inland portions of the Roanoke River. Limited commercial fishing seasons were implemented in Albemarle Sound in 1984 (October–May; Henry et al. 1992). State regulations enacted in 1985 prohibited the sale of hook-and-line-caught striped bass.

#### **1.4.1.2 Current**

The ASMA commercial striped bass fishery from 1990 through 1997 operated on a 44.45-metric ton (98,000-lb) TAL (Table 1.6). The TAL was split to have a spring and fall season. The commercial fishery operated with net yardage restrictions, mesh size restrictions, size limit restrictions, and daily landing limits. The A-R stock was declared recovered in 1997 by the ASMFC. In 1998, the commercial TAL was increased to 56.88 metric tons (125,400 lb) and additional increases in poundage occurred in 1999 and 2000. From 2000 through 2002, the commercial TAL remained at 102.1 metric tons (225,000 lb). In 2015, the TAL was adjusted to a total of 124.7 metric tons (275,000 lb) for all sectors, based on projections from the 2014 benchmark stock assessment (NCDMF 2014). Since the initial TAL was set in 1990, seasons, yardage, mesh size restrictions, and daily landing limits have been used to control harvest and maintain the fishery as a bycatch fishery.

### **1.4.2 Recreational Fishery**

Striped bass are landed recreationally in the ASMA and RRMA by hook and line, primarily by trolling or casting artificial lures and using live or cut bait. In recent years, the catch-and-release

fly fishery in the RRMA has seen an increase in angler effort. Combined recreational harvest from both management areas has ranged from 5.9 metric tons (13,095 lb) in 1985 to 106.9 metric tons (235,747 lb) in 2000 (Table 1.7). Since 1997, harvest steadily increased from 25.2 metric tons (55,653 lb) to 106.9 metric tons (235,747 lb) in 2000. Since 2000, harvest has shown an overall decline, except for a slight increase in 2011–2012 for the ASMA, 2012 for the RRMA, 2015 for the ASMA, and 2015–2016 for the RRMA. The harvest estimate for 2017 in the ASMA stands as the third lowest on record since 1982.

## **1.5 Fisheries Management**

### **1.5.1 Management Authority**

Fisheries management includes all activities associated with maintenance, improvement, and utilization of the fisheries resources of the coastal area, including research, development, regulation, enhancement, and enforcement.

North Carolina’s existing fisheries management system for striped bass is adaptive, with rulemaking authority vested in the North Carolina Marine Fisheries Commission (NCMFC) and the North Carolina Wildlife Resources Commission (NCWRC) within their respective jurisdictions. The NCMFC also has the authority to delegate to the fisheries director the ability to issue public notices, called proclamations, suspending or implementing particular commission rules that may be affected by variable conditions.

Fisheries management includes all activities associated with maintenance, improvement, and utilization of the fisheries resources of the coastal area, including research, development, regulation, enhancement, and enforcement. North Carolina’s existing fisheries management system is powerful and flexible, with rulemaking (and proclamation) authority vested in the NCMFC and the NCWRC within their respective jurisdictions.

The North Carolina Department of Environmental Quality (NCDEQ) is the parent agency of the NCMFC and the NCDMF. The NCMFC is responsible for managing, protecting, preserving and enhancing the marine and estuarine resources under its jurisdiction, which include all state coastal fishing waters extending to three miles offshore. In support of these responsibilities, the NCDMF conducts management, enforcement, research, monitoring statistics, and licensing programs to provide information on which to base these decisions. The NCDMF presents information to the NCMFC and NCDEQ in the form of fisheries management and coastal habitat protections plans and proposed rules. The NCDMF also administers and enforces the NCMFC’s adopted rules.

The NCWRC is a state government agency authorized by the General Assembly to conserve and sustain the state’s fish and wildlife resources through research, scientific management, wise use and public input. The Commission is the regulatory agency responsible for the creation and enforcement of hunting, trapping and boating laws statewide and fishing laws within its jurisdictional boundaries including all designated inland fishing waters. The NCWRC and NCDMF share authority for regulating recreational fishing activity in joint fishing waters.

### **1.5.2 Management Unit Definition**

There are two geographic management units defined in the estuarine striped bass FMP and include the fisheries throughout the coastal systems of North Carolina (NCDMF 2004). The management unit for this assessment is the ASMA and RRMA and is defined as:

Albemarle Sound Management Area (ASMA) includes the Albemarle Sound and all its joint and inland water tributaries, (except for the Roanoke, Middle, Eastmost and Cashie rivers), Currituck, Roanoke and Croatan sounds and all their joint and inland water tributaries, including Oregon Inlet, north of a line from Roanoke Marshes Point across to the north point of Eagle Nest Bay in Dare county. The Roanoke River Management Area (RRMA) includes the Roanoke River and its joint and inland water tributaries, including Middle, Eastmost and Cashie rivers, up to the Roanoke Rapids Dam. The striped bass stock in these two harvest management areas is referred to as the Albemarle Sound-Roanoke River (A-R) stock, and its spawning grounds are located in the Roanoke River in the vicinity of Weldon, NC. Management of recreational and commercial striped bass regulations within the ASMA is the responsibility of the North Carolina Marine Fisheries Commission (NCMFC). Within the RRMA commercial regulations are the responsibility of the NCMFC while recreational regulations are the responsibility of the North Carolina Wildlife Resources Commission (NCWRC). The A-R stock is also included in the management unit of the Atlantic States Marine Fisheries Commission (ASMFC) Amendment #6 to the Interstate Fishery Management plan (FMP) for Atlantic Striped Bass and includes Albemarle Sound and all its joint and Inland Water tributaries, (except for the Roanoke, Middle, Eastmost and Cashie rivers), Currituck, Roanoke, and Croatan sounds and all their Joint and Inland Water tributaries, including Oregon Inlet, north of a line from Roanoke Marshes Point 35 48'.5015' N – 75 44'.1228' W across to the north point of Eagle Nest Bay 35 44'.1710' N - 75 31'.0520' W (Figure 1.1).

### **1.5.3 Regulatory History**

The ASMA commercial striped bass fishery from 1991 through 1997 operated on a 44.45-metric ton TAL (Table 1.6). The TAL was split to have a spring and fall season. The commercial fishery operated with net yardage restrictions, mesh size restrictions, size limit restrictions, and daily landing limits. The A-R stock was declared recovered in 1997 by the ASMFC. In 1998, the commercial TAL was increased to 56.88 metric tons and additional increases in the TAL occurred in 1999 and 2000. From 2000 through 2002, the commercial TAL remained at 102.1 metric tons. The ASMFC Striped Bass Management Board approved another TAL increase in 2003. From 2003 to 2014, the TAL remained at 249.5 metric tons. Based on a stock assessment benchmark, the TAL was reduced to 124.7 metric tons in 2015. Since the initial TAL was set in 1990, seasons, yardage, mesh size restrictions, and daily landing limits have been used to control harvest and maintain the fishery as a bycatch fishery.

Striped bass have been managed as a bycatch of the multi-species commercial fishery in the ASMA since 1991. Since 1991, when the striped bass season was open, commercial fishermen were allowed to land from seven to 15 fish per day, not to exceed 50% by weight of the total catch and fish had to meet the 18-inch TL minimum size limit. Gill nets continue to account for the highest percentage of the commercial harvest, followed by pound nets.

### **1.5.4 Current Regulations**

Striped bass from the A-R stock are harvested commercially within the ASMA and recreationally in both the RRMA and the ASMA. Commercial harvest is currently limited to the ASMA although there was a small commercial fishery operating in the Roanoke River during the early 1980s. The commercial fishery is regulated as a bycatch fishery with a TAL, size limits, daily possession limits, seasonal (closed May 1 through September 30) and gear restrictions, net attendance

requirements, and permitting and reporting requirements all imposed to prevent TAL overages and limit discard losses. Finfish dealers who purchase striped bass are required to obtain a striped bass dealer permit from NCDMF. The dealers are required to report their landings daily to NCDMF for the quota to be monitored. Dealers are also required to affix striped bass sale tags, provided by NCDMF, to the fish when purchased from the fishermen.

The recreational fishery within the RRMA is regulated through a creel limit, minimum size limit including a protective slot, and a fixed length spring season, while the ASMA recreational fishery is regulated through a creel limit, minimum size, and the variable spring and fall seasons that close once harvest targets are reached or set season closure dates are reached (closed May 1 through September 30). The A-R striped bass stock is managed by the NCDMF, the NCWRC, and the South Atlantic Fisheries Coordination Office (SAFCO) of the U.S. Fish and Wildlife Service (USFWS) under guidelines established in the ASMFC Interstate FMP for Atlantic Striped Bass and the North Carolina Estuarine Striped Bass FMP.

### **1.5.5 Management Performance**

Management strategies for the A-R striped bass stock have met with variable success over the last several decades. Unrestricted harvest and poor habitat conditions led to a stock collapse in the 1980s; however, severe harvest restrictions and Roanoke River streamflow improvements led to population recovery spurred by increases in recruitment, spawning stock biomass growth, and age structure expansion in the late 1990s and 2000s. Consequently, commercial and recreational harvest restrictions were eased, and the TAL was increased throughout the 2000s. From 1990 through 2002, harvest reached the TAL easily, with the season often having to close after only weeks or months to prevent harvest from exceeding the TAL. Starting in 2003, with the increase in TAL to 249 metric tons, harvest started to consistently decline through 2008, even with extended commercial and recreational seasons in the ASMA. From 2009 through 2014, harvest was still well below the TAL (Figure 1.7). The reason for the decline in harvest even with extended seasons is likely due to declining stock abundance due to several poor year classes produced from 2001 to present. Even with a reduction in the TAL in 2015 to 125 metric tons, harvest has not reached the TAL, although a reduced American shad season starting in 2014 could have contributed to the commercial quota not being reached as the majority of commercial harvest historically came during the American shad commercial season in the ASMA. Recent survey data and stock assessments have supported managers' concerns about declining landings, poor recruitment, reductions in population abundance, and a truncation of age structure (NCDMF 2014, 2018).

## **1.6 Assessment History**

### **1.6.1 Review of Previous Methods & Results**

The A-R stock has an extensive assessment history. Dorazio (1995) and Gibson (1995) prepared the first comprehensive assessment of the A-R striped bass stock based on a Virtual Population Analysis (VPA using CAGEAN, Deriso et al. 1985) and a Brownie tag-return model analysis (Brownie et al. 1985). Schaaf (1997) later provided CAGEAN-based VPA results through 1996 based on the methodology established in Gibson (1995). Smith (1996) used the MARK software program to estimate survival of striped bass in Albemarle Sound through analysis of release and recovery data. Carmichael (1998) updated the CAGEAN assessment through 1997 and later developed an ADAPT VPA assessment of the A-R stock using age-specific indices from the Albemarle Sound Independent Gill-Net surveys, the Roanoke River Electrofishing Survey, and

juvenile and yearling abundance indices from Albemarle Sound (Carmichael 1999). The 1999 assessment also included an analysis of tag-return data based on the MARK program. The ADAPT catch-at-age and MARK tag-return assessment framework was updated in 2000 (Carmichael 2000). Analysis of tag-return data for estimation of mortality was discontinued after 2000 as the results were deemed similar to those from the VPA and was duplicative work; subsequent assessments focused on the catch-at-age data. The VPA stock assessment was conducted annually until 2006 to determine stock status and to evaluate potential changes to the TAL (Carmichael 2001, 2002, 2003; Grist 2004, 2005; Takade 2006). The assessment shifted to an ASAP2 model for the 2010 assessment and a yield-per-recruit (YPR) model was used to calculate the benchmarks externally (Takade 2010). The 2014 assessment was performed similarly using an ASAP3 model and benchmarks were calculated with a YPR model. Projections were made using the Age Structured Projection Model (AGEPRO). The most recent stock assessments indicated that the stock was not overfished and overfishing was not occurring (Mroch and Godwin 2014; Flowers et al. 2016).

### **1.6.2 Progress on Research Recommendations**

- Incorporate high reward tagging into the current tagging program to provide estimates of tag return rates for each sector; this will allow for more precise estimates of natural mortality and fishing mortality from tag-based analyses.

There is an ongoing multi-species tagging study that was initiated in 2014 and funded through the NCDMF Coastal Recreational Fishing Fund. The study employs both high reward and double tags to estimate tag loss and angler reporting rates.

- Improve estimates of discard losses from the Albemarle Sound Management Area (ASMA) commercial gill-net fisheries.

NCDMF's Programs 466 and 467 monitor commercial gill-net fisheries and record bycatch (see also section 2.1.2). These programs are continually expanding and should lead to improved estimates of commercial discards over time.

- Re-evaluate hook-and-release mortality rates from the ASMA and RRMA recreational fisheries incorporating different hook types and angling methods at various water temperatures (e.g., live bait, artificial bait, and fly fishing).

No progress.

- Improve estimates of hook-and-release discard losses in the recreational fishery during the closed harvest season

There is a plan in place starting in May 2021 to provide additional funding to the existing striped bass creel survey in the ASMA that will extend intercepts during the closed harvest season (May–September).

## **2 DATA**

### **2.1 Fisheries-Dependent**

#### **2.1.1 Commercial Landings**

##### **2.1.1.1 Survey Design & Methods**

Prior to 1978, North Carolina's commercial landings data were collected by the National Marine Fisheries Service (NMFS). Between 1978 and 1993, landings information was gathered through the NMFS/North Carolina Cooperative Statistics program. Reporting was voluntary during this period, with North Carolina and NMFS port agents sampling the state's major dealers (Lupton and Phalen 1996). Beginning in 1994, the NCDMF instituted a mandatory dealer-based trip-ticket system to track commercial landings.

On January 1, 1994, the NCDMF initiated a Trip Ticket Program (NCTTP) to obtain more complete and accurate trip-level commercial landings statistics (Lupton and Phalen 1996). Trip ticket forms are used by state-licensed fish dealers to document all transfers of fish sold from coastal fishing waters from the fishermen to the dealer. The data reported on these forms include transaction date, area fished, gear used, and landed species as well as fishermen and dealer information.

The majority of trips reported to the NCTTP only record one gear per trip; however, as many as three gears can be reported on a trip ticket and are entered by the program's data clerks in no particular order. When multiple gears are listed on a trip ticket, the first gear may not be the gear used to catch a specific species if multiple species were listed on the same ticket but caught with different gears. In 2004, electronic reporting of trip tickets became available to commercial dealers and made it possible to associate a specific gear for each species reported. This increased the likelihood of documenting the correct relationship between gear and species.

##### **2.1.1.2 Sampling Intensity**

North Carolina dealers are required to record the transaction at the time of the transactions and report trip-level data to the NCDMF on a monthly basis. For further information on the sampling methodology for the NCTTP, see NCDMF 2019.

##### **2.1.1.3 Biological Sampling**

Biological sampling occurs during the spring and fall fishery. NCDMF personnel have a target of 600 samples from the spring fishery and 300 samples from the fall fishery. Fish are sampled monthly from various fish houses throughout the ASMA, throughout each season. Fish are measured to the nearest mm for fork length (FL) and TL and weighed to the nearest 0.01 kg. Sex is determined using the Sykes (1957) method and scales are removed from the left side of the fish, above the lateral line and between the posterior of the first dorsal fin and the insertion of the second dorsal fin. Scales are cleaned and pressed on acetate sheets using a Carver heated hydraulic press. NCDMF employees read scales using a microfiche reader set on 24x or 33x magnification. For each sex, a minimum of 15 scales per 25-mm size class is read and subsequently used to assign ages to the remainder of the sample.

##### **2.1.1.4 Potential Biases & Uncertainties**

All fish that are caught are not required to be landed (discards) or sold so some fish may be taken home for personal consumption and are not reported in the landings. The reporting of multiple



gears on a single trip ticket could also be a source of bias since the order in which gears are reported are not indicative of the primary method of capture.

#### **2.1.1.5 Development of Estimates**

Commercial landings were summarized by year using the NCTTP data. Length data collected from the commercial fish house sampling program were used to compute annual length-frequency distributions by sex.

#### **2.1.1.6 Estimates of Commercial Landings Statistics**

The NCTTP is considered a census of North Carolina commercial landings, though reliability of the data decreases as one moves back in time. Commercial landings were highest in the late 1960s and have substantially decreased through recent years (Figure 2.1). Landings have been constrained with a TAL since 1991.

The minimum lengths and ages observed in the commercial fisheries landings are strongly tied to the minimum length regulations at the time fish are collected, measured, and aged. The most noticeable impact is the implementation of the 18-inch minimum TL length limit in 1991; striped bass less than 45 cm TL (~18 inches; Figures 2.2, 2.3) and younger than age 3 (Figures 2.4, 2.5) have been rarely observed since 1991. The length and age compositions show that fewer larger and older fish have been observed in recent years (Figures 2.2–2.5).

### **2.1.2 Commercial Gill-Net Discards**

#### **2.1.2.1 Survey Design & Methods**

NCDMF's Program 466 (Onboard Observer Monitoring) was designed to monitor fisheries for protected species interactions in the gill-net fishery by providing onboard observations. Additionally, this program monitors finfish bycatch and characterizes effort in the fishery. The onboard observer program requires the observer to ride onboard the commercial fishermen's vessel and record detailed gill-net catch, bycatch, and discard information for all species encountered. Observers contact licensed commercial gill-net fishermen holding an Estuarine Gill-Net Permit (EGNP) throughout the state to coordinate observed fishing trips. Observers may also observe fishing trips from NCDMF vessels under Program 467 (Alternative Platform Observer Program), but these data were not used in this stock assessment due to the lack of biological data collected through the program.

#### **2.1.2.2 Sampling Intensity**

Fishing trips targeting striped bass are observed throughout the year; however, most observed trips occur during the fall when landings are the greatest in the Albemarle and the spring for the Pamlico Sound, both areas of which have a history of Atlantic sturgeon and sea turtle interactions.

#### **2.1.2.3 Biological Sampling**

Data recorded includes species, weight, length, and fate (landed, live discard, or dead discard).

#### **2.1.2.4 Potential Biases & Uncertainties**

Program 466 began sampling statewide in May 2010. To provide optimal coverage throughout the state, management units were created to maintain proper coverage of the fisheries. Management units were delineated based on four primary factors: (1) similarity of fisheries and management, (2) extent of known protected species interactions in commercial gill-net fisheries, (3) unit size, and (4) the ability of the NCDMF to monitor fishing effort. Total effort for each management unit

can vary annually based on fishery closures due to protected species interactions or other regulatory actions. Therefore, the number of trips and effort sampled each year by management unit varies both spatially and temporally.

Program 466 data do not span the entire time series for the assessment (no data are available for 1991–2000) and statewide sampling began in May 2010 decreasing the variability of observed trips with better spatial and temporal sampling beginning in 2012.

Striped bass discard data were not available in sufficient quantities to estimate discards or post-release mortality from commercial pound net or gig fisheries; however, these fisheries and others are known to have discards of striped bass. Additionally, commercial discards likely occur in other states, so the estimates presented here likely underestimate the total number of striped bass commercial discards removed from the A-R stock.

It is also important to note that this survey was designed to target trips that occur in times and areas where protected species interactions are highest; the program does not target striped bass trips. For this reason, a high number of zero-catch trips relative to striped bass occur in the data.

#### **2.1.2.5 Development of Estimates**

A generalized linear model (GLM) framework was used to predict striped bass discards in the A-R gill-net fishery based on data collected during 2012 through 2017. Only those variables available in all data sources were considered as potential covariates in the model. Available variables were year, season, mesh category (small: <5 inches and large:  $\geq 5$  inches) and management area (Figure 2.6), which were all treated as categorical variables in the model. Effort was measured as soak time (days) multiplied by net length (yards). Live and dead discards were modeled separately.

All available covariates were included in the initial model and assessed for significance using the appropriate statistical test. Non-significant covariates were removed using backwards selection to find the best-fitting predictive model. The offset term was included in the model to account for differences in fishing effort among observations (Zuur et al. 2009, 2012). Using effort as an offset term in the model assumes the number of striped bass discards is proportional to fishing effort (A. Zuur, Highland Statistics Ltd., personal communication).

Examination of the data indicated they were significantly zero inflated for both the live and dead discards. There are two types of models commonly used for count data that contain excess zeros. Those models are zero-altered (two-part or hurdle models) and zero-inflated (mixture) models (see Minami et al. 2007 and Zuur et al. 2009 for detailed information regarding the differences of these models). Minami et al. (2007) suggests that zero-inflated models may be more appropriate for catches of rarely encountered species; therefore, zero-inflated models were initially considered though were unable to converge. For this reason, zero-altered models were pursued.

The best-fitting model for live discards and for dead discards was applied to available effort data from the NCTTP to estimate the total number of live discards and dead discards for the A-R gill-net fishery.

In order to develop estimates of commercial discards for years prior to 2012, a hindcasting approach was used. The ratio of live or dead discards in numbers to A-R gill-net landings was computed by year for 2012 to 2017. As these ratios were variable among years (Figure 2.7), the working group decided to apply the median ratio over 2012 to 2017 separately for live and dead discards. The median ratio for either live or dead discards was multiplied by the commercial gill-

net landings in 1991 to 2011 to estimate the live and dead commercial gill-net discards for those years.

Because only dead discards were input into the assessment model, the estimates of live commercial gill-net discards were multiplied by 43%, an estimate of post-release mortality described in section 1.2.6.2. These estimates of live discards that did not survive were added to the estimates of commercial dead discards to produce an estimate of total dead discards for the commercial gill-net fishery for 2012 to 2017.

The available length samples from the NCDMF's Program 466 were summarized by year and used to characterize the length distribution of striped bass commercial discards by year.

#### **2.1.2.6 Estimates of Commercial Gill-Net Discard Statistics**

The best-fitting GLM for the commercial gill-net live discards assumed a zero-altered Poisson distribution (dispersion=2.9). The significant covariates for both the count and binary part of the model were year, season, mesh, and area. The best-fitting GLM for the dead discards assumed a zero-altered Poisson (dispersion=2.7). The significant covariates for the count part of the model were year, season, mesh, and area and the significant covariates for the binary part of the model were season and mesh.

Estimates of annual commercial dead discards ranged from a low of 2,500 striped bass in 2008 to a high of just over 11,600 striped bass in 2001 between 1991 and 2017 (Table 2.1; Figure 2.8). Total lengths of commercial discards have ranged from 10 cm to 85 cm (Figure 2.9). The majority of discards have been less than 60 cm TL.

#### **2.1.3 Albemarle Sound Recreational Fishery Monitoring**

From the 1950s through the late 1980s, various researchers conducted creel surveys in the Albemarle Sound and Roanoke River, although the Roanoke River has the most complete historical time series of catch and effort data (Hassler et al. 1981). Starting in 1988 and 1990 respectively, the NCWRC and NCDMF initiated annual creel surveys in the RRMA and ASMA that have continued to date.

##### **2.1.3.1 Survey Design & Methods**

The NCDMF collects catch and effort data through on-site interviews at boat ramps during allowed harvest days for each of four ASMA sampling zones (Figure 2.10). Statistics were calculated through a non-uniform probability access-point creel survey (Pollock et al. 1994). Site probabilities were set in proportion to the likely use of a site according to time of day, day of week, and season. Probabilities for this survey were assigned based on seasonal striped bass fishing pressure observed during past surveys, in addition to anecdotal information (S. Winslow and K. Rawls, NCDMF, personal communication). Probabilities can be adjusted during the survey period according to angler counts to provide more accurate estimates. Morning and afternoon periods were assigned unequal probabilities of conducting interviews, with each period representing half a fishing day. A fishing day was defined as one and a half hours after sunrise until one hour after sunset. These values varied among sites within zones due to differing fishing pressure.

##### **2.1.3.2 Sampling Intensity**

The ASMA striped bass creel survey data series includes estimates of effort, catch, and discards for years 1990–2017. The survey does not operate during the closed harvest season, so estimates of catch and release during this time are not available. In the early years of the survey when the

TAL was very low, the seasons may have only lasted a few days to a few weeks. In recent years as the TAL has increased, the harvest season occurs from October 1 through April 30. Creel clerks work all three weekend days (Friday–Sunday) and two weekdays. Interview sessions are approximately five hours and 45 minutes long, either in the morning or afternoon.

#### **2.1.3.3 Biological Sampling**

In the ASMA creel survey, all striped bass are sampled during the surveys and measured for TL (mm) and weighed to the nearest 0.1 kg by NCDMF personnel. No scales are collected for ageing purposes. Striped bass are not sexed during the creel survey.

#### **2.1.3.4 Potential Biases & Uncertainties**

One bias that has increased over time in the ASMA creel survey is the number of private access sites that are not included in the pool of public access points available to the survey. The increase in private sites is due to increased development of single-family dwellings and developments on the Albemarle Sound and tributaries in the last 20 years.

Another bias inherent in any non-uniform probability access-point creel survey is accurately matching the site probabilities to actual fishing pressure throughout the harvest season. Determining accurate probabilities is made more difficult when the harvest area is a large, open system such as a coastal estuary, and the species of interest is migratory in nature and movement (and hence fishing pressure) varies throughout the harvest area seasonally.

The bias associated with the increase in the number of private access points not included in the survey serves to systematically underestimate harvest and effort statistics, while the bias associated with varying probabilities throughout the season is not systematic and can produce under or over estimates of harvest and effort on an annual basis.

#### **2.1.3.5 Development of Estimates**

In the ASMA from 1990 to the spring season of 2005, a non-uniform probability roving access-point creel survey was used to estimate recreational hook-and-line effort and catch and release of striped bass during the allowed harvest seasons. Catch and effort data are collected daily for each of four ASMA sampling zones. Fishing effort was estimated by counting empty boat trailers at public and private boating access sites and using interview data to remove trailer counts for other users, including recreational fishermen targeting other species, hunters, recreational boaters, and commercial fishermen. Harvest was estimated as the product of catch rates and total fishing effort stratified by day and zone (Pollock et al. 1994).

In the ASMA from the fall of 2005 to present, angler catch statistics were calculated through a non-uniform probability access-point creel survey (Pollock et al. 1994). Site probabilities were set in proportion to the likely use of a site according to time of day, day of week, and season. Probabilities for this survey were assigned based on seasonal striped bass fishing pressure observed during past surveys, in addition to anecdotal information (S. Winslow and K. Rawls, NCDMF, personal communication). Probabilities can be adjusted during the survey period according to angler counts to provide more accurate estimates. Morning and afternoon periods were assigned unequal probabilities of conducting interviews, with each period representing half a fishing day. A fishing day was defined as one and a half hours after sunrise until one hour after sunset. These values varied among sites within zones due to differing fishing pressure. Harvest was estimated by applying the sample unit probabilities to interview data stratified by day and zone (Pollock et al. 1994).

Dead discards (no live) were input into the assessment model, so the estimates of Albemarle Sound recreational discards were multiplied by 6.4%, an estimate of post-release mortality described in section 1.2.6.2.

Lengths sampled from the Albemarle Sound recreational creel survey were used to characterize the length distribution of striped bass harvested by the Albemarle Sound recreational fishery by year.

In the absence of length samples from the recreational fisheries characterizing the releases, tagging data of striped bass recaptured by recreational anglers was used to develop length frequencies for the recreational releases. The composition of the total catch was derived first and then the length composition of the harvested fish was subtracted to estimate the length composition of the recreational releases. Due to the very low numbers of recaptured fish in some years, the recaptured fish length data were pooled across all years. For recaptures without lengths associated with them, if they were caught within three months of initial release, negligible growth was assumed and they were assigned a recapture length equal to the initial tagging length. The number of recaptures with associated lengths per year for the Albemarle Sound ranged from 3 to 127 with a mean of 39. Effective sample size was determined as the average number of unique locations and dates per year for recaptures in the associated management area. The proportion of fish recaptured per 2-cm length bin,  $t_l$ , was calculated from these pooled data such that:

$$t_l = \frac{\sum_{y=1997}^{y=2017} T_{y,l}}{\sum_{y=1997}^{y=2017} T_y}$$

where  $T_{y,l}$  is the number of fish tagged in year  $y$  and length bin  $l$ . A smoother was applied across the resulting proportion data using the following centrally-weighted five-point moving average:

$$Smoothed[t_l] = \frac{[t_{l-2} + t_{l-1} + 3t_l + 2t_{l+1} + t_{l+2}]}{9}$$

The length composition of the total catch per year and length bin,  $C_{y,l}$ , was then estimated as:

$$Smoothed[C_{y,l}] = Smoothed[t_l]C_y$$

where  $C_y$  is the total catch numbers of striped bass per year.

A smoother was applied to recreational harvest length frequencies,  $H_{y,l}$ , and the numbers of recreational releases per year and length bin,  $D_{y,l}$ , were then estimated as:

$$D_{y,l} = Smoothed[C_{y,l}] - [H_{y,l}]$$

In some instances, this produced length bins with negative discard values. The negative values were truncated to zero, and the data set for each year was then rescaled to match the original total number of releases per year.

### 2.1.3.6 Estimates of Albemarle Sound Recreational Fishery Statistics

Annual recreational harvest of striped bass in the Albemarle Sound has ranged from a low of 3,500 fish in 2010 to a high of just over 40,000 fish in 2001 (Table 2.2; Figure 2.11). No overall trend is apparent in the recreational harvest time series, but estimates in the most recent two years (2016 and 2017) are among the lowest observed since 1991.

Estimates of recreational dead discards in the Albemarle Sound have been variable from 1991 through 2017 (Table 2.2; Figure 2.12). Recreational dead discards have ranged from a low of 605 striped bass in 2006 to a high of over 5,800 striped bass in 1998.

The length distribution of recreational harvested striped bass has remained relatively consistent from 1996 through 2017 (Figure 2.13). The majority of lengths fall between 45 and 60 cm TL. Lengths of striped bass observed in the Albemarle Sound recreational discards have also demonstrated consistency over the years in which lengths are available (1997–2017; Figure 2.14); the majority of these recreational discards range between 40 and 60 cm TL.

## **2.1.4 Roanoke River Recreational Fishery Monitoring**

### **2.1.4.1 Survey Design & Methods**

The NCWRC conducts the RRMA striped bass creel survey to estimate angler effort, catch, and harvest during the spring harvest season. In some years, estimates of angler effort and catch and release of striped bass after the harvest season closes are also made (depending on available funding). The creel survey employs a non-uniform probability, stratified access-point creel survey design (Pollock et al. 1994) to estimate recreational fishing effort (angler hours, and angler trips), harvest of striped bass, and numbers of striped bass caught and released. The creel survey is stratified by area (upper zone or lower zone), time (AM or PM), and type of day (weekdays and weekend days). The upper zone includes the river segment from Roanoke Rapids Lake dam downstream to the U.S. Highway 258 Bridge near Scotland Neck (Figure 2.15). The lower zone extends from U.S. Highway 258 Bridge downstream to Albemarle Sound. Because past analyses depict differential catch rates through progression of the open harvest season, the survey was stratified into two-week sample periods. Within periods, samples and estimates are further stratified by type of day because fishing effort and catch is also known to vary as a function of day type. Selection of access points where interviews occurred was based on probability of boat trailer counts generated from prior RRMA creel surveys as well as expert opinion by biological and enforcement staff. Probabilities of fishing activity for time of day (0.4 for AM and 0.6 for PM during periods one and two and equal probabilities during all other periods) are estimated based upon prior experience with the RRMA striped bass fishery.

### **2.1.4.2 Sampling Intensity**

The RRMA striped bass creel survey data series includes 1988–2017 for harvest season estimates and 1995–1999, 2005–2008, and 2010–2017 for closed season catch and effort estimates. The creel survey is conducted during March, April, and May of each year. Creel clerks typically work two weekdays and both weekend days each week. Interview sessions last three hours and one session is conducted in each zone each sample day.

### **2.1.4.3 Biological Sampling**

RRMA striped bass creel clerks record the total number of striped bass caught and the number of striped bass harvested. Creel clerks measure TL (mm), weight (kg), and determine sex of each striped bass harvested when possible. Counts and total weights of harvested striped bass (i.e., no individual data) are recorded for angling parties when interview sessions are busy. In some years, creel clerks also record the number of striped bass released within length limit categories (e.g., short, legal, slot, over-slot), type of bait used, angler residency, and trip expenditures.

#### **2.1.4.4 Potential Biases & Uncertainties**

In the RRMA creel survey, sample unit probabilities are adjusted each year depending on current conditions and expected trends in angler effort. Additionally, construction of new boating access areas has necessitated addition and deletion of creel locations. The NCWRC Jamesville-Astoria Rd. boating access area was added to the survey in 2011, and the two private ramps in Jamesville were subsequently removed from the survey. In 2016, a new boating access area in Lewiston-Woodville was added to the survey. Calculation of fishing effort was made using expansions of trailer count data from 1988–2001, but from 2002–2017, fishing effort was calculated by expanding interview data by the sample unit probability.

#### **2.1.4.5 Development of Estimates**

From 1988–2001, total fishing effort was estimated from counts of empty boat trailers at boating access areas along the entire river. Trailer counts were conducted each day of the open season. Total numbers of anglers were estimated by expanding trailer counts by the mean number of anglers per party as determined from interviews at access areas. The starting point for effort counts was randomly selected. Counts were made during mid-morning, or mid-afternoon periods. Based on interview data, trailer counts were adjusted to eliminate commercial fishermen, hunters, and recreational boaters. Data were adjusted based on the proportion of recreational anglers interviewed by creel clerks within each zone by period and kind of day. Harvest was estimated as the product of catch rates and total fishing effort stratified by period, zone, and kind of day (weekday or weekend day).

From 2002–2017, a specifically designed creel survey program was used to provide estimates of catch, harvest, and effort using formulas derived from Pollock et al. (1994). Estimates of striped bass catch, harvest, and effort for each sample day were made by expanding interview data by the sample unit probability (product of the access point probability and time of day probability). Within sample periods, catch, harvest, and effort estimates for weekdays and weekend days are separately averaged. The averages are then expanded to the total number of days of each type for that sample period. Separate estimates of total catch, harvest, and effort are made for each zone. Finally, sample period and zone totals are added to calculate the annual estimates.

Only dead discards were input into the assessment model, so the estimates of Roanoke River recreational discards were multiplied by 6.4%, an estimate of post-release mortality described in section 1.2.6.2.

As discard estimates were only available starting in 1995, a hindcasting approach was used to develop estimates back to 1991. The ratio of dead discards to harvest in numbers was calculated for 1995 through 2017 (Figure 2.16). The median ratio over those years was multiplied by the Roanoke River recreational harvest in 1991 to 1994 to estimate the dead discards for these earlier years.

Lengths sampled from the Roanoke River recreational creel survey were used to characterize the length distribution of striped bass harvested by the Roanoke River recreational fishery by year.

Roanoke River discard length compositions were derived using the same methodology as the Albemarle Sound discard length compositions described in section 2.1.3.5. The number of recaptures with associated lengths per year for the Roanoke River ranged from 18 to 191 with a mean of 88.

#### **2.1.4.6 Estimates of Roanoke River Recreational Fishery Statistics**

Estimates of recreational harvest in the Roanoke River have ranged from a low of about 3,100 fish in 1985 to a high of just over 38,000 fish in 2000 (Table 2.3; Figure 2.17). Recreational harvest increased from the beginning of the time series in 1982 to the early 2000s. Since then, recreational harvest in the Roanoke River has shown an overall slight decline.

Discards from the Roanoke River recreational fishery have been variable (Table 2.3; Figure 2.18). Estimates have ranged from a low of 4,215 striped bass in 2017 to a high of over 18,600 striped bass in 1997. There is no clearly discernable trend in these discard estimates over time.

As was observed with the Albemarle Sound recreational harvest and discard lengths, there was consistency in the total lengths observed in the Roanoke River recreational harvest (Figure 2.19) and discards (Figure 2.20) observed over time. The majority of striped bass collected from the Roanoke River recreational fishery were between 40 cm and 55 cm TL for both the harvest and discards.

### **2.2 Fisheries-Independent**

#### **2.2.1 Juvenile Abundance Survey (Program 100)**

##### **2.2.1.1 Survey Design & Methods**

The NCDMF Juvenile Anadromous Survey, also known as Program 100 (P100), targets young-of-year (YOY) striped bass using a bottom trawl in Albemarle Sound. The survey was taken over by the NCDMF in 1984 and continues to sample the same seven fixed stations in western Albemarle Sound initiated in 1955 by Dr. William Hassler of N.C. State University, making it one of the longest continuous time series of striped bass fisheries-independent abundance data on the east coast (Figure 2.21). The sampled habitats are preferred nursery habitat for YOY striped bass in the Albemarle Sound as they increase in size and move from near-shore nursery areas to more open water habitats (Hassler et. al 1981).

The survey uses an 18-foot semi-balloon trawl with a body mesh size of 0.75-inch bar mesh and a 0.125-inch bar mesh tail bag. Tow duration is 15 minutes. Temperature, salinity, and dissolved oxygen are recorded.

##### **2.2.1.2 Sampling Intensity**

Trawl sampling is conducted bi-weekly for eight weeks starting in mid-July at seven established locations in the western Albemarle Sound area for a total of 56 samples. Trawl sites are located at the edge of breaks and contours, usually within the 2.4 m–3.7 m (8 feet–12 feet) depth profile.

##### **2.2.1.3 Biological Sampling**

All striped bass captured are counted and a subsample (maximum of 30) is measured (mm; TL and FL). In the event a striped bass is captured that may overlap with the size range of a YOY and a 1-year old striped bass, the specimen is brought back to the lab for examination of otoliths and/or scale samples to determine its age. In recent years, a subsample of YOY and age-1 striped bass has been weighed to the nearest gram for improved length at age relationships.

##### **2.2.1.4 Potential Biases & Uncertainties**

The Juvenile Abundance Survey is a fixed survey that the division appropriated from another source, so the fixed stations were retained for the continuity of data. A fixed-station survey can run the risk of bias if the sites selected do not adequately represent the sampling frame.



Additionally, even if the sites adequately cover the sampling frame, the increased variation that would come about from sampling randomly is not accounted for and is therefore at risk of being neglected.

Indices derived from fixed-station surveys such as P100 may not accurately reflect changes in population abundance (Warren 1994, 1995). The accuracy of the estimates is tied to the degree of spatial persistence in catch data of the species (Lee and Rock 2018). The persistence of the P100 data were evaluated following the approach of Lee and Rock (2018) and results suggested a lack of year\*station interaction, which indicates the presence of spatial persistence and so suggests the survey is likely tracking trends in relative abundance.

#### **2.2.1.5 Development of Estimates**

A nominal index was calculated by year using a standard arithmetic mean (numbers per tow). A generalized linear model (GLM) framework was also used to model the relative abundance of YOY striped bass. Potential covariates were evaluated for collinearity by calculating variance inflation factors. Collinearity exists when there is correlation between covariates and its presence causes inflated p-values. The Poisson distribution is commonly used for modeling count data; however, the Poisson distribution assumes equidispersion; that is, the variance is equal to the mean. Count data are more often characterized by a variance larger than the mean, known as overdispersion. Some causes of overdispersion include missing covariates, missing interactions, outliers, modeling non-linear effects as linear, ignoring hierarchical data structure, ignoring temporal or spatial correlation, excessive number of zeros, and noisy data (Zuur et al. 2009, 2012). A less common situation is underdispersion in which the variance is less than the mean. Underdispersion may be due to the model fitting several outliers too well or inclusion of too many covariates or interactions (Zuur et al. 2009).

Data were first fit with a standard Poisson GLM and the degree of dispersion was then evaluated. If over- or underdispersion was detected, an attempt was made to identify and eliminate the cause of the over- or underdispersion (to the extent allowed by the data) before considering alternative models, as suggested by Zuur et al. (2012). For example, the negative binomial distribution allows for overdispersion relative to the Poisson distribution whereas a quasi-Poisson GLM can be used to correct the standard errors for overdispersion. If the overdispersion is the result of an excessive number of zeros (more than expected for a Poisson or negative binomial), then a model designed to account for these excess zeros can be applied. There are two types of models that are commonly used for count data that contain excess zeros: zero-altered (two-part or hurdle models) and zero-inflated (mixture) models (see Minami et al. 2007 and Zuur et al. 2009 for detailed information regarding the differences of these models). Minami et al. (2007) suggests that zero-inflated models may be more appropriate for catches of rarely encountered species; therefore, zero-inflated models were considered here when appropriate.

All available covariates were included in the initial model and assessed for significance using the appropriate statistical test. Non-significant covariates were removed using backwards selection to find the best-fitting predictive model.

#### **2.2.1.6 Estimates of Survey Statistics**

Available covariates were year, depth, surface and bottom temperature, and surface and bottom salinity. The best-fitting GLM model assumed a negative binomial distribution (dispersion=1.4) and the significant covariates were year and bottom temperature.

The nominal and GLM-standardized indices were similar throughout the time series (Figure 2.22). Both exhibit substantial inter-annual variability over time.

## **2.2.2 Independent Gill-Net Survey**

### **2.2.2.1 Survey Design & Methods**

In October 1990, the NCDMF initiated the Striped Bass Independent Gill-Net Survey, also known as Program 135 (P135). The survey was designed to monitor the striped bass population in the Albemarle and Croatan sounds.

The survey follows a random stratified design, stratified by geographic area. This survey divides the water bodies comprising the Albemarle region into six sample zones that are further subdivided into one-mile square quadrants with an average of 22 quadrants per zone (Figure 2.23). Albemarle Sound, Croatan Sound, and Alligator River sample zones (Zones 2–7) were selected for this survey, based on previous sampling and historical abundance information (Street and Johnson 1977). Sampling in Zone 1 was discontinued shortly after the survey began in favor of sampling Zone 7, to allow for tagging to produce estimates of mixing of the Albemarle-Roanoke striped bass stock and the migratory portion of the Atlantic migratory stock which may utilize the eastern portion of the Albemarle Sound during the winter months while overwintering. The survey gear is a multi-mesh monofilament gill net. Four gangs of twelve meshes (2.5-, 3.0-, 3.5-, 4.0-, 4.5-, 5.0-, 5.5-, 6.0-, 6.5-, 7.0-, 8.0-, 10.0-inch stretched mesh, ISM) of gill nets are set in each quadrant by the fishing crew. One two-gang set is weighted to fish at the bottom (sink net), and the other is floating unless the area is unsuitable for gill-net sampling (marked waterways and areas with excessive submerged obstructions). The use of 12 different mesh sizes allowed for the capture of fish age one and older. Alternate zones and quadrants are randomly selected if the primary selection cannot be fished. A fishing day is defined as the two crews fishing the described full complement of nets for that segment for one day. One unit of effort is defined as each 40-yard net fished for 24 hours.

The fishing year is divided into two segments: (1) fall/winter survey period, 1 November through 28 February; and (2) spring survey period, 1 March through late May. The sampling methods remain the same during each sampling season. Areas fished, sampling frequency, and sampling effort is altered seasonally.

For the fall/winter segment, two survey crews fish replicate 40-yard anchored, floating, and sinking monofilament gill nets from 2.5- to 4.0- ISM in one-half inch increments with a twine size of 0.33 mm (#104), 5.0- to 7.0-ISM with a twine size of 0.40 mm (#139), and 8.0-ISM and 10.0-ISM, with a twine size of 0.57 mm (#277). Heavier twine sizes in the larger mesh nets are intended to improve retention of larger, heavier fish. Gill nets were constructed with a hanging coefficient of 0.5. Gear soak time is 48 hours for each selected quadrant.

In the spring segment, gill-net effort is concentrated in western Albemarle Sound (Zone 2) near the mouth of the Roanoke River (Figure 2.23). The shift to Zone 2 was designed to increase the chance of intercepting mature striped bass congregated in this area during their migration to the Roanoke River spawning grounds. Effort is concentrated in this zone to determine differences in the size, age, and sex composition of the spring spawning migration relative to the fall/winter resident population. Zone 2 is sub-divided into southern and northern areas.

#### **2.2.2.2 Sampling Intensity**

The NCDMF monitors the adult striped bass population in Albemarle Sound through spring (March–May) and fall (November–February). The fishing year is divided into two segments: (1) fall/winter survey period, 1 November through 28 February; and (2) spring survey period, 1 March through late May. All zones are sampled equally, except in the spring when effort is shifted to Zone 2. Each crew samples each of the six zones, providing 24 fishing days per month and a total of 96 fishing days for the season. A fishing day is defined as one crew, fishing the full complement of nets specified, for that segment for one day (24 hours).

The southern area, adjacent to the Roanoke River, received increased effort at a 2:1 ratio south to north, based on the historical seasonal abundance of mature striped bass (Harris et al. 1985). Quadrants sampled are randomly selected as previously noted. Fishing effort is conducted continuously, seven days a week weather permitting, until the end of late May.

#### **2.2.2.3 Biological Sampling**

All striped bass are counted and measured and healthy striped bass that survived entanglement are tagged with internal anchor tags and then measured to the nearest mm for FL and TL. Scales are removed from the left side of the fish, above the lateral line and between the posterior of the first dorsal fin and the insertion of the second dorsal fin. When possible, sex is determined by applying directional pressure to the abdomen towards the vent and observing the presence of milt or eggs.

For both the fall/winter and spring segment, fish that did not survive entanglement are processed at the NCDMF laboratory. Fish are measured to the nearest mm for FL and TL and weighed to the nearest 0.01 kg. Sex is determined by visual inspection and scales are removed as previously described. Scales are cleaned and pressed on acetate sheets using a Carver heated hydraulic press. Scales are read using a microfiche reader set on 24x or 33x magnification. For each sex, a minimum of 15 scales per 25 mm size class is read and subsequently used to assign ages to the remainder of the sample.

#### **2.2.2.4 Potential Biases & Uncertainties**

The P135 Survey deploys a passive gear of an array of nets with varying mesh size over a variety of randomly selected locations. The effort expended on survey design should result in estimates with relatively low bias. The survey design was informed by previous abundance and sampling data. It is possible that changes in the stock (habitat use, migration corridors, etc.) since the implementation of the sampling program may cause estimates to vary.

Many factors affect gill-net catch efficiency including net visibility and turbidity (Berst 1961; Hansson and Rudstam 1995), though setting nets overnight may offset some concerns of net visibility. Efficiency can also decrease if nets become tangled or fouled with debris. In the P135 Survey, performance of individual net panels is evaluated and recorded and catch is evaluated at the sample level (catch from a gang of nets is a sample), so performance of individual net panels may not have a large impact on catch from a sample.

#### **2.2.2.5 Development of Estimates**

Nominal indices of abundance were developed for both the fall/winter and spring components of the P135 Survey and were calculated using stratified average estimator (numbers per gang of net, 480 yards of 12 mesh sizes). For both the fall/winter and spring segments, only catches observed during the first 24 hours of the soak were included in the development of the index. Standardized indices were also calculated using the GLM approach described in section 2.2.1.5.

Biological data collected during the survey were summarized to characterize both the length and age frequencies of striped bass observed by sex and survey component.

#### **2.2.2.6 Estimates of Survey Statistics**

Available covariates for the GLM standardization included year, quad (fall/winter only), depth, and surface temperature. The best-fitting GLM for the fall/winter index assumed a negative binomial distribution (dispersion=1.6) and the significant covariates were year, quad, and surface temperature. The best-fitting GLM for the spring index assumed a negative binomial distribution (dispersion=1.5) and the significant covariates were year, depth, and surface temperature.

The GLM-standardized indices tracked well with the nominal indices for both the fall/winter (Figure 2.24) and spring (Figure 2.25) components of the P135 Survey. Indices from both components of the survey indicate decreasing trends in the most recent years of the time series (Figures 2.24, 2.25).

Females observed during the fall/winter component of the P135 Survey have ranged from 15 cm to 95 cm TL and males have ranged from 15 cm to 80 cm TL (Figure 2.26). Striped bass observed during the spring component of this survey were generally larger; females have ranged from 20 cm to 115 cm TL and males have ranged from 15 cm to 90 cm TL (Figure 2.27).

Females ranging from ages 1 to 10 have been collected during the fall/winter component of the P135 Survey (Figure 2.28). Males collected during the fall/winter have ranged in age from 1 to 7. Older striped bass tend to be observed during the spring component of this survey (Figure 2.29). Female striped bass as old as 15 and males as old as 10 have been observed in the spring. The modal age has varied over time for both females and males in both the fall/winter and spring components of the P135 Survey.

### **2.2.3 Roanoke River Electrofishing Survey**

#### **2.2.3.1 Survey Design & Methods**

The NCWRC Electrofishing Survey on the Roanoke River spawning grounds began in 1991 to meet the ASMFC FMP requirements to monitor spawning stock abundance (Figure 2.30). A boat-mounted electrofishing unit (Smith-Root 7.5 GPP) is used (1 dip netter) to capture fish during daylight hours. Sampling is conducted at stations within strata. Sampling stations are located on main and secondary river channel habitats. Three strata are sampled each day, and strata selection is dependent on flow conditions. Flows of approximately 7,000 cubic feet per second (cfs) or less restrict access to strata above the rapids in proximity to the Weldon boating access area. To minimize size selection during sampling, striped bass were netted as they were encountered regardless of size. Water temperature (°C) is recorded each sample day.

#### **2.2.3.2 Sampling Intensity**

NCWRC personnel collect striped bass weekly between mid-April and May, on the historic spawning grounds of the Roanoke River near Weldon (RM 130) and Roanoke Rapids (RM 137), North Carolina. Sampling begins as the water temperature approaches 15.0°C (59.0°F) and continues through the range of optimal spawning temperatures until water temperatures surpass 22°C or until striped bass spawning is complete; optimum spawning temperatures range from 18.0° to 22.0°C (64.4° to 71.6°F) for striped bass in the Roanoke River.

### **2.2.3.3 Biological Sampling**

Information on sex, age, and size composition of the spawning stock is also collected. Each fish is measured to the nearest mm for TL and sex is determined by assessing the presence of eggs or milt when pressure is applied to the fish's abdomen. Weight (kg) and scales are obtained from a subsample (target maximum of five fish of each 25-mm size group and sex per sample day) of fish. Weight and scales are collected from all fish greater than 700 mm. Scales are removed from the left side of the fish, above the lateral line and between the posterior of the first dorsal fin and the insertion of the second dorsal fin. Scales are aged using an EyeCom 3000 microfiche reader at 24x or 36x magnification. A primary reader ages up to 15 individuals per 25-mm length group per sex, and a subsample (20% of aged scales) is aged by a secondary reader for age verification. Age discrepancies between the readers are reconciled in concert.

### **2.2.3.4 Potential Biases & Uncertainties**

The electrofishing survey spans a seven-mile section of the Roanoke River, determined to be the spatial extent of the spawning grounds. Site selection in early years of the survey was opportunistic to some degree, but multiple strata were always sampled so that sites were spread out within the spawning habitat/survey area each sample day. In more recent years, sites have been randomly selected within each of the three strata and the strata selections are based on flow conditions; however, some sample sites cannot be sampled due to flow conditions or angling activity. Inability to access sampling sites due to flow conditions or angler presence could bias the abundance estimates either by concentrating striped bass in the accessible areas or allowing striped bass to go undetected. Additionally, it is possible that fish may be missed by the dip netter. If striped bass are not universally available to the dip netter at all population densities, it could bias abundance estimates.

Other biases could be due to the gear itself; striped bass of abnormal size may not be as vulnerable to the stunning effects of the electrofishing gear and could escape capture. Electrofishing tends to select for larger fish as they are more visible to the dip netters and have a lower immobilization threshold (Sullivan 1956; Reynolds 1996; Dolan and Miranda 2003; Ruetz et al. 2007). For this reason, the relative abundance of smaller fish is likely biased too low (Reynolds 1996). Collection of fish by netting may be associated with bias. Daugherty and Sutton (2005) demonstrated that capture efficiency was affected by moderate flow rates due to movement of fish out of range of the netters. Schoenebeck and Hansen (2005) indicated how gear saturation caused electrofishing catch rate to be non-linearly related to abundance. Some fish may be less likely to be immobilized by electrofishing gear. Dolan and Miranda (2003) demonstrated how immobilization thresholds were inversely proportional to body size. Conductivity, water temperature, water transparency, dissolved oxygen, depth, flow, and electric current are some of the factors that can impact the efficiency of electrofishing gear (Reynolds 1996; McInerney and Cross 2000; Speas et al. 2004; Buckmeier and Schlechte 2009).

### **2.2.3.5 Development of Estimates**

A nominal index was calculated using a ratio estimator (numbers per minute; Pollock et al. 1994). A standardized index was also calculated using the GLM approach described in section 2.2.1.5. An offset term was included in the model to account for differences in survey effort (measured in minutes) among sampling events (Zuur et al. 2009, 2012).

Biological data collected during the survey were summarized to characterize both the length and age frequencies of striped bass observed by sex.

### **2.2.3.6 Estimates of Survey Statistics**

Available covariates for the GLM were year, stratum, discharge, and temperature. The final best-fitting model assumed a negative binomial distribution (dispersion=1.3) and the significant covariates were year, stratum, and temperature. The nominal and GLM-standardized indices were similar throughout the time series (Figure 2.31). Both series exhibit inter-annual variation and both demonstrate a general declining trend since the early 2000s.

The total lengths of females observed in the Roanoke River Electrofishing Survey have ranged from 20 cm to 120 cm TL (Figure 2.32). Males have ranged in length from 10 cm to 110 cm TL. Some truncation of the length distributions is apparent in the most recent years of the survey.

A broad range of ages have been collected during this survey (Figure 2.33). Females have ranged in age from 1 to 17 years while males have ranged in age from 1 to 15 years. The age distributions have shown a truncation in the last few years of the survey.

## **3 ASSESSMENT**

### **3.1 Method—Stock Synthesis**

#### **3.1.1 Scope**

The unit stock was defined as all striped bass within the ASMA and RRMA.

#### **3.1.2 Description**

This assessment is based on a forward-projecting length-based, age-structured model. A two-sex model is assumed. The stock was modeled using Stock Synthesis (SS) text version 3.30.14 software (Methot 2000; Methot and Wetzel 2013; Methot et al. 2019). Stock Synthesis is an integrated statistical catch-at-age model that is widely used for stock assessments throughout the world. SS was also used to estimate reference point values. All input files are available upon request.

#### **3.1.3 Dimensions**

The assessment model was applied to data collected from within the range of the assumed biological stock unit (ASMA-RRMA; section 1.2.1).

The time period modeled was 1991 through 2017 using an annual time step based on the calendar year. The year 1991 was selected as the start year because it was the earliest year for which landings from the Albemarle Sound recreational fleet were available (section 2.1.3). The terminal year, 2017, was selected because it was the most recent year from which data were available at the start of the assessment process.

#### **3.1.4 Structure / Configuration**

##### **3.1.4.1 Catch**

The model initially incorporated three fishing fleets: ASMA commercial fishery (ARcomm), ASMA recreational fishery (ASrec), and the RRMA recreational fishery (RRrec). Landings (i.e., “retained” catch) were entered for each of these fleets (ARcomm: weight; ASrec: numbers; RRrec: numbers; Table 3.1; Figure 3.1). Dead discards (in numbers) were also included for each of the three fleets (Table 3.2; Figure 3.2). After evaluation of initial model runs, it was decided to treat the RRrec discards as a separate fleet (see section 3.1.4.8).

#### **3.1.4.2 Survey Indices**

Four indices of relative abundance were selected for input into the model. All indices were derived from fisheries-independent surveys (Table 3.3; Figure 3.3). The index derived from the Program 100 Juvenile Trawl Survey (P100juv) was input as an index of age-0 recruitment and so associated biological data (lengths or ages) were not required as inputs into the model. Indices derived from the fall/winter component of the Program 135 Independent Gill-Net Survey (P135fw), the spring component of the Program 135 Independent Gill-Net Survey (P135spr), and the Roanoke River Electrofishing Survey (RRef) were also used.

Changes in indices over time can occur due to factors other than changes in abundance; the fisheries-independent indices were standardized using a GLM approach to attempt to remove the impact of some of these factors (Maunder and Punt 2004; see sections 2.2.1–2.2.3). Catchability ( $q$ ) was assumed to be time-invariant for each survey and all survey indices were assumed to have a linear relation to abundance.

#### **3.1.4.3 Length Composition**

Annual length frequencies were input for each fleet's landings and discards for the years in which lengths were available for the particular fleet (see sections 2.1.1–2.1.3). Annual length frequencies characterizing the P135fw, P135spr, and RRef surveys were also input (see sections 2.2.2 and 2.2.3). Where possible, sex-specific length frequencies were used. Length frequencies were input by 2-cm length bins ranging from 10 cm to 130 cm TL.

#### **3.1.4.4 Age Composition**

Annual sex-specific age data were input for the AScomm landings as well as the P135fw, P135spr, and RRef surveys. The age data were input as raw age-at-length data, rather than age compositions generated from applying age-length keys to the catch-at-length compositions. The input compositions are therefore the distribution of ages obtained from samples in each length bin (conditional age-at-length). This approach is considered a superior approach because it avoids double use of fish for both age and length information, it contains more detailed information about the age-length relationship and so improves the estimation of growth parameters, and the approach can match the protocols of sampling programs where age data are collected in a length-stratified program (Methot et al. 2019).

Age 15 was treated as a plus group that included ages 15 through 17, the maximum age within the data input into the stock assessment model. Ages were assumed to be associated with small bias and negligible imprecision.

#### **3.1.4.5 Biological Parameters**

##### *Natural Mortality*

Natural mortality is one of the most important parameters in a stock assessment and one of the most difficult to estimate. The availability of an empirical estimate is rare. The empirical estimate of natural mortality from the Harris and Hightower (2017) study (0.72, see section 1.2.6.1) was assumed for both females and males in the model presented to the peer reviewers (see section 5) and treated as an age-invariant, fixed input. While the peer reviewers were pleased with the working group's attempt to incorporate an empirical estimate of natural mortality, they felt the value was too high given the species maximum age (see section 1.2.6.1).

Given the uncertainty in the assumed rate of natural mortality, a series of sensitivity runs were performed at the second peer review workshop in which the assumption regarding natural mortality

was varied (see section 3.1.7.2). The values assumed for natural mortality in these runs were selected from the range estimated based on the species life history (Table 1.4; section 1.2.6.1). After discussion between the working group and the peer review panel, a value of 0.40 was settled on for use in the final base run. This value was assumed for both sexes and treated as an age-invariant, fixed input. Both the working group and the peer review panel felt this value was more appropriate given the species' life history and maximum age and was closer to the empirical estimate of natural mortality estimated in the Harris and Hightower (2017) study than other values explored.

### *Growth*

Growth (age-length) was assumed to be sex specific and was modeled using the von Bertalanffy growth curve. In the SS model, when fish recruit at the real age of 0.0, their length is set equal to the lower edge of the first population length bin (here, 10 cm; Methot et al. 2019). Fish then grow linearly until they reach a real age equal to a user-specified age (here, age 1). As the fish continue to age, they grow according to the von Bertalanffy growth equation.

Allowing SS to estimate the growth curve ensures that the assumptions about selectivity are consistent with other parts of the model and that uncertainty in the growth estimates is incorporated into the estimates of spawning stock biomass, fishing mortality, and reference points (Hall 2013). All age-length growth parameters were estimated for both sexes. The estimated growth parameters for each sex were  $L_{\infty}$ ,  $K$ , coefficient of variation (CV) for length at age 1, and CV for  $L_{\infty}$ . Initial values for  $L_{\infty}$  and  $K$  were derived by fitting the von Bertalanffy model to the available age-length data by sex (see also section 1.2.4; Table 1.1). Initial values for the CVs for length at age 1 and  $L_{\infty}$  were derived empirically for each sex. The initial values for the growth parameters were treated as informative priors (prior standard deviation=0.05 for  $L_{\infty}$  and  $K$ ; prior standard deviation=0.8 for CV1 and CV2) assuming a normal distribution. Examination of the observed data was used to set reasonable bounds on all growth parameters for males and females.

Parameters of the length-weight relationship were fixed (i.e., not estimated) for both males and females. The assumed values were those estimated in this report as described in section 1.2.4 (Table 1.2).

### *Maturity & Reproduction*

Female maturity at age as estimated by Boyd (2011; section 1.2.5.4) was treated as a fixed input in the model. Reproduction was assumed to occur on January 1 each year.

### *Fecundity*

The selected fecundity option in SS was such that causes eggs to be equivalent to spawning biomass.

#### **3.1.4.6 Stock-Recruitment**

A Beverton-Holt stock-recruitment relationship was assumed. Virgin recruitment,  $R_0$ , was estimated within the model. Steepness,  $h$ , was fixed at 0.9 and the standard deviation of  $\log(\text{recruitment})$ ,  $\sigma_R$ , was fixed at 0.6. Recruitment deviations were estimated from 1980 to 2015. The deviations are assumed to sum to zero over this time period. Setting the first year in which to estimate recruitment deviations (1974) earlier than the model start year (1991) allows for a non-equilibrium age structure at the start of the assessment time series (Methot et al. 2019).



#### **3.1.4.7 Fishing Mortality**

SS allows several options for reporting fishing mortality ( $F$ ). The  $F$  values reported here represent a real annual  $F$  calculated as a numbers-weighted  $F$  (see Methot et al. 2019) for ages 3–5. This age range was selected based on the high selectivity for this age range by the fleets and the large percentage of the total catch this age range comprises. Note the last NCDMF stock assessment for striped bass reported apical  $F$  values ( $F$  at age 4) and so are not directly comparable to the results of this assessment (Flowers et al. 2016).

#### **3.1.4.8 Selectivity**

In SS, selectivity can be a function of length and/or age. In the current assessment, selectivity was assumed to be a function of length for all fleets and surveys due to the high confidence in the length data for characterizing these data sources. Retention for the fleets was also assumed to be a function of length (the only option for retention parameters).

In initial runs, all selectivity patterns were modeled using the recommended double normal curve. The double normal curve is extremely flexible and can take on shapes ranging from asymptotic to dome shaped. Evaluation of the initial model fits to the length composition data indicated some potential issues with the predicted selectivity patterns (i.e., strong patterns in the length residuals). Fits to the RRrec harvest lengths were especially poor so the decision was made to fix the selectivity to match the protective slot (section 1.5.4) and treat the discard portion of this fishery as a separate fleet. The presence of strong residual patterns in the fits to the length composition data prompted consideration of an even more flexible selectivity function, the cubic spline. Use of the cubic spline for the ARcomm fleet (six nodes) and the P135fw survey (three nodes) provided improvements in fits to the length composition data associated with these fleets and so was assumed in the final base model.

Early model runs suggested difficulty in predicting the female and male length composition data from the RRef survey. Investigation of the data and discussion with the model developer suggested this was due to the highly skewed sex ratio and different length frequency patterns between female and male striped bass observed in the survey. The SS model allows for selectivity for male fish to differ from selectivity for female fish and this option was selected for the RRef survey. The male selectivity parameters were modeled as an offset of the female selectivity parameters.

#### **3.1.4.9 Equilibrium Catch**

The SS model needs to assume an initial condition of the population dynamics for the period prior to the estimation period. Typically, two approaches are used to meet this assumption. The first approach starts the model as far back as necessary to satisfy the notion that the period prior to the estimation of dynamics was in an unfished or near unfished state. For striped bass, reliable catch records back to the start of the fishery are not available. For this reason, the model developer recommended use of the second approach, which is to estimate (where possible) initial conditions assuming equilibrium catch (R.D. Methot Jr., NOAA Fisheries, personal communication). The equilibrium catch is the catch taken from a fish stock when it is in equilibrium with removals and natural mortality balanced by stable recruitment and growth.

#### **3.1.5 Optimization**

The SS model assumes an error distribution for each data component and assigns a variance to each observation. The ARcomm landings, ASrec and RRrec harvests, and RRrec discards were fit in the model assuming a lognormal error structure. These data were assumed precise and assigned

a minimal observation error. The standard errors (SEs) of the annual ARcomm landings were assumed equal to 0.02 prior to the start of the Trip Ticket program (1994; section 2.1.1) and were assumed equal to 0.01 for the remainder of the time series. As the commercial landings data are derived from a census and recreational data are derived from a survey, a slightly higher standard error was assumed for the annual ASrec and RRrec harvest estimates ( $SE=0.02$ ). The RRrec discard estimates were based on a hindcast method in earlier years (1991–1994) of the time series and were assumed to have a CV equal to 0.06. Discard estimates from this fleet in subsequent years were assumed to have a CV equal to 0.04.

As dead discards are part of the overall total removals, they were also assumed to be precise, though were assumed to have higher variance than the landings and harvest due to the increased uncertainty in the estimation methods. The coefficient of variation (CV) assumed for the ARcomm discards was derived from the GLM standardization (see section 2.1.2.5). The CVs for discards from the ASrec fleet were derived empirically. A normal distribution was assumed for the error structure of the discards for each fleet.

Survey indices were fit assuming a lognormal error distribution with variance estimated from the GLM standardization.

Composition information was fit assuming a multinomial error structure with variance described by the effective sample size. For each fleet and survey, the effective sample size was the number of sampled trips and a maximum of 200 was imposed.

The objective function for the base model included likelihood contributions from the landings and harvest, discards, survey indices, length compositions, age data, and recruitment deviations. The total likelihood is the weighted sum of the individual components. All likelihood components with the exception of the age data, were initially assigned a lambda weight equal to 1.0. Based on a recommendation from the model developer, the likelihood components for the age data were reduced to 0.25 (R.D. Methot Jr., NOAA Fisheries, personal communication).

The model results are dependent, sometimes highly, on the weighting of each data set (Francis 2011). Francis (2011) points out that there is wide agreement on the importance of weighting, but there is lack of consensus as to how it should be addressed. In integrated models that use multiple data sets, it is not uncommon for the composition data to drive the estimation of absolute abundance when inappropriate data weightings are applied or the selectivity process is misspecified (Lee et al. 2014). Francis (2011) argues that abundance information should primarily come from indices of abundance and not from composition data. Following the recommendation of Francis (2011), the model was weighted in two stages. Stage 1 weights were largely empirically derived (standard errors, CVs, and effective sample sizes described earlier in this section) and applied to individual data observations. Stage 2 weights were applied to reweight the length and age composition data by adjusting the input effective sample sizes. The stage 2 weights were estimated based on method TA1.8 (Appendix A in Francis 2011) using the SSMethod.TA1.8 function within the r4ss package (Taylor et al. 2019) in R (R Core Team 2019).

### **3.1.6 Diagnostics**

Several approaches were used to assess model convergence. The first diagnostic was to check whether the Hessian matrix (i.e., matrix of second derivatives of the likelihood with respect to the parameters) inverted. Next, the model convergence level was compared to the convergence criteria (0.0001, common default value). Ideally, the model convergence level will be less than the criteria.

Model stability was further evaluated using a “jitter” analysis. This analysis is a built-in feature of SS in which the initial parameter values are varied by a user-specified fraction. This allows evaluation of varying input parameter values on model results to ensure the model has converged on a global solution. A model that is well behaved should converge on a global solution across a reasonable range of initial parameter estimates (Cass-Calay et al. 2014). Initial parameters were randomly jittered by 10% for a series of 50 random trials. The final model total likelihood value, annual estimates of spawning stock biomass (SSB), annual  $F$  values, and associated thresholds (see section 4) from the jitter runs were compared to the base run results.

Additional diagnostics included evaluation of fits to landings and harvest, discards, indices, and length compositions and comparison of predicted growth parameters to empirical values. The evaluation of fits to the various data components included a visual comparison of observed and predicted values and calculation of standardized residuals for the fits to the fisheries-independent survey indices and length composition data. The standardized residuals were first visually inspected to evaluate whether any obvious patterns were present. In a model that is fit well, there should be no apparent pattern in the standardized residuals. If most of the residuals are within one standard deviation of the observed value, there is evidence of under-dispersion. This is indicative of a good predictive model for the data. That is, the model is fitting the data much better than expected, given the assumed sample size.

Checking for patterns in standardized residuals over time can be done via the runs test, which was applied to the standardized residuals of the fits to the fisheries-independent survey indices. The runs test was applied using the RunsTest function in the DescTools package (Signorell et al. 2019) in R (R Core Team 2019). In a perfectly fit model, the standardized residuals have a normal distribution with mean equal to 0 and standard deviation equal to 1. The Shapiro-Wilk distribution test was applied to determine whether the standardized residuals of the fits to the fisheries-independent survey indices were normally distributed. This test was conducted using the shapiro.test function within the stats package in R (R Core Team 2019). An alpha level of 0.05 was used for both the runs test and Shapiro-Wilk distribution test to determine significance.

### **3.1.7 Uncertainty & Sensitivity Analyses**

#### **3.1.7.1 Evaluate Data Sources**

Uncertainty can also be explored by assessing the contribution of each source of information (Methot 1990). The contribution of a data source or other parameter(s) can be manipulated by changing the weight, or emphasis, of the associated likelihood component.

The contribution of different fisheries-independent surveys was explored by removing the data from each survey one at a time in a series of model runs. In each of these runs, the survey under evaluation was effectively removed by assigning a lambda weight of 0.0 to the likelihood component for that survey’s index and associated biological data (if present).

Annual estimates of female spawning stock biomass and  $F$  were compared to those from the base run.

#### **3.1.7.2 Alternative Natural Mortality**

Natural mortality was assumed to be constant across sexes and ages in the final base run ( $M=0.40$ ; section 3.1.4.5); however, natural mortality that varies by sex and age may be more realistic. In one sensitivity run, natural mortality was assumed equal to the values derived using the modified Lorenzen approach described in section 1.2.6.1 (assumed sex-specific and age-variable).

Additionally, a run was performed in which natural mortality was assumed equal to the empirical estimate of 0.72 derived from the Harris and Hightower (2017) study (assumed sex- and age-constant). Finally, a run was performed in which natural mortality was assumed equal to 0.30 to provide a run that used a lower range value for natural mortality (assumed sex- and age-constant).

### 3.1.8 Results

A summary of the input data used in the base run of the striped bass stock assessment model is shown in Figure 3.4.

#### 3.1.8.1 Base Run—Diagnostics

The final base run resulted in an inverted Hessian matrix, but the model's final convergence level was 0.00673183. This value is higher than the convergence criteria, which was set at 0.0001. It is not unusual for models with hundreds of parameters to produce higher convergence levels and so values less than 1.0 for such models are typically deemed acceptable (R.D. Methot Jr., NOAA Fisheries, personal communication). Four out of 111 estimated parameters were estimated near their bounds (Table 3.4). These are the CV for female age at  $L_\infty$ , CV for male age at  $L_\infty$ , initial equilibrium  $F$  for the RRrec discard fleet, and one of the selectivity parameters for the ARcomm fleet.

Twenty one of the 50 jitter runs successfully converged (Table 3.5). None of the converged jitter runs resulted in a likelihood value that was lower than the base run (Figure 3.5). The majority of the converged runs produced similar trends in female SSB and  $F$  to the base run (Figure 3.6). The results of one of the converged runs (run 46) was not included in these plots as it estimated female SSB to be an order of magnitude higher and  $F$  an order of magnitude lower than the other converged runs. Overall, the jitter analysis gives evidence that the base model converged to the global solution.

There is near identical agreement between observed and predicted landings and harvest for the ARcomm, ASrec, and RRrec fleets (Figure 3.7). This is not unexpected given the small amount of error assumed for these data (section 3.1.5). The SS model tended to underestimate discards for the ARcomm fleet (Figure 3.8A). For the ASrec discards, the model overestimated in some years and underestimated in others (Figure 3.8B). The RRrec discards were fit well by the model (Figure 3.8C).

Model fits to the fisheries-independent survey indices are reasonable (Figures 3.9–3.12). The model-predicted indices tended to capture the overall trend in the observed values for the P100juv (Figure 3.9), P135fw (Figure 3.10), and RRef (Figure 3.12) survey indices but did a poor job of predicting the trend for the P135spr survey index (Figure 3.11). The model did not capture the same degree of inter-annual variability seen in the observed index. Visual inspection of the standardized residuals indicates no clear temporal patterns for any of the survey indices and this was confirmed by the results of the runs tests, which produced non-significant ( $\alpha=0.05$ )  $P$ -values (Table 3.6). None of the standardized residuals for the fisheries-independent survey indices were found to be significantly different from a normal distribution based on the results of the Shapiro-Wilk test for normality.

The fits to the length compositions aggregated across time appear reasonable for most of the fleets and surveys with the exception of the fit to the ARcomm discard lengths (Figure 3.13). This poor fit is likely due, in part, to the small effective sample sizes associated with the ARcomm discard length compositions. Examination of the fits to the length composition data by individual year

indicates fits ranging from good to poor (Figures 3.14–3.28). Again, the poor fit to the ARcomm discard lengths is evident (Figure 3.16). The presence of bimodality in the P135fw survey lengths provided some difficulty in model fitting (Figures 3.23, 3.24). This was also true for the P135spr survey lengths (Figures 3.25, 3.26). Residuals from the fits to the length composition data for the different data sources are shown in Figures 3.29–3.37. The fits to the length composition data from the P135fw survey (Figures 3.35), P135spr survey (Figure 3.36), and RRef survey (Figure 3.37) show residual patterns which suggest the periodic presence of strong year classes. The strongest length composition residual patterns are evident in the ASrec harvest (Figure 3.31) and ASrec discard (Figure 3.32) fits. Fits to the ASrec harvest lengths suggest underestimation at mid-range lengths and overestimation at the smallest and largest lengths (Figure 3.31). The opposite pattern is seen in the fits to the ASrec discard lengths, which shows overestimation at mid-range lengths and underestimation at the smallest and largest lengths (Figure 3.32).

The growth curves estimated by the model are similar to the curves derived empirically (Figure 3.38). The predicted growth curves for both females and males suggest a small degree of underestimation of length at age.

### **3.1.8.2 Base Run—Selectivity & Population Estimates**

The predicted selectivity curves are shown in Figures 3.39–3.41 and are considered reasonable.

Annual predicted recruitment is variable among years and demonstrates a general decrease over the time series (Table 3.7; Figure 3.42). Predicted recruitment deviations are shown in Figure 3.43 and show no obvious concerning pattern.

There is less inter-annual variability in predicted female spawning stock biomass (SSB; Table 3.7; Figure 3.44) than that exhibited in the predicted recruitment values (Figure 3.42). Female SSB values were highest in the late 1990s through the mid-2000s and have generally decreased since. The predicted stock-recruitment relationship indicates the relation is not particularly strong (Figure 3.45). This is not unexpected given the model assumed a fixed value of 0.9 for the steepness parameter. Predicted values of spawner potential ratio (SPR) show a slightly decreasing trend over the time series (Table 3.7; Figure 3.46).

Predicted population numbers at age suggest 60–65% of the population has been dominated by age-0 and age-1 fish (Tables 3.8–3.9). These predicted numbers at age show an increase in the numbers of older fish through the mid-2000s, followed by a possible truncation of age structure in recent years. The predictions of landings at age for the ARcomm fleet indicate that most (~82%) of the fish captured are ages 3 through 5 (Table 3.10). The majority (84%) of the discards for the ARcomm fleet are ages 2 through 5 (Table 3.11). The harvest for the ASrec fleet is dominated (nearly 81%) by ages 3 through 6 (Table 3.12). Approximately 74% of the discards for the ASrec fleet are ages 3 and 4 (Table 3.13). The RRrec fleet captures mostly (93%) age-3 to age-5 striped bass in the harvest (Table 3.14) while most (67%) of the RRrec discards are age 3 and 4 (Table 3.15).

Model predictions of annual  $F$  (numbers-weighted, ages 3–5) exhibit moderate inter-annual variability throughout the assessment time series and peaks are observed in 2012 and 2016 (Table 3.16; Figure 3.47). Predicted  $F$  values range from a low of 0.15 in 1997, 1999, and 2003 to a high of 1.3 in 2012. There a decline in  $F$  in the last year of the time series.

### 3.1.8.3 Evaluate Data Sources

The removal of the different survey data sets had minimal impact on estimates of female SSB and  $F$  (Figure 3.48).

### 3.1.8.4 Alternative Natural Mortality

Assuming age-varying natural mortality (Lorenzen  $M$ ) and a lower value of natural mortality ( $M=0.30$ ) produced estimates of female SSB that were lower than those in the base run while the overall trends were similar (Figure 3.49A). Using the higher empirically-derived value of natural mortality ( $M=0.72$ ) resulted in higher estimates of female SSB than those predicted in the base run. The model that assumed the empirical estimate of natural mortality resulted in lower estimates of  $F$  relative to the base run as did the run that assumed natural mortality varied with age and sex (Figure 3.49B). Predicted  $F$  values were slightly higher when the lower value of natural mortality was assumed ( $M=0.30$ ). Estimates of recruitment increased by an order of magnitude when using the empirically-derived natural mortality and when using the Lorenzen natural mortality (Figure 3.50).

## 3.2 Discussion of Results

The current stock assessment for striped bass indicates some concerning trends. Observed recruitment in recent years of the assessment time series (Figures 2.22, 3.3A) has been relatively low and predicted recruitment has been showing a general decline recently (Figure 3.42). Overall, recruitment is highly variable and has been generally lower in recent years relative to that observed and predicted from 1991 through 2000. From 1993 through 2000, the stock produced seven of the top nine year classes in terms of age-0 abundance. The 2000 cohort is the largest produced in the entire time series. Since then, from 2001 through 2006, five out of the six cohorts produced were below-average in terms of numbers and only the 2005-year class is considered a strong year class (Table 3.7; Figure 3.42). These observations suggest there is another factor besides simply the size of SSB that has an influence on producing strong year classes. Much research from the 1950s through the 1980s supports the importance of flow in the Roanoke River during the spawning period and subsequent weeks while eggs and larvae are being transported down the Roanoke River to the nursery habitat in the western Albemarle Sound and the importance of flow in supporting abundant striped bass year-class production (Hassler et al. 1981; Rulifson and Manooch 1990; Zincone and Rulifson 1991).

The length (Figures 2.2, 2.3) and age (Figures 2.4, 2.5) compositions of striped bass sampled from the commercial landings show that fewer larger and older fish have been observed in recent years. A truncation of the length (Figure 2.32) and age (Figure 2.33) structure is also evident in the observations from the Roanoke River Electrofishing Survey. Recent observations from the Roanoke River Electrofishing Survey of abundance are the lowest in the time series (Figure 2.31). The abundance of age 9+ fish in the survey has also been declining in recent years. Predicted population numbers at age show a truncation in the most recent years of the time series and an overall decline in total population abundance (Tables 3.8, 3.9). Predicted female SSB (Figure 3.44) has also shown a declining trend in recent years and, estimates in recent years have been the lowest in the entire time series. The 2016 estimate of fishing mortality was the second highest in the time series and declined in 2017 (Figure 3.47).

Performance of the stock assessment model was considered good in terms of predicting the observed data. The quality of the fits is strongly tied to the input variance and effective sample sizes. Fits to the observed landings, harvest, and discard were reasonable and this was expected

given the low variance assumed for these data sources. Of the fisheries-independent survey indices, all but the P135spr index were fit well and no issues were detected among the residuals for any of the survey indices. The model was insensitive to the removal of the various sources of fisheries-independent survey data suggesting the different surveys share similar signals in the data with regard to population trends.

Striped bass commonly migrate outside the bounds of the A-R management unit, either to other internal waters of North Carolina such as western Pamlico Sound and the Tar-Pamlico, Pungo, and Neuse rivers or by joining the migratory ocean stock. The probability of migration increases with age and has increased over time (Callihan et al. 2014). In the most recent years examined in Callihan et al. (2014), the probability has been most significant for fish age 6 and older (20% or greater). In addition, smaller adults show evidence of density-dependent movements and habitat utilization, as the likelihood of recapture outside the ASMA in adjacent systems increases during periods of higher stock abundance. When a striped bass migrates, it may not return to its natal waterbody; this could be due to harvest outside of the ASMA and RRMA and is not accounted for in the harvest losses here. This loss of fish from the system will likely be interpreted by the model as losses due to natural and/or fishing mortality. The most recent assessments of the A-R striped bass stocks attempted to account for these migration losses by adjusting the natural mortality rate by the probability of migration and fishing mortality occurring in the Atlantic Ocean, thereby creating an estimate of total unobserved mortality that accounted for both natural mortality and losses not attributable to North Carolina fisheries (Mroch and Godwin 2014; Flowers et al. 2016). In this assessment, migration losses were not specifically modeled; this total unobserved mortality was treated as fixed in the modeling process.

The ages in this assessment were derived from scales and were assumed to be associated with small bias and negligible imprecision; however, Welch et al. (1993) found that scales tend to underage striped bass for fish that are older than age ten. This suggests that the maximum age assumed for this assessment, age 17, may be an underestimate of the true maximum age. Assuming maximum age that is too young can positively bias the estimates of SPR (Goodyear 1993) and the derived reference points.

There is additional recent evidence that age 17 may not be the maximum age for the A-R stock. In 2017, an angler returned a striped bass tag from a fish that had been tagged on the spawning grounds in 2007, which was aged at the time to 13 years old, increasing the oldest known age fish in the A-R stock to 23. In April 2020, an angler caught and cut the tag off a striped bass in the Roanoke River that was originally tagged in 1995 and estimated to be age 6, which suggests the oldest known fish in the stock is now at 31 years old, likely from the 1989 year class. Note that these instances are of single tag returns and it is not known how reflective they are of the relative abundance of these older fish in the stock. The available observed data suggested few fish older than age 9 are present in the stock, especially in recent years.

#### **4 STATUS DETERMINATION CRITERIA**

The General Statutes of North Carolina define overfished as “the condition of a fishery that occurs when the spawning stock biomass of the fishery is below the level that is adequate for the recruitment class of a fishery to replace the spawning class of the fishery” (NCGS § 113-129). The General Statutes define overfishing as “fishing that causes a level of mortality that prevents a fishery from producing a sustainable harvest.”

The working group decided that the spawner potential ratio (SPR) was an appropriate proxy for developing reference points. Levels of SPR ranging from 20% to 50% have been found to be appropriate for various stocks, but historical analysis of SPR shows increased risk of recruitment overfishing levels if SPR falls below 30% (Walters and Martell 2004). For this assessment, threshold values were based on 35% SPR and targets were based on 45% SPR.

The fishing mortality reference points and the values of  $F$  that are compared to them represent numbers-weighted values for ages 3 to 5 (section 3.1.4.7). The SS model estimated a value of 0.13 for  $F_{\text{Target}}$  ( $F_{45\%}$ ). The estimate of  $F_{\text{Threshold}}$  ( $F_{35\%}$ ) from the SS model was 0.18. The estimated value of fishing mortality in the terminal year (2017) of the model was 0.27, which is greater than the threshold value and suggests that overfishing is currently occurring in the stock ( $F_{2017} > F_{\text{Threshold}}$ ; Figure 4.1).

The target level for female spawning stock biomass ( $SSB_{\text{Target}}$  or  $SSB_{45\%}$ ) was estimated at 159 metric tons by the SS model. The estimated threshold for SSB ( $SSB_{\text{Threshold}}$  or  $SSB_{35\%}$ ) was 121 metric tons. Terminal year (2017) female SSB was 35.6 metric tons, which is less than the threshold value and suggests the stock is currently overfished ( $SSB_{2017} < SSB_{\text{Threshold}}$ ; Figure 4.2).

The estimates in the most recent years are often associated with large uncertainty in stock assessment models. Approaching the ending year of the time series, the estimates of the most recent years lack data support from subsequent years during calibration. Nevertheless, stock status is often based on the terminal year estimates of fishing mortality and population size (or a proxy) to address the management needs and interests.

## 5 SUITABILITY FOR MANAGEMENT

Stocks assessments performed by the NCDMF in support of management plans are subject to an extensive review process, including a review by an external panel of experts. External reviews are designed to provide an independent peer review and are conducted by experts in stock assessment science and experts in the biology and ecology of the species. The goal of the external review is to ensure the results are based on the best science available and provide a valid basis for management.

The review workshop allows for discussion between the working group and review panel, enabling the reviewers to ask for and receive timely updates to the models as they evaluate the sensitivity of the results to different model assumptions. The workshop also allows the public to observe the peer review process and better understand the development of stock assessments.

The external peer review panel first met with the working group in person in December 2019. The reviewers were concerned with the external fit of the von Bertalanffy growth model to the observed age-length data; model predicted size was consistently smaller than empirical size for larger, older fish. The reviewers were also concerned with residual patterns in the fits to the length composition data indicative of model misspecification. Another major concern was failure of the model to capture trends observed in the empirical data. The peer reviewers did not support the presented model for management use but agreed to a second review after the working group addressed their concerns. In preparing the updated model, the working group noted an error in the input data that invalidated the first model. The working group corrected the data issue and also addressed the peer reviewer concerns regarding model fitting. A second assessment was presented to the peer review panel via webinar in June 2020.

The external peer reviewers worked with the working group to develop a model (presented in section 3) that the peer review endorsed for management use for at least the next five years and



agreed the determination of stock status (overfished and overfishing) for the North Carolina Albemarle Sound-Roanoke River striped bass in the terminal year concurs with professional opinion and observations. The reviewers also agreed that: (1) the justification of inclusion and exclusion of data sources are appropriate; (2) the data sources used in this assessment are appropriate; (3) determination of stock status for the terminal year is robust to model assumptions on natural mortality and growth; (4) the extensive exploration of sensitivities to model assumptions and configurations, especially the sensitivity analysis regarding the natural mortality and growth assumptions, resolves the reviewers' primary areas of concerns such as the concerns over the fitting to growth data and length composition data and the concern regarding the overestimation of abundance for the last three years of the time series; (5) reviewers recommend future assessments consider key abiotic drivers of poor recruitment such as river flow and key biotic drivers such as catfish predation and competition; (6) reviewers also recommend collection of sex-specific growth data from juveniles and old fish to better inform growth estimates and length- or age-specific natural mortality estimates, and to resolve the concern on growth estimates showing little difference between males and females. Detailed comments from the external peer reviewers are provided in the Appendix.

While the peer reviewers did approve the model for management use and were confident in the declining trend in recruitment based on assessment results and results from the Juvenile Abundance Survey (P100; Figure 5.1), there was a great deal of uncertainty in the potential causes of the decline in recruitment (Appendix). One key uncertainty was related to the impacts of changes in river flow on YOY abundance. The review panel recognized the declining recruitment in the time series did not appear to result solely from reduced stock abundance due to harvest (i.e., overfishing). The review panel suggested future assessments consider formally incorporating the flow-recruitment relationship into the stock assessment as spring flow conditions are believed to influence recruitment and ultimately stock abundance. Another area of potential influence on the striped bass stock is the prevalence of the non-native blue catfish (*Ictalurus furcatus*). The population of blue catfish in the Roanoke River and western Albemarle Sound and tributaries has increased dramatically in recent years (Darsee et al. 2019; NCDMF 2019). The reviewers felt predation by blue catfishes could potentially impact recruitment of striped bass directly or could influence food resources for striped bass through competition for prey (e.g., Pine et al. 2005). The review panel recognized the degree to which this occurs is not known, but future assessments should consider this as a factor that may influence abundance but is not tied to striped bass harvest.

## **6 RESEARCH RECOMMENDATIONS**

The research recommendations listed below are offered by the working group to improve future stock assessments of the A-R striped bass stock.

### High

- Improve estimates of discard mortality rates and discard losses from the ASMA commercial gill-net fisheries (ongoing through observer program)
- Collect data to estimate catch-and-release discard losses in the ASMA recreational fishery during the closed harvest season
- Investigate relationship between river flow and striped bass recruitment for consideration of input into future stock assessment models

### Medium

- Transition to an assessment that is based on ages derived from otoliths
- Improve estimates of catch-and-release discard losses in the RRMA recreational fishery during the closed harvest season
- Incorporate tagging data directly into the statistical catch-at-age model
- Improve the collection of length and age data to characterize commercial and recreational discards
- Explore the direct input of empirical weight-at-age data into the stock assessment model in lieu of depending on the estimated growth relationships

### Low

- Re-evaluate catch-and-release mortality rates from the ASMA and RRMA recreational fisheries incorporating different hook types and angling methods at various water temperatures (e.g., live bait, artificial bait, and fly fishing)
- Investigate the potential impact of blue catfish on the A-R striped bass population (e.g., habitat, predation, forage)

## 7 LITERATURE CITED

- Alverson, D.L., and M.J. Carney. 1975. A graphic review of the growth and decay of population cohorts. *Journal du Conseil international pour l'Exploration de la Mer* 36(2):133–143.
- Artedi, P., C.V. Linnaeus, and J.J. Walbaum. 1792. *Petri Artedi Sueci Genera piscium : in quibus systema totum ichthyologiae proponitur cum classibus, ordinibus, generum characteribus, specierum differentiis, observationibus plurimis : redactis speciebus 242 ad genera 52: Ichthyologiae*. Available (July 2020): <https://www.biodiversitylibrary.org/bibliography/61537#/summary>
- Atlantic States Marine Fisheries Commission (ASMFC). 1998. Source document to amendment 5 to the interstate fisheries management plan for Atlantic Striped Bass. ASMFC, Fisheries Management Report No. 34, Arlington, Virginia. 117 p.
- ASMFC. 2003. Amendment 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 41, Washington, DC.
- ASMFC. 2007. Addendum 1 to Amendment 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 16, Washington, DC.
- ASMFC. 2009. 2009 stock assessment report for Atlantic striped bass. A Report prepared by the Atlantic Striped Bass Technical Committee. Accepted for management use November 2009.
- Benton, J.C. 1992. Atlantic migratory striped bass adult monitoring program - North Carolina and Virginia offshore mixed stocks, 1988–1992. U.S. Fish and Wildlife Service, South Atlantic Fisheries Coordination Office, Morehead City, North Carolina.
- Berst, A.H. 1961. Selectivity and efficiency of experimental gill nets in South Bay and Georgian Bay of Lake Huron. *Transactions of the American Fisheries Society* 90(4):413–418.
- Bigelow, H.B., and W.C. Schroeder. 1953. *Fishes of the Gulf of Maine*. U.S. Fish and Wildlife Service Fisheries Bulletin 53.
- Boreman, J., and R.R. Lewis. 1987. Atlantic coastal migration of striped bass. *American Fisheries Society, Symposium* 1, Bethesda, Maryland.
- Boyd, J.B. 2011. Maturation, fecundity, and spawning frequency of the Albemarle/Roanoke striped bass stock. Master's thesis. East Carolina University, Greenville, North Carolina. 132 p.
- Buckler, D.R., P.M. Mehrle, L. Cleveland, and F.J. Dwyer. 1987. Influence of pH on the toxicity of aluminium and other inorganic contaminants to East Coast striped bass. *Water, Air, and Soil Pollution* 35:97–106. <https://doi.org/10.1007/BF00183846>
- Buckmeier, D.L., and J.W. Schlechte. 2009. Capture efficiency and size selectivity of channel catfish and blue catfish sampling gears. *North American Journal of Fisheries Management* 29(2):404–416.
- Cubillos, L.A. 2003. An approach to estimate the natural mortality rate in fish stocks. *Naga, Worldfish Center Quarterly* 26(1):17–19.

- Callihan, J.L., C.H. Godwin, and J.A. Buckel. 2014. Effect of demography on spatial distribution: movement patterns of Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*) in relation to their stock recovery. *Fisheries Bulletin* 112(2-3):131–143.
- Carmichael, J.T. 1998. Status of the Albemarle Sound-Roanoke River stock of striped bass, 1982–1997. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Carmichael, J.T. 1999. Status of the Albemarle Sound-Roanoke River stock of striped bass, 1982–1998. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Carmichael, J.T. 2000. Status of the Albemarle Sound-Roanoke River stock of striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Carmichael, J.T. 2001. Status of the Albemarle Sound-Roanoke River stock of striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Carmichael, J.T. 2002. Status of the Albemarle Sound-Roanoke River stock of striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Carmichael, J.T. 2003. Status of the Albemarle Sound-Roanoke River stock of striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Cass-Calay, S.L., J.C. Tetzlaff, N.J. Cummings, and J.J. Isely. 2014. Model diagnostics for Stock Synthesis 3: examples from the 2012 assessment of cobia in the U.S. Gulf of Mexico. *Collective Volume of Scientific Papers ICCAT* 70(5):2069–2081.
- Chapoton, R.B., and J.E. Sykes. 1961. Atlantic Coast migration of large striped bass as evidenced by fisheries and tagging. *Transactions of the American Fisheries Society* 90(1):13–20.
- Cooper, J.E., R.A. Rulifson, J.J. Isely, and S.E. Winslow. 1998. Food habits and growth of juvenile striped bass, *Morone saxatilis*, in Albemarle Sound, North Carolina. *Estuaries* 21(2):307–317.
- Darsee, S.P., T. Mathes and J. Facendola 2019. North Carolina Striped Bass monitoring. Federal Aid in Sport Fish Restoration, Project F-56 Segment 26, Independent Gill-Net Survey 2019 Technical Report. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 61 p.
- Daugherty, D.J., and T.M. Sutton. 2005. Use of a chase boat for increasing electrofishing efficiency for flathead catfish in lotic systems. *North American Journal of Fisheries Management* 25(4):1528–1532.
- Davis, W.S., and J.E. Sykes. 1960. Commercial harvest and catch composition of striped bass in Albemarle Sound, North Carolina. National Marine Fisheries Service, Atlantic Estuarine Fisheries Center, Beaufort, North Carolina. 44 p.
- Deriso, R.B., T.J. Quinn II, and P.R. Neal. 1985. Catch at age analysis with auxiliary information. *Canadian Journal of Fisheries and Aquatic Sciences* 42:815–824.
- Dorazio, R.M. 1995. Mortality estimates of striped bass caught in the Albemarle Sound and Roanoke River, North Carolina. *North American Journal of Fisheries Management* 15(2): 290–299.
- Diodati, P.J. 1991. Estimating mortality of hook and released striped bass. Project AFC-22, Final Report. Massachusetts Division of Marine Fisheries, Salem.

- Dolan, C.R., and L.E. Miranda. 2003. Immobilization thresholds of electrofishing relative to fish size. *Transactions of the American Fisheries Society* 132(5):969–976.
- Flowers, J., S. Darsee, L. Lee, and C. Godwin. 2016. Stock status of Albemarle Sound-Roanoke River striped bass: update 1982–2014. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2016-01, Morehead City, NC. 87 p.
- Francis, R.I.C.C. 2011. Data weighting in statistical fisheries stock assessment models. *Canadian Journal of Fisheries and Aquatic Sciences* 68(6):1124–1138.
- Gibson, M.R. 1995. Status of the Albemarle Sound-Roanoke River striped bass stock in 1994. Rhode Island Division of Fish and Wildlife, Wickford, Rhode Island. 14 p.
- Goodyear, C.P. 1993. Spawning stock biomass per recruit in fisheries management: foundation and current use. Pages 67–81 *In*: S.J. Smith, J.J. Hunt, D. Rivard (editors), Risk evaluation and biological reference points for fisheries management. Canadian Special Publication of Fisheries and Aquatic Sciences 120.
- Grant, G.C. 1974. The age composition of striped bass catches in Virginia rivers, 1967–1971, and a description of the fishery. *Fisheries Bulletin* 72(1):193–199.
- Greene, K.E., J.L. Zimmerman, R.W. Laney, and J.C. Thomas-Blate. 2009. Atlantic coast diadromous fish habitat: a review of utilization, threats, recommendations for conservation, and research needs. Atlantic States Marine Fisheries Commission, Habitat Management Series No. 9, Washington D.C. 464 p.
- Grist, J. 2004. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Grist, J. 2005. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Hall, N.G. 2013. Report on the SEDAR 28 desk review of the stock assessments for Gulf of Mexico cobia and Spanish mackerel. 66 p. Available (November 2019): [https://www.st.nmfs.noaa.gov/Assets/Quality-Assurance/documents/peer-review-reports/2013/2013\\_02\\_19%20Hall%20SEDAR%2028%20GM%20spanish%20mackerel%20cobia%20assessment%20report%20review%20report.pdf](https://www.st.nmfs.noaa.gov/Assets/Quality-Assurance/documents/peer-review-reports/2013/2013_02_19%20Hall%20SEDAR%2028%20GM%20spanish%20mackerel%20cobia%20assessment%20report%20review%20report.pdf)
- Hall, L.W., S.E. Finger, and M.C. Ziegenfuss. 1993. A review of in situ and on-site striped bass contaminant and water-quality studies in Maryland waters of the Chesapeake Bay watershed. Pages 3–15 *In*: L.A. Fuiman (editor), Water quality and the early life stages of fishes. American Fisheries Society, Symposium 14, Bethesda, Maryland.
- Hansson, S., and L.G. Rudstam. 1995. Gillnet catches as an estimate of fish abundance: a comparison between vertical gillnet catches and hydroacoustic abundance of Baltic Sea herring (*Clupea harengus*) and sprat (*Sprattus sprattus*). *Canadian Journal of Fisheries and Aquatic Sciences* 52(1):75–83.
- Harrell, R.M. 1988. Catch and release mortality of striped bass caught with artificial lures and baits. *Proceedings of the Annual Conference Southeastern Association of Fish and Wildlife Agencies* 41(1987):70–75.
- Harris Jr., R.C., and B.L. Burns. 1983. An investigation of size, age, and sex of North Carolina striped bass. Project AFC-18-2, Annual Progress Report. North Carolina Department of

- Natural Resources and Community Development, Division of Marine Fisheries, Morehead City, North Carolina.
- Harris Jr., R.C., B.L. Burns, and H.B. Johnson. 1985. An investigation of size, age, and sex of North Carolina striped bass. Project AFC-18, Completion Report. North Carolina Department of Natural Resources and Community Development, Division of Marine Fisheries, Morehead City, North Carolina. 136 p.
- Harris, J.E., and J.E. Hightower. 2017. An integrated tagging model to estimate mortality rates of Albemarle Sound-Roanoke River striped bass. *Canadian Journal of Fisheries and Aquatic Sciences* 74(7):1061–1076.
- Hassler, W.W., N.L. Hill, and J.T. Brown 1981. The status and abundance of striped bass, *Morone saxatilis*, in the Roanoke River and Albemarle Sound, North Carolina, 1956–1980. Report to the North Carolina Department of Natural Resources and Community Development, Division of Marine Fisheries. Special Scientific Report 38.
- Hassler, W.W., and S.D. Taylor. 1984. The status, abundance, and exploitation of striped bass in the Roanoke River and Albemarle Sound, North Carolina, 1982, 1983. Project AFC-19, Completion Report. North Carolina Division of Marine Fisheries. NCDMF Publication No. 136.
- Henry, L.T., S.D. Taylor, and S.E. Winslow. 1992. North Carolina striped bass. Project AFS-26, Completion Report. North Carolina Department of Environment, Health, and Natural Resources, Division of Marine Fisheries. Morehead City, North Carolina.
- Hewitt, D.A., and J.M. Hoenig. 2005. Comparison of two approaches for estimating natural mortality based on longevity. *Fishery Bulletin* 103(2):433–437.
- Hightower, J.E., A.M. Wicker, and K.M. Endres. 1996. Historical trends in abundance of American shad and river herring in Albemarle Sound, North Carolina. *North American Journal of Fisheries Management* 16(2):257–271.
- Hill, J., J.W. Evans, and M.J. Van Den Avyle. 1989. Species profiles: life histories and environmental requirements of coastal fishes and invertebrates (South Atlantic)—striped bass. Biological Report 82(11.118), U.S. Department of the Interior, Fish and Wildlife Service, Washington, D.C. 35 p.
- Hoenig, J.M. 1983. Empirical use of longevity data to estimate mortality rates. *Fishery Bulletin* 82(1):898–903.
- Holland, B.F., and G.F. Yelverton. 1973. Distribution and biological studies of anadromous fishes offshore North Carolina. North Carolina Department of Natural and Economic Resources, Division of Commercial and Sport Fisheries. Morehead City, North Carolina. 156 p.
- Humphries, M., and J.W. Kornegay. 1985. An evaluation of the use of bony structures for aging Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*). Federal Aid in Sport Fish Restoration, Project F-22. North Carolina Wildlife Resources Commission, Raleigh.
- Hutchinson, T. [1764] 1936. The history of the colony and province of Massachusetts-Bay, Volume I, with a memoir and additional notes by L. S. Mayo. Harvard University Press, Cambridge, MA. 467 p.

- Jensen, A.L. 1996. Beverton and Holt life history invariants result from optimal trade-off of reproduction and survival. *Canadian Journal of Fisheries and Aquatic Sciences* 53(4):820–822.
- Lee, H-H., K.R. Piner, R.D. Methot Jr., and M.N. Maunder. 2014. Use of likelihood profiling over a global scaling parameter to structure the population dynamics model: an example using blue marlin in the Pacific Ocean. *Fisheries Research* 158:138–146.
- Lee, L.M., and J.E. Rock. 2018. The forgotten need for spatial persistence in catch data from fixed station surveys. *Fishery Bulletin* 116(1):69–74.
- Lewis, R.M., and R.R. Bonner, Jr. 1966. Fecundity of the striped bass, *Roccus saxatilis* (Walbaum). *Transactions of the American Fisheries Society* 95(3):328–331.
- Limburg, K.E., and J.R. Waldman. 2009. Dramatic declines in north Atlantic diadromous fishes. *BioScience* 59(11):955–965.
- Lorenzen, K. 1996. The relationship between body weight and natural mortality in juvenile and adult fish: a comparison of natural ecosystems and aquaculture. *Journal of Fish Biology* 49(4):627–647.
- Lorenzen, K. 2005. Population dynamics and potential of fisheries stock enhancement: practical theory for assessment and policy analysis. *Philosophical Transactions of the Royal Society of London, Series B* 360(1453):171–189.
- Lupton, B.Y., and P.S. Phalen. 1996. Designing and implementing a trip ticket program. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 32 p + appendices.
- Manooch, C. 1973. Food habits of yearling and adult striped bass, *Morone saxatilis* (Walbaum) from Albemarle Sound, North Carolina. *Chesapeake Science* 14(2):73–86.
- McFarland, R. 1911. History of New England fisheries. University of Pennsylvania Press, Philadelphia, PA. 455 p.
- Maunder, M.N., and A.E. Punt. 2004. Standardizing catch and effort data: a review of recent approaches. *Fisheries Research* 70(2-3):141–159.
- McInerney, M.C., and T.K. Cross. 2000. Effects of sampling time, intraspecific density, and environmental variables on electrofishing catch per effort of largemouth bass in Minnesota lakes. *North American Journal of Fisheries Management* 20(2):328–336.
- Merriman, D. 1941. Studies on the striped bass (*Roccus saxatilis*) of the Atlantic Coast. U.S. Fish and Wildlife Service Fisheries Bulletin 50(1):1–77.
- Methot, R.D. 1990. Synthesis model: an adaptable framework for analysis of diverse stock assessment data. *International North Pacific Fisheries Commission Bulletin* 50:259–277.
- Methot, R.D. 2000. Technical description of the stock synthesis assessment program. NOAA Technical Memorandum NMFS-NWFSC-43. 46 p.
- Methot Jr., R.D., and C.R. Wetzel. 2013. Stock synthesis: a biological and statistical framework for fish stock assessment and fishery management. *Fisheries Research* 142:86–99.
- Methot Jr., R.D., C.R. Wetzel, I.G. Taylor, and K. Doering. 2019. Stock synthesis user manual, version 3.30.14. NOAA Fisheries, Seattle, WA. 212 p.

- Moseley, A., W.B. Robertson, and M.G. Ellzey. 1877. Annual reports of the fish commissioners of the state of Virginia for the years 1875-6 and 1876-7, together with the laws relating to fish and game passed during the session of 1876-7. Printed by order of the Senate. R.F. Walker, Superintendent Public Printing, Richmond.
- Mroch, R., and C. Godwin. 2014. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 193 p.
- National Marine Fisheries Service (NMFS) and U.S. Fish and Wildlife Service (USFWS). 2016. Roanoke River diadromous fishes restoration plan. Raleigh, North Carolina. May 2016.
- Nichols, P.R., and R.P. Cheek. 1966. Tagging summary of American shad, *Alosa sapidissima* (Wilson) and striped bass, *Morone saxatilis* (Walbaum). Bureau of Commercial Fisheries, Biological Laboratory, Beaufort, North Carolina, 1950–1965. U.S. Fish and Wildlife Service, SSR No. 539. 8 p.
- North Carolina Department of Environmental Quality (NCDEQ). 2016. North Carolina coastal habitat protection plan. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 33 p.
- North Carolina Division of Marine Fisheries (NCDMF). 2004. North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environment and Natural Resources, Division of Marine Fisheries, Morehead City, North Carolina. 374 p.
- NCDMF. 2010. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 128 p.
- NCDMF. 2014. November 2014 Revision to amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environmental and Natural Resources, Division of Marine Fisheries, Elizabeth City, North Carolina. 15 p.
- NCDMF. 2019. 2019 License and Statistics annual report. North Carolina Department of Environmental Quality, Division of Marine Fisheries. Morehead City, North Carolina. 430 p.
- N.C. Striped Bass Study Management Board. 1991. Report on the Albemarle Sound-Roanoke River stock of striped bass. N.C. Striped Bass Study Management Board, U.S. Fish and Wildlife Service, Atlanta, Georgia. 56 p. + appendices.
- Ostrach, D.J., J.M. Low-Marchelli, K.J. Eder, S.J. Whiteman, and J.G. Zinkl. 2008. Maternal transfer of xenobiotics and effects on larval striped bass in the San Francisco Estuary. *Proceedings of the National Academy of Sciences of the United States of America* 105(49): 19354–19359.
- Pine III, W.E., T.J. Kwak, D.S. Waters, and J.A. Rice. 2005. Diet selectivity of introduced flathead catfish in coastal rivers. *Transactions of the American Fisheries Society* 134(4):901–909.
- Pollock, K.H., C.M. Jones, and T.L. Brown. 1994. Angler survey methods and their applications in fisheries management. *American Fisheries Society, Symposium* 25, Bethesda, Maryland.
- Ralston, S. 1987. Mortality rates of snappers and groupers. Pages 375–404 *In*: J.J. Polovina and S. Ralston (eds.), *Tropical Snappers and Groupers: Biology and Fisheries Management*. Westview Press, Boulder Colorado. 659 p.



- Reynolds, J.B. 1996. Electrofishing. Pages 221–253 *In*: B.R. Murphy and D.W. Willis (editors), Fisheries techniques, 2nd edition. American Fisheries Society, Bethesda, Maryland.
- Rudershausen, P.J., J.E. Tuomikoski, J.A. Buckel, and J.E. Hightower. 2005. Prey selectivity and diet of striped bass in western Albemarle Sound, North Carolina. *Transactions of the American Fisheries Society* 134(5):1059–1074.
- Ruetz III, C.R., D.G. Uzarski, D.M. Krueger, and E.S. Rutherford. 2007. Sampling a littoral fish assemblage: comparison of small-mesh fyke netting and boat electrofishing. *North American Journal of Fisheries Management* 27(3):825–831.
- Rulifson, R.A. 1990. Abundance and viability of striped bass eggs spawned in the Roanoke River, North Carolina, in 1989. North Carolina Department of Environmental Management, Health and Natural Resources and U.S. Environmental Protection Agency, Albemarle-Pamlico Estuarine Study, Raleigh, NC. Project No. 90-11. 96 p.
- Rulifson, R.A. 1991. Comparing the abundance and viability of striped bass eggs spawned in the Roanoke River, North Carolina, at two locations in 1991. Interim report to the North Carolina Striped Bass Study Management Board. Institute for Coastal and Marine Resources, and Department of Biology, East Carolina University, Greenville, NC.
- Rulifson, R.A., and D. Bass. 1991. Food analyses of young-of-year. Page 217-219 in NOAA Technical Memorandum NMFS-SEFC-291.
- Rulifson, R.A., M.T. Huish, and R.W. Thoesen. 1982. Anadromous fish in the Southeastern United States and recommendations for development of a management plan. U.S. Fish and Wildlife Service, Fisheries Resource, Region 4, Atlanta, Georgia. 525 p.
- Rulifson, R.A., and C.S. Manooch III. 1990. Recruitment of juvenile striped bass in the Roanoke River, North Carolina, as related to reservoir discharge. *North American Journal of Fisheries Management* 10(4):397–407.
- Schaaf, W. 1997. Status of the Albemarle Sound-Roanoke River striped bass stock in 1997. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Schoenebeck, C.W., and M.J. Hansen. 2005. Electrofishing catchability of walleyes, largemouth bass, smallmouth bass, northern pike, and muskellunge in Wisconsin lakes. *North American Journal of Fisheries Management* 25(4):1341–1352.
- Secor, D.H. 2000. Longevity and resilience of Chesapeake Bay striped bass. *ICES Journal of Marine Science* 57(4):808–815.
- Setzler, E.M., W.R. Boynton, K.V. Wood, H.H. Zion, L. Lubbers, N.K. Mountford, P. Frere, L. Tucker, and J.A. Mihursky. 1980. Synopsis of biological data on striped bass. NOAA Technical Report, NMFS Circular 443: FAO Synopsis No. 121. 69 p.
- Signorell, A. et mult. al. 2019. DescTools: tools for descriptive statistics. R package version 0.99.30.
- Smith, H.M. 1907. North Carolina geological and economic survey. Volume II. The fishes of North Carolina. E.M. Uzzell & Co., State Printers and Binders, Raleigh. 452 p.
- Smith, D. 1996. Annual survival of Albemarle Sound striped bass: a report to the ASMFC Striped Bass Stock Assessment Committee. ASMFC, Washington, D.C.

- Speas, D.W., C.J. Walters, D.L. Ward, and R.S. Rogers. 2004. Effects of intraspecific density and environmental variables on electrofishing catchability of brown and rainbow trout in the Colorado River. *North American Journal of Fisheries Management* 24(2):586–596.
- Stevens, D.E. 1966. Food habits of striped bass, *Morone saxatilis*, in the Sacramento-San Joaquin Delta. Pages 68–96 *In*: J.L. Turner and D.W. Kelley (compilers), *Ecological studies of the Sacramento-San Joaquin Delta. Part H. Fishes of the delta.* California Department Fish Game Fishery Bulletin 136.
- Street, M.W., A.S. Deaton, W.S. Chappell, and P.D. Mooreside. 2005. North Carolina Coastal Habitat Protection Plan. North Carolina Department of Environment and Natural Resources, Division of Marine Fisheries, Morehead City, North Carolina. 656 p.
- Street, M.W., and H.B. Johnson. 1977. Striped bass in North Carolina. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Street, M.W., P.P. Pate, Jr., B.F. Holland, Jr., and A.B. Powell. 1975. Anadromous fisheries research program, northern coastal region. Project AFCS-8, Completion Report. North Carolina Division of Marine Fisheries. 193 + 62 p and Append.
- Sullivan, C. 1956. The importance of size grouping in population estimates employing electric shockers. *Progress Fish-Culturist* 18(4):188–190.
- Sykes, J.E. 1957. A method of determining the sex of the striped bass, *Morone saxatilis* (Walbaum). *Transactions of the American Fisheries Society* 87(1):104–107.
- Takade, H.M. 2006. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Takade, H.M. 2010. Stock status of Albemarle Sound-Roanoke River striped bass. North Carolina Division of Marine Fisheries, Morehead City, North Carolina.
- Taylor, I.G., I.J. Stewart, A.C. Hicks, T.M. Garrison, A.E. Punt, J.R. Wallace, C.R. Wetzel, J.T. Thorson, Y. Takeuchi, K. Ono, C.C. Monnahan, C.C. Stawitz, Z.T. A'mar, A.R. Whitten, K.F. Johnson, R.L. Emmet, S.C. Anderson, G.I. Lambert, M.M. Stachura, A.B. Cooper, A. Stephens, N.L. Klaer, C.R. McGilliard, I. Mosqueira, W.M. Iwasaki, K. Doering, and A.M. Havron. 2019. r4ss: R code for stock synthesis. R package version 1.35.3. <https://github.com/r4ss>
- Taylor, M.J., and K.R. White. 1992. A meta-analysis of hooking mortality of nonanadromous trout. *North American Journal of Fisheries Management* 12(4):760–767.
- Then, A.Y., J.M. Hoenig, N.G. Hall, and D.A. Hewitt. 2015. Evaluating the predictive performance of empirical estimators of natural mortality rate using information on over 200 fish species. *ICES Journal of Marine Science* 72(1):82–92.
- Trent, L., and W.W. Hassler. 1966. Feeding behavior of adult striped bass, *Morone saxatilis*, in relation to stages of sexual maturity. *Chesapeake Science* 7(4):189–192.
- Trent, L., and W.W. Hassler. 1968. Gill net selection, migration, size and age composition, sex ratio, harvest efficiency, and management of striped bass in the Roanoke River, North Carolina. *Chesapeake Science* 9(4):217–232.

- Tuomikoski, J.E., P.J. Rudershausen, J.A. Buckel, and J.E. Hightower. 2008. Effects of age-1 striped bass predation on juvenile fish in western Albemarle Sound. *Transactions of the American Fisheries Society* 137(1):324–339.
- Vladykov, V.D., and D.H. Wallace. 1952. Studies of the striped bass, *Morone saxatilis* (Walbaum), with special reference to the Chesapeake Bay region during 1936–1938. *Bulletin of the Bingham Oceanographic Collection (Yale University)* 14(1):132–177.
- Walters, C.J., and S.J.D. Martell. 2004. *Fisheries ecology and management*. Princeton University Press, Princeton, New Jersey. 448 p.
- Warren, W.G. 1994. The potential of sampling with partial replacement for fisheries surveys. *ICES Journal of Marine Science* 51(3):315–324.
- Warren, W.G. 1995. Juvenile abundance index workshop—consultant’s report. Appendix 1 *In*: P.J. Rago, C.D. Stephen, and H.M. Austin (editors), *Report of the juvenile abundances indices workshop*. Atlantic States Marine Fisheries Commission, Special Report No. 48, Washington, D.C. 83 p.
- Welch, T.J., M.J. van den Avyle, R.K. Betsill, and E.M. Driebe. 1993. Precision and relative accuracy of striped bass age estimates from otoliths, scales, and anal fin rays and spines. *North American Journal of Fisheries Management* 13(3):616–620.
- Worth, S.G. 1904. Report on operations with the striped bass at the Weldon North Carolina sub-station in May 1904. Department of Commerce and Labor, Bureau of Fisheries.
- Zincone, L.H., and R.A. Rulifson. 1991. Instream flow and striped bass recruitment in the lower Roanoke River, North Carolina. *Rivers* 2(2):125–137.
- Zuur, A.F., E.N. Ieno, N.J. Walker, A.A. Saveliev, and G.M. Smith. 2009. *Mixed effects models and extensions in ecology with R*. Springer-Verlag, New York. 574 p.
- Zuur, A.F., A.A. Saveliev, and E.N. Ieno. 2012. *Zero inflated models and generalized linear mixed models with R*. Highland Statistics Ltd, United Kingdom. 324 p.

## 8 TABLES

**Table 1.1.** Parameter estimates and associated standard errors (in parentheses) of the von Bertalanffy age-length growth curve by sex. The function was fit to total length in centimeters.

<b>Sex</b>	<b>n</b>	<b><math>L_{\infty}</math></b>	<b><math>K</math></b>	<b><math>t_0</math></b>
Female	29,991	160 (0.81)	0.071 (0.00063)	-0.62 (0.014)
Male	29,691	161 (1.3)	0.064 (0.00082)	-0.87 (0.017)

**Table 1.2.** Parameter estimates and associated standard errors (in parentheses) of the length-weight function by sex. The function was fit to total length in centimeters and weight in kilograms.

<b>Sex</b>	<b>n</b>	<b><math>a</math></b>	<b><math>b</math></b>
Female	28,814	2.8E-06 (4.4E-08)	3.2 (2.3E-03)
Male	33,411	5.9E-06 (1.0E-07)	3.1 (2.7E-03)

**Table 1.3.** Percent maturity of female striped bass as estimated by Boyd (2011).

<b>Age</b>	<b>% Maturity</b>
<b>0</b>	0
<b>1</b>	0
<b>2</b>	0
<b>3</b>	28.6
<b>4</b>	96.8
<b>5</b>	100
<b>6</b>	100
<b>7</b>	100
<b>8</b>	100
<b>9</b>	100
<b>10</b>	100
<b>11</b>	100
<b>12</b>	100
<b>13</b>	100
<b>14</b>	100
<b>15</b>	100
<b>16</b>	100
<b>17</b>	100

**Table 1.4.** Age-constant estimates of natural mortality derived from life history characteristics.

<b>Method</b>	<b>Female</b>	<b>Male</b>	<b>Average</b>
Alverson and Carney 1975	0.37	0.44	0.40
Hoening 1983 (regression)	0.26	0.30	0.28
Hoening 1983 (rule-of-thumb)	0.25	0.28	0.26
Ralston 1987 (linear regression)	0.16	0.15	0.16
Jensen 1996 (theoretical)	0.11	0.095	0.10
Jensen 1996 (derived from Pauly 1980)	0.11	0.10	0.11
Cubillos 2003	0.099	0.090	0.094
Hewitt and Hoening 2005	0.25	0.28	0.26
Hoening (nls; from Then et al. 2015)	0.37	0.41	0.39
Then et al. 2015	0.30	0.34	0.32
Average	0.23	0.25	0.24

**Table 1.5.** Estimates of natural mortality at age by sex based on the method of Lorenzen (1996).

<b>Age</b>	<b>Female</b>	<b>Male</b>
<b>0</b>	2.8	2.2
<b>1</b>	1.4	1.3
<b>2</b>	1.0	1.0
<b>3</b>	0.88	0.88
<b>4</b>	0.79	0.80
<b>5</b>	0.73	0.74
<b>6</b>	0.69	0.70
<b>7</b>	0.66	0.67
<b>8</b>	0.64	0.65
<b>9</b>	0.62	0.63
<b>10</b>	0.60	0.62
<b>11</b>	0.59	0.60
<b>12</b>	0.58	0.59
<b>13</b>	0.57	0.58
<b>14</b>	0.56	0.57
<b>15</b>	0.56	0.57
<b>16</b>	0.55	0.56
<b>17</b>	0.55	0.56

**Table 1.6.** Changes in the total allowable landings (TAL) in metric tons and pounds (in parentheses) for the ASMA-RRMA, 1991–2017.

<b>Regulatory Period</b>	<b>ASMA Commercial</b>	<b>ASMA Recreational</b>	<b>RRMA Recreational</b>	<b>Combined TAL</b>
1991–1997	44.45 (98,000)	13.34 (29,400)	13.34 (29,400)	71.12 (156,800)
1998	56.88 (125,400)	28.44 (62,700)	28.44 (62,700)	113.8 (250,800)
1999	62.57 (137,940)	31.28 (68,970)	31.28 (68,970)	125.2 (275,968)
2000–2002	102.1 (225,000)	51.03 (112,500)	51.03 (112,500)	204.1 (450,000)
2003–2014	124.7 (275,000)	62.37 (137,500)	62.37 (137,500)	249.5 (550,000)
2015–2017	62.37 (137,500)	31.18 (68,750)	31.18 (68,750)	124.7 (275,000)

**Table 1.7.** Striped bass commercial landings and discards and recreational harvest and discards from the ASMA-RRMA, 1991–2017.

Year	Commercial Landings	Commercial Discards	Recreational Harvest		Recreational Discards	
	ASMA	ASMA	ASMA	RRMA	ASMA	RRMA
	metric tons	numbers	numbers	numbers	numbers	numbers
1991	49.24	10,267	14,395	26,934	1,507	9,516
1992	45.65	8,434	10,542	13,372	1,279	4,725
1993	49.70	8,952	11,404	14,325	847.4	5,061
1994	46.48	4,302	8,591	8,284		2,927
1995	39.88	4,938	7,343	7,471		3,373
1996	40.92	4,150	7,433	8,367		10,461
1997	43.64	3,967	6,901	9,364	1,969	18,673
1998	56.26	5,817	19,566	23,109	5,881	12,159
1999	73.94	7,401	16,967	22,479	2,581	10,468
2000	97.17	10,500	38,085	38,206	5,052	5,961
2001	100.0	11,630	40,127	35,231	3,931	4,544
2002	101.2	6,633	27,896	36,422	3,300	3,570
2003	120.9	10,394	15,124	11,157	1,618	2,448
2004	124.2	4,475	28,004	26,506	2,627	11,989
2005	105.6	9,566	17,954	34,122	1,358	10,093
2006	84.62	6,715	10,711	25,355	605.1	4,194
2007	77.94	4,803	7,143	19,305	870.3	3,360
2008	34.01	2,538	10,048	10,541	2,366	12,137
2009	43.49	3,294	12,069	23,248	2,596	8,702
2010	90.72	10,017	3,504	22,445	1,037	7,930
2011	61.86	6,646	13,341	22,102	1,381	6,894
2012	52.48	4,256	22,345	28,847	1,598	4,033
2013	31.03	6,706	4,299	7,718	1,048	4,750
2014	32.23	2,794	5,529	11,058	1,478	10,594
2015	51.98	3,539	23,240	20,031	3,170	6,927
2016	55.89	3,989	4,794	21,260	662.5	3,369
2017	34.50	2,762	4,215	9,899	1,578	5,021

**Table 2.1.** Annual estimates of commercial gill-net discards (numbers of fish), 1991–2017. Note that values prior to 2012 were estimated using a hindcasting approach.

<b>Year</b>	<b>Discards</b>
<b>1991</b>	10,267
<b>1992</b>	8,434
<b>1993</b>	8,952
<b>1994</b>	4,302
<b>1995</b>	4,938
<b>1996</b>	4,150
<b>1997</b>	3,967
<b>1998</b>	5,817
<b>1999</b>	7,401
<b>2000</b>	10,500
<b>2001</b>	11,630
<b>2002</b>	6,633
<b>2003</b>	10,394
<b>2004</b>	4,475
<b>2005</b>	9,566
<b>2006</b>	6,715
<b>2007</b>	4,803
<b>2008</b>	2,538
<b>2009</b>	3,294
<b>2010</b>	10,017
<b>2011</b>	6,646
<b>2012</b>	4,256
<b>2013</b>	6,706
<b>2014</b>	2,794
<b>2015</b>	3,539
<b>2016</b>	3,989
<b>2017</b>	2,762



**Table 2.2.** Annual estimates of recreational harvest and dead discards (numbers of fish) for the ASMA, 1991–2017.

<b>Year</b>	<b>Harvest</b>	<b>Discards</b>
<b>1991</b>	14,395	1,507
<b>1992</b>	10,542	1,279
<b>1993</b>	11,404	847
<b>1994</b>	8,591	
<b>1995</b>	7,343	
<b>1996</b>	7,433	
<b>1997</b>	6,901	1,969
<b>1998</b>	19,566	5,881
<b>1999</b>	16,967	2,581
<b>2000</b>	38,085	5,052
<b>2001</b>	40,127	3,931
<b>2002</b>	27,896	3,300
<b>2003</b>	15,124	1,618
<b>2004</b>	28,004	2,627
<b>2005</b>	17,954	1,358
<b>2006</b>	10,711	605
<b>2007</b>	7,143	870
<b>2008</b>	10,048	2,366
<b>2009</b>	12,069	2,596
<b>2010</b>	3,504	1,037
<b>2011</b>	13,341	1,381
<b>2012</b>	22,345	1,598
<b>2013</b>	4,299	1,048
<b>2014</b>	5,529	1,478
<b>2015</b>	23,240	3,170
<b>2016</b>	4,794	663
<b>2017</b>	4,215	1,578

**Table 2.3.** Annual estimates of recreational harvest and dead discards (numbers of fish) for the RRMA, 1991–2017. Note that discard values prior to 1995 were estimated using a hindcasting approach.

<b>Year</b>	<b>Harvest</b>	<b>Discards</b>
<b>1991</b>	26,934	9,516
<b>1992</b>	13,372	4,725
<b>1993</b>	14,325	5,061
<b>1994</b>	8,284	2,927
<b>1995</b>	7,471	3,373
<b>1996</b>	8,367	10,461
<b>1997</b>	9,364	18,673
<b>1998</b>	23,109	12,159
<b>1999</b>	22,479	10,468
<b>2000</b>	38,206	5,961
<b>2001</b>	35,231	4,544
<b>2002</b>	36,422	3,570
<b>2003</b>	11,157	2,448
<b>2004</b>	26,506	11,989
<b>2005</b>	34,122	10,093
<b>2006</b>	25,355	4,194
<b>2007</b>	19,305	3,360
<b>2008</b>	10,541	12,137
<b>2009</b>	23,248	8,702
<b>2010</b>	22,445	7,930
<b>2011</b>	22,102	6,894
<b>2012</b>	28,847	4,033
<b>2013</b>	7,718	4,750
<b>2014</b>	11,058	10,594
<b>2015</b>	20,031	6,927
<b>2016</b>	21,260	3,369
<b>2017</b>	4,215	5,021

**Table 3.1.** Annual estimates of commercial landings and recreational harvest that were input into the SS model, 1991–2017. Values assumed for the coefficients of variation (CVs) are also provided.

Year	ASMA Commercial		ASMA Recreational		RRMA Recreational	
	metric tons	CV	numbers	CV	numbers	CV
1991	49.24	0.02	14,395	0.02	26,934	0.02
1992	45.65	0.02	10,542	0.02	13,372	0.02
1993	49.70	0.02	11,404	0.02	14,325	0.02
1994	46.48	0.01	8,591	0.02	8,284	0.02
1995	39.88	0.01	7,343	0.02	7,471	0.02
1996	40.92	0.01	7,433	0.02	8,367	0.02
1997	43.64	0.01	6,901	0.02	9,364	0.02
1998	56.26	0.01	19,566	0.02	23,109	0.02
1999	73.94	0.01	16,967	0.02	22,479	0.02
2000	97.17	0.01	38,085	0.02	38,206	0.02
2001	99.99	0.01	40,127	0.02	35,231	0.02
2002	101.18	0.01	27,896	0.02	36,422	0.02
2003	120.91	0.01	15,124	0.02	11,157	0.02
2004	124.20	0.01	28,004	0.02	26,506	0.02
2005	105.64	0.01	17,954	0.02	34,122	0.02
2006	84.62	0.01	10,711	0.02	25,355	0.02
2007	77.94	0.01	7,143	0.02	19,305	0.02
2008	34.01	0.01	10,048	0.02	10,541	0.02
2009	43.49	0.01	12,069	0.02	23,248	0.02
2010	90.72	0.01	3,504	0.02	22,445	0.02
2011	61.86	0.01	13,341	0.02	22,102	0.02
2012	52.48	0.01	22,345	0.02	28,847	0.02
2013	31.03	0.01	4,299	0.02	7,718	0.02
2014	32.23	0.01	5,529	0.02	11,058	0.02
2015	51.98	0.01	23,240	0.02	20,031	0.02
2016	55.89	0.01	4,794	0.02	21,260	0.02
2017	34.50	0.01	4,215	0.02	9,899	0.02

**Table 3.2.** Annual estimates of dead discards that were input into the SS model, 1991–2017. Values assumed for the coefficients of variation (CVs) are also provided.

Year	Albemarle/Roanoke Commercial		Albemarle Sound Recreational		Roanoke River Recreational	
	numbers	CV	numbers	CV	numbers	CV
1991	10,267	0.82	1,507	0.060	9,516	0.06
1992	8,434	0.67	1,279	0.051	4,725	0.06
1993	8,952	0.72	847	0.034	5,061	0.06
1994	4,302	0.34			2,927	0.06
1995	4,938	0.40			3,373	0.04
1996	4,150	0.33			10,461	0.04
1997	3,967	0.32	1,969	0.079	18,673	0.04
1998	5,817	0.47	5,881	0.24	12,159	0.04
1999	7,401	0.59	2,581	0.10	10,468	0.04
2000	10,500	0.84	5,052	0.20	5,961	0.04
2001	11,630	0.93	3,931	0.16	4,544	0.04
2002	6,633	0.53	3,300	0.13	3,570	0.04
2003	10,394	0.83	1,618	0.065	2,448	0.04
2004	4,475	0.36	2,627	0.11	11,989	0.04
2005	9,566	0.77	1,358	0.054	10,093	0.04
2006	6,715	0.54	605	0.024	4,194	0.04
2007	4,803	0.38	870	0.035	3,360	0.04
2008	2,538	0.20	2,366	0.095	12,137	0.04
2009	3,294	0.26	2,596	0.10	8,702	0.04
2010	10,017	0.80	1,037	0.041	7,930	0.04
2011	6,646	0.53	1,381	0.055	6,894	0.04
2012	4,256	0.17	1,598	0.064	4,033	0.04
2013	6,706	0.27	1,048	0.042	4,750	0.04
2014	2,794	0.11	1,478	0.059	10,594	0.04
2015	3,539	0.14	3,170	0.13	6,927	0.04
2016	3,989	0.16	663	0.027	3,369	0.04
2017	2,762	0.11	1,578	0.063	5,021	0.04

**Table 3.3.** GLM-standardized indices of relative abundance derived from fisheries-independent surveys that were input into the SS model, 1991–2017. The empirically-derived standard errors (SEs) are also provided.

Year	Program 100 Juvenile		Program 135 Fall/Winter		Program 135 Spring		Roanoke River Electrofishing	
	Index	SE	Index	SE	Index	SE	Index	SE
1991	0.709	0.19	0.44	0.043				
1992	2.12	0.51	0.44	0.037	0.48	0.034		
1993	42.4	8.8	0.42	0.039	0.28	0.021		
1994	59.4	12	0.79	0.071	0.18	0.017	125	21
1995	8.54	1.8	0.31	0.024	0.94	0.063	42.1	7.0
1996	35.0	7.2	0.59	0.051	0.67	0.048	29.0	5.0
1997	5.12	1.1	0.54	0.031	0.84	0.057	75.7	12
1998	5.24	1.3	0.94	0.066	1.1	0.074	102	16
1999	0.968	0.26	0.49	0.034	1.1	0.069	92.1	15
2000	55.9	12	0.37	0.042	0.92	0.061	72.1	12
2001	3.52	0.82	0.50	0.053	1.1	0.072	210	35
2002	5.68	1.2	0.31	0.028	0.83	0.057	110	24
2003	0.253	0.095	0.80	0.060	0.38	0.029	221	39
2004	1.72	0.43	0.47	0.036	0.86	0.064	57.1	11
2005	23.0	4.8	0.65	0.057	0.71	0.051	104	17
2006	2.87	0.64	0.20	0.016	1.0	0.072	120	20
2007	4.94	1.1	0.83	0.085	0.41	0.031	53.0	8.8
2008	5.35	1.2	0.55	0.058	1.2	0.089	77.2	12
2009	0.363	0.11	0.54	0.048	0.71	0.057	76.5	13
2010	6.75	1.4	0.60	0.081	0.99	0.081	106	19
2011	15.3	3.2	0.20	0.018	1.1	0.094	46.3	7.7
2012	3.42	0.79	0.23	0.020	1.2	0.11	58.2	9.1
2013	0.369	0.11	0.37	0.032	1.4	0.12	39.6	7.6
2014	17.0	3.6	0.32	0.037	0.93	0.081	66.7	13
2015	18.4	3.8	0.17	0.017	0.51	0.039	46.4	9.1
2016	5.39	1.1	0.12	0.018	0.31	0.026	20.1	3.7
2017	1.29	0.30			0.36	0.030	14.5	2.5

**Table 3.4.** Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds.

ID	Label	Value	SD[Value]	Phase	Status
1	NatM_p_1_Fem_GP_1	0.40		-2	fixed
2	L_at_Amin_Fem_GP_1	17	0.050	3	estimated
3	L_at_Amax_Fem_GP_1	160	0.050	3	estimated
4	VonBert_K_Fem_GP_1	0.065	0.0010	3	estimated
5	CV_young_Fem_GP_1	0.19	0.0053	3	estimated
6	CV_old_Fem_GP_1	0.0010	8.4E-07	3	LO
7	Wtlen_1_Fem_GP_1	4.6E-06		-3	fixed
8	Wtlen_2_Fem_GP_1	3.2		-3	fixed
9	Mat50%_Fem_GP_1	1		-3	fixed
10	Mat_slope_Fem_GP_1	0		-3	fixed
11	Eggs/kg_inter_Fem_GP_1	1		-3	fixed
12	Eggs/kg_slope_wt_Fem_GP_1	0		-3	fixed
13	NatM_p_1_Mal_GP_1	0.40		-2	fixed
14	L_at_Amin_Mal_GP_1	18	0.050	4	estimated
15	L_at_Amax_Mal_GP_1	161	0.050	4	estimated
16	VonBert_K_Mal_GP_1	0.060	0.0011	4	estimated
17	CV_young_Mal_GP_1	0.19	0.0060	4	estimated
18	CV_old_Mal_GP_1	0.0010	8.0E-07	4	LO
19	Wtlen_1_Mal_GP_1	7.5E-06		-3	fixed
20	Wtlen_2_Mal_GP_1	3.1		-3	fixed
21	CohortGrowDev	1.0		-1	fixed
22	FracFemale_GP_1	0.50		-99	fixed
23	SR_LN(R0)	6.2	0.039	1	estimated
24	SR_BH_steep	0.90		-4	fixed
25	SR_sigmaR	0.60		-4	fixed
26	SR_regime	0		-4	fixed
27	SR_autocorr	0		-99	fixed
28	Main_InitAge_17	-0.37	0.52	4	estimated
29	Main_InitAge_16	-0.20	0.55	4	estimated
30	Main_InitAge_15	-0.23	0.55	4	estimated

**Table 3.4. (continued)** Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds.

ID	Label	Value	SD[Value]	Phase	Status
31	Main_InitAge_14	-0.30	0.53	4	estimated
32	Main_InitAge_13	-0.36	0.52	4	estimated
33	Main_InitAge_12	-0.38	0.50	4	estimated
34	Main_InitAge_11	-0.53	0.48	4	estimated
35	Main_InitAge_10	-0.75	0.45	4	estimated
36	Main_InitAge_9	-0.77	0.39	4	estimated
37	Main_InitAge_8	-0.76	0.34	4	estimated
38	Main_InitAge_7	-0.79	0.31	4	estimated
39	Main_InitAge_6	-0.88	0.30	4	estimated
40	Main_InitAge_5	-0.70	0.28	4	estimated
41	Main_InitAge_4	-0.23	0.22	4	estimated
42	Main_InitAge_3	0.65	0.091	4	estimated
43	Main_InitAge_2	0.037	0.11	4	estimated
44	Main_InitAge_1	-0.48	0.12	4	estimated
45	Main_RecrDev_1991	-0.54	0.12	4	estimated
46	Main_RecrDev_1992	-0.25	0.11	4	estimated
47	Main_RecrDev_1993	0.72	0.081	4	estimated
48	Main_RecrDev_1994	1.2	0.076	4	estimated
49	Main_RecrDev_1995	0.89	0.099	4	estimated
50	Main_RecrDev_1996	1.6	0.074	4	estimated
51	Main_RecrDev_1997	0.81	0.11	4	estimated
52	Main_RecrDev_1998	1.2	0.086	4	estimated
53	Main_RecrDev_1999	0.36	0.14	4	estimated
54	Main_RecrDev_2000	1.5	0.062	4	estimated
55	Main_RecrDev_2001	0.38	0.098	4	estimated
56	Main_RecrDev_2002	0.00039	0.085	4	estimated
57	Main_RecrDev_2003	-0.92	0.13	4	estimated
58	Main_RecrDev_2004	-0.12	0.088	4	estimated
59	Main_RecrDev_2005	0.81	0.077	4	estimated
60	Main_RecrDev_2006	0.47	0.098	4	estimated

**Table 3.4. (continued)** Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds.

ID	Label	Value	SD[Value]	Phase	Status
61	Main_RecrDev_2007	0.56	0.083	4	estimated
62	Main_RecrDev_2008	-0.24	0.082	4	estimated
63	Main_RecrDev_2009	-1.6	0.12	4	estimated
64	Main_RecrDev_2010	0.065	0.077	4	estimated
65	Main_RecrDev_2011	0.77	0.059	4	estimated
66	Main_RecrDev_2012	-0.0074	0.089	4	estimated
67	Main_RecrDev_2013	-0.91	0.16	4	estimated
68	Main_RecrDev_2014	0.43	0.095	4	estimated
69	Main_RecrDev_2015	0.39	0.11	4	estimated
70	Main_RecrDev_2016	0.020	0.13	4	estimated
71	Main_RecrDev_2017	-0.47	0.15	4	estimated
72	InitF_seas_1flt_1ARcomm	0.085	0.0064	1	estimated
73	InitF_seas_1flt_2ASrec	0.011	0.00055	1	estimated
74	InitF_seas_1flt_3RRrecharv	0.019	0.00089	1	estimated
75	InitF_seas_1flt_8RRrecdisc	0.0057	0.00031	1	LO
76	LnQ_base_P100juv(4)	-8.2	0.56	5	estimated
77	Q_power_P100juv(4)	0.60	0.086	6	estimated
78	LnQ_base_P135fw(5)	-3.0	0.17	5	estimated
79	Q_power_P135fw(5)	-0.54	0.033	6	estimated
80	LnQ_base_P135spr(6)	-1.7	0.19	5	estimated
81	Q_power_P135spr(6)	-0.74	0.033	6	estimated
82	LnQ_base_RRef(7)	1.8	0.22	5	estimated
83	Q_power_RRef(7)	-0.37	0.056	6	estimated
84	SizeSpline_Code_ARcomm(1)	2.0		-99	fixed
85	SizeSpline_GradLo_ARcomm(1)	0.060	0.046	3	estimated
86	SizeSpline_GradHi_ARcomm(1)	0.0010	9.0E-05	3	HI
87	SizeSpline_Knot_1_ARcomm(1)	29		-99	fixed
88	SizeSpline_Knot_2_ARcomm(1)	45		-99	fixed
89	SizeSpline_Knot_3_ARcomm(1)	49		-99	fixed
90	SizeSpline_Knot_4_ARcomm(1)	52		-99	fixed



**Table 3.4. (continued)** Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds.

ID	Label	Value	SD[Value]	Phase	Status
91	SizeSpline_Knot_5_ARcomm(1)	55		-99	fixed
92	SizeSpline_Knot_6_ARcomm(1)	88		-99	fixed
93	SizeSpline_Val_1_ARcomm(1)	-6.1	0.29	2	estimated
94	SizeSpline_Val_2_ARcomm(1)	-4.4	0.23	2	estimated
95	SizeSpline_Val_3_ARcomm(1)	-2.1	0.13	2	estimated
96	SizeSpline_Val_4_ARcomm(1)	-1.0		-99	fixed
97	SizeSpline_Val_5_ARcomm(1)	-1.1	0.072	2	estimated
98	SizeSpline_Val_6_ARcomm(1)	-2.6	0.30	2	estimated
99	Retain_L_infl_ARcomm(1)	30	3.6	1	estimated
100	Retain_L_width_ARcomm(1)	9.6	1.7	2	estimated
101	Retain_L_asymptote_logit_ARcomm(1)	999		-4	fixed
102	Retain_L_maleoffset_ARcomm(1)	0		-4	fixed
103	Size_DbIN_peak_ASrec(2)	53	0.28	1	estimated
104	Size_DbIN_top_logit_ASrec(2)	0.13	209	1	estimated
105	Size_DbIN_ascend_se_ASrec(2)	3.7	0.057	2	estimated
106	Size_DbIN_descend_se_ASrec(2)	3.5	123	2	estimated
107	Size_DbIN_start_logit_ASrec(2)	-999		-4	fixed
108	Size_DbIN_end_logit_ASrec(2)	15		-5	fixed
109	Retain_L_infl_ASrec(2)	40	0.38	1	estimated
110	Retain_L_width_ASrec(2)	5.1	0.19	2	estimated
111	Retain_L_asymptote_logit_ASrec(2)	999		-4	fixed
112	Retain_L_maleoffset_ASrec(2)	0		-4	fixed
113	Size_DbIN_peak_RRrecharv(3)	46		-3	fixed
114	Size_DbIN_top_logit_RRrecharv(3)	-2.2		-3	fixed
115	Size_DbIN_ascend_se_RRrecharv(3)	-4.0		-4	fixed
116	Size_DbIN_descend_se_RRrecharv(3)	-2.0		-4	fixed
117	Size_DbIN_start_logit_RRrecharv(3)	-999		-4	fixed
118	Size_DbIN_end_logit_RRrecharv(3)	-999		-5	fixed
119	SizeSpline_Code_P135fw(5)	2.0		-99	fixed
120	SizeSpline_GradLo_P135fw(5)	0.56	0.11	3	estimated

**Table 3.4. (continued)** Parameter values, standard deviations (SD), phase of estimation, and status from the base run of the stock assessment model. LO or HI indicates parameter values estimated near their bounds.

ID	Label	Value	SD[Value]	Phase	Status
121	SizeSpline_GradHi_P135fw(5)	-0.41	0.091	3	estimated
122	SizeSpline_Knot_1_P135fw(5)	25		-99	fixed
123	SizeSpline_Knot_2_P135fw(5)	42		-99	fixed
124	SizeSpline_Knot_3_P135fw(5)	57		-99	fixed
125	SizeSpline_Val_1_P135fw(5)	-4.6	0.38	2	estimated
126	SizeSpline_Val_2_P135fw(5)	-1.0		-99	fixed
127	SizeSpline_Val_3_P135fw(5)	-1.4	0.26	2	estimated
128	Size_DblN_peak_P135spr(6)	47	2.2	1	estimated
129	Size_DblN_top_logit_P135spr(6)	-0.018	222	1	estimated
130	Size_DblN_ascend_se_P135spr(6)	5.1	0.22	2	estimated
131	Size_DblN_descend_se_P135spr(6)	3.5	123	2	estimated
132	Size_DblN_start_logit_P135spr(6)	-999		-4	fixed
133	Size_DblN_end_logit_P135spr(6)	15		-5	fixed
134	Size_DblN_peak_RRef(7)	57	1.1	1	estimated
135	Size_DblN_top_logit_RRef(7)	0.014	219	1	estimated
136	Size_DblN_ascend_se_RRef(7)	4.4	0.099	2	estimated
137	Size_DblN_descend_se_RRef(7)	3.5	123	2	estimated
138	Size_DblN_start_logit_RRef(7)	-999		-4	fixed
139	Size_DblN_end_logit_RRef(7)	15		-5	fixed
140	SzSel_MaleDogleg_RRef(7)	59	1.8	1	estimated
141	SzSel_MaleatZero_RRef(7)	7.9	1.1	1	estimated
142	SzSel_MaleatDogleg_RRef(7)	0		-4	fixed
143	SzSel_MaleatMaxage_RRef(7)	-6.2	5.6	2	estimated
144	Size_DblN_peak_RRecdisc(8)	51	0.69	3	estimated
145	Size_DblN_top_logit_RRecdisc(8)	0.052	222	3	estimated
146	Size_DblN_ascend_se_RRecdisc(8)	4.4	0.095	4	estimated
147	Size_DblN_descend_se_RRecdisc(8)	3.5	123	4	estimated
148	Size_DblN_start_logit_RRecdisc(8)	-999		-4	fixed
149	Size_DblN_end_logit_RRecdisc(8)	15		-5	fixed

**Table 3.5.** Results of the base run compared to the results of 50 jitter trials in which initial parameter values were jittered by 10%. A single asterisk (\*) indicates that the Hessian matrix did not invert. Two asterisks (\*\*) indicate that the convergence level was greater than 1.

<b>Run</b>	<b>Total LL</b>	<b>SSB<sub>2017</sub></b>	<b>SSB<sub>Threshold</sub></b>	<b><i>F</i><sub>2017</sub></b>	<b><i>F</i><sub>Threshold</sub></b>
<b>base</b>	4,879	35.6	121	0.266	0.18
<b>1</b>	*				
<b>2</b>	**				
<b>3</b>	**				
<b>4</b>	*				
<b>5</b>	*				
<b>6</b>	*				
<b>7</b>	5,061	41.7	115	0.22	0.18
<b>8</b>	4,879	35.3	121	0.27	0.18
<b>9</b>	*				
<b>10</b>	4,956	35.5	115	0.26	0.18
<b>11</b>	*				
<b>12</b>	6,138	51.3	29.7	0.05	0.30
<b>13</b>	*				
<b>14</b>	4,879	35.3	121	0.27	0.18
<b>15</b>	4,879	35.6	121	0.27	0.18
<b>16</b>	4,879	35.6	121	0.27	0.18
<b>17</b>	5,298	45.5	40.2	0.07	0.20
<b>18</b>	**				
<b>19</b>	**				
<b>20</b>	4,879	35.6	121	0.27	0.18
<b>21</b>	*				
<b>22</b>	**				
<b>23</b>	4,879	35.3	121	0.27	0.18
<b>24</b>	*				
<b>25</b>	*				

**Table 3.5. (continued)** Results of the base run compared to the results of 50 jitter trials in which initial parameter values were jittered by 10%. A single asterisk (\*) indicates that the Hessian matrix did not invert. Two asterisks (\*\*) indicate that the convergence level was greater than 1.

<b>Run</b>	<b>Total LL</b>	<b>SSB<sub>2017</sub></b>	<b>SSB<sub>Threshold</sub></b>	<b>F<sub>2017</sub></b>	<b>F<sub>Threshold</sub></b>
<b>26</b>	4,879	35.3	121	0.27	0.18
<b>27</b>	4,879	35.3	121	0.27	0.18
<b>28</b>	*				
<b>29</b>	4,886	35.6	122	0.27	0.19
<b>30</b>	*				
<b>31</b>	4,879	35.3	121	0.27	0.18
<b>32</b>	**				
<b>33</b>	**				
<b>34</b>	**				
<b>35</b>	4,879	35.3	121	0.27	0.18
<b>36</b>	*				
<b>37</b>	*				
<b>38</b>	7,009	50.4	42	0.087	0.19
<b>39</b>	4,956	35.5	115	0.26	0.18
<b>40</b>	**				
<b>41</b>	*				
<b>42</b>	*				
<b>43</b>	4,879	35.6	121	0.27	0.18
<b>44</b>	4,879	35.6	121	0.27	0.18
<b>45</b>	**				
<b>46</b>	7,390	1,667	739	0.026	0.27
<b>47</b>	*				
<b>48</b>	**				
<b>49</b>	*				
<b>50</b>	4,879	35.6	121	0.27	0.18

**Table 3.6.** Results of the runs test for temporal patterns and results of the Shapiro-Wilk test for normality applied to the standardized residuals of the fits to the fisheries-independent survey indices from the base run of the assessment model. *P*-values were considered significant at  $\alpha = 0.05$ .

Survey	Runs Test		Shapiro-Wilk	
	median	<i>P</i> -value	W	<i>P</i> -value
P100juv	-0.029	0.70	0.98	0.80
P135fw	0.016	1.0	0.98	0.81
P135spr	0.017	0.31	0.97	0.70
RRef	0.019	0.30	0.97	0.67

**Table 3.7.** Annual estimates of recruitment (thousands of fish), female spawning stock biomass (SSB; metric tons), and spawner potential ratio (SPR) and associated standard deviations (SDs) from the base run of the stock assessment model, 1991–2017.

Year	Recruitment		SSB		SPR	
	Value	SD	Value	SD	Value	SD
1991	227	27	148	10	0.22	0.012
1992	299	30	129	8.0	0.30	0.011
1993	780	57	116	7.0	0.26	0.011
1994	1,211	83	87	6.1	0.25	0.013
1995	876	82	67	4.9	0.23	0.011
1996	1,720	110	66	4.0	0.23	0.0096
1997	850	88	105	5.5	0.31	0.012
1998	1,284	98	165	8.2	0.31	0.012
1999	564	79	203	10	0.35	0.012
2000	1,736	87	266	12	0.29	0.010
2001	583	53	255	12	0.28	0.010
2002	398	31	243	11	0.28	0.010
2003	157	20	220	10	0.32	0.010
2004	356	29	259	8.1	0.27	0.0062
2005	889	60	209	5.7	0.24	0.0061
2006	618	57	140	4.2	0.20	0.0065
2007	643	46	81	3.3	0.14	0.0061
2008	277	20	60	3.1	0.21	0.0078
2009	75	9	94	4.6	0.24	0.0096
2010	404	28	108	4.6	0.22	0.0082
2011	810	40	100	2.7	0.21	0.0054
2012	357	29	68	1.7	0.11	0.0044
2013	111	17	21	1.0	0.13	0.0053
2014	510	49	41	1.9	0.20	0.0065
2015	541	62	76	2.7	0.17	0.0058
2016	359	49	58	2.3	0.16	0.0076
2017	202	31	36	2.7	0.18	0.012

**Table 3.8.** Predicted population numbers (numbers of fish) at age at the beginning of the year from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	226,690	168,260	188,106	233,819	63,912	25,981	13,654	9,380	6,190	3,942	2,602	2,091	1,583	1,047	721	502	336	528
1992	298,814	151,951	112,634	125,023	136,282	24,395	7,538	4,169	3,328	2,451	1,652	1,118	908	690	457	315	219	378
1993	779,868	200,297	101,736	75,069	77,339	64,844	9,498	2,946	1,778	1,527	1,172	806	550	448	341	226	156	295
1994	1,211,036	522,750	134,083	67,734	45,664	34,408	22,844	3,376	1,163	766	690	542	376	258	210	160	106	212
1995	875,700	811,762	349,814	89,216	41,084	19,718	11,354	7,542	1,252	478	333	309	246	171	118	96	73	146
1996	1,720,200	586,983	543,056	232,456	53,319	16,624	5,845	3,361	2,552	476	195	140	132	106	74	51	41	94
1997	850,404	1,153,053	392,701	360,342	138,727	21,982	5,069	1,757	1,136	961	191	81	59	56	45	31	22	58
1998	1,283,700	570,034	771,993	261,187	222,840	67,949	8,925	2,033	754	520	457	93	39	29	27	22	15	39
1999	564,216	860,478	381,751	514,639	162,098	108,982	27,753	3,635	887	349	249	222	45	19	14	13	11	27
2000	1,736,040	378,201	576,252	254,690	323,729	83,014	47,650	12,152	1,702	440	179	130	116	24	10	7	7	20
2001	582,912	1,163,685	253,259	384,410	157,504	153,276	32,110	18,429	5,091	762	205	85	62	56	11	5	4	13
2002	398,252	390,732	779,193	168,910	236,515	72,748	56,893	11,898	7,437	2,208	344	94	39	29	26	5	2	8
2003	157,198	266,953	261,601	519,606	103,739	108,157	26,827	21,318	4,941	3,354	1,042	166	46	19	14	13	3	5
2004	355,698	105,371	178,669	174,420	326,834	51,302	43,366	10,649	9,240	2,326	1,659	528	85	24	10	7	7	4
2005	889,434	238,426	70,529	118,948	106,898	148,739	18,382	15,420	4,162	3,930	1,039	759	244	40	11	5	3	5
2006	617,552	596,193	159,578	46,919	71,316	44,860	48,553	6,191	5,931	1,778	1,777	483	357	115	19	5	2	4
2007	642,528	413,945	398,816	106,011	27,249	25,795	11,768	13,588	2,106	2,341	760	788	217	162	52	8	2	3
2008	277,352	430,673	276,335	263,098	56,240	6,450	3,405	1,699	2,766	562	726	253	271	76	56	18	3	2
2009	75,442	185,910	288,136	183,127	153,665	21,566	1,767	911	513	931	202	268	95	102	29	21	7	2
2010	404,054	50,569	124,449	191,666	109,788	65,088	7,117	592	343	212	404	90	121	43	46	13	10	4
2011	809,868	270,836	33,815	82,579	113,573	42,732	18,416	2,083	207	139	94	186	42	57	20	22	6	6
2012	357,286	542,855	181,202	22,451	48,267	42,752	11,647	5,122	675	76	55	38	77	17	24	8	9	5
2013	110,836	239,483	362,573	119,121	10,411	6,946	2,761	821	530	93	12	9	7	14	3	4	2	3
2014	509,662	74,290	159,688	237,869	61,499	2,172	691	274	115	100	21	3	2	2	4	1	1	1
2015	541,110	341,625	49,683	105,708	137,920	22,681	561	177	82	39	37	8	1	1	1	1	0	1
2016	358,590	362,706	228,496	32,914	59,484	44,092	4,617	110	40	21	11	11	2	0	0	0	0	0
2017	201,758	240,360	242,368	151,168	18,131	16,999	7,995	913	29	13	8	4	4	1	0	0	0	0

**Table 3.9.** Predicted population numbers (numbers of fish) at age at mid-year from the base run of the stock assessment model, 1991–2017.

<b>Year</b>	<b>0</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>	<b>8</b>	<b>9</b>	<b>10</b>	<b>11</b>	<b>12</b>	<b>13</b>	<b>14</b>	<b>15</b>	<b>16</b>	<b>17</b>
<b>1991</b>	185,596	137,665	153,355	178,506	39,479	13,994	7,544	5,587	3,895	2,551	1,706	1,378	1,046	692	477	332	222	349
<b>1992</b>	244,646	124,334	91,953	98,331	93,998	15,222	4,712	2,722	2,255	1,695	1,154	784	638	486	322	222	154	266
<b>1993</b>	638,495	163,879	83,012	58,548	51,580	38,486	5,662	1,851	1,167	1,027	797	551	377	307	234	155	107	202
<b>1994</b>	991,500	427,629	109,372	52,752	30,003	19,764	13,126	2,056	745	505	462	365	254	174	142	108	72	143
<b>1995</b>	716,952	663,952	285,161	68,969	26,130	10,735	6,177	4,387	772	305	216	202	161	113	77	63	48	96
<b>1996</b>	1,408,361	480,113	442,364	179,575	34,230	9,179	3,204	1,954	1,566	302	125	91	86	69	48	33	27	61
<b>1997</b>	696,247	943,477	320,264	283,368	97,083	14,007	3,210	1,151	768	662	133	56	41	39	31	22	15	40
<b>1998</b>	1,050,997	466,488	630,316	205,761	155,829	43,425	5,696	1,342	513	359	318	65	28	20	19	15	11	27
<b>1999</b>	461,938	704,168	311,814	408,170	115,996	72,061	18,364	2,487	624	250	179	161	33	14	10	10	8	19
<b>2000</b>	1,421,338	309,488	470,656	200,285	222,738	51,628	29,633	7,865	1,139	300	123	89	80	16	7	5	5	14
<b>2001</b>	477,245	952,227	206,828	301,525	107,033	93,380	19,546	11,707	3,352	512	139	58	42	38	8	3	2	9
<b>2002</b>	326,059	319,712	636,296	132,372	159,925	44,176	34,825	7,667	4,994	1,517	239	66	27	20	18	4	2	5
<b>2003</b>	128,701	218,394	213,608	412,096	72,947	68,484	16,902	14,035	3,390	2,359	742	119	33	14	10	9	2	3
<b>2004</b>	291,217	86,208	145,782	136,546	220,461	30,708	25,859	6,657	6,026	1,554	1,123	359	58	16	7	5	4	3
<b>2005</b>	728,199	195,058	57,526	92,102	69,239	84,979	10,668	9,562	2,720	2,643	708	520	168	27	8	3	2	3
<b>2006</b>	505,602	487,618	130,066	35,756	42,880	22,975	25,683	3,610	3,726	1,162	1,183	324	240	78	13	4	1	3
<b>2007</b>	526,041	338,213	323,925	77,210	13,248	9,370	4,470	6,127	1,088	1,303	438	462	128	96	31	5	1	2
<b>2008</b>	227,074	352,268	224,954	201,066	34,819	3,376	1,762	933	1,604	337	441	155	166	46	35	11	2	1
<b>2009</b>	61,766	152,106	235,001	141,791	99,996	12,389	1,023	559	329	614	134	180	64	68	19	14	5	1
<b>2010</b>	330,805	41,352	101,375	147,538	68,481	34,620	3,850	350	218	141	274	61	83	29	32	9	7	3
<b>2011</b>	663,054	221,530	27,553	63,132	69,667	22,308	9,712	1,185	125	87	60	120	27	37	13	14	4	4
<b>2012</b>	292,513	443,650	146,918	15,287	18,284	10,862	3,091	1,646	251	30	23	16	32	7	10	4	4	2
<b>2013</b>	90,741	195,557	293,675	85,586	4,751	2,190	870	306	230	44	6	5	3	7	2	2	1	1
<b>2014</b>	417,269	60,753	129,924	181,124	37,339	1,104	350	150	67	61	13	2	1	1	2	1	1	1
<b>2015</b>	443,017	279,392	40,438	79,294	77,954	10,232	249	84	42	21	20	4	1	1	0	1	0	0
<b>2016</b>	293,582	296,493	185,853	24,428	31,785	18,774	2,053	56	23	13	7	7	1	0	0	0	0	0
<b>2017</b>	165,182	196,503	197,152	114,032	10,402	7,901	3,755	476	16	8	5	3	3	1	0	0	0	0



**Table 3.10.** Predicted landings at age (numbers of fish) for the ARcomm fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	1	71	343	5,471	6,939	4,564	2,537	1,507	802	424	249	188	139	91	62	43	29	46
1992	1	56	180	2,626	14,205	4,219	1,355	632	401	244	146	93	73	55	36	25	17	30
1993	3	84	185	1,781	8,912	12,240	1,869	492	237	168	115	74	49	40	30	20	14	26
1994	6	280	310	2,048	6,627	8,068	5,564	702	194	106	85	63	43	29	24	18	12	24
1995	5	509	948	3,137	6,788	5,182	3,098	1,768	237	75	47	41	32	22	15	12	9	19
1996	9	353	1,410	7,831	8,514	4,236	1,538	755	461	72	26	18	16	13	9	6	5	11
1997	3	414	609	7,365	14,253	3,764	897	261	133	93	16	6	5	4	3	2	2	4
1998	3	163	953	4,251	18,195	9,279	1,264	242	71	40	31	6	2	2	2	1	1	2
1999	2	253	485	8,674	13,903	15,772	4,171	458	88	29	18	15	3	1	1	1	1	2
2000	5	121	796	4,627	29,136	12,388	7,379	1,585	176	37	13	9	8	2	1	1	0	1
2001	2	401	377	7,519	15,131	24,258	5,271	2,552	560	69	16	6	5	4	1	0	0	1
2002	1	149	1,284	3,653	25,030	12,703	10,383	1,845	920	226	31	8	3	2	2	0	0	1
2003	1	130	553	14,578	14,580	25,101	6,437	4,322	799	449	124	19	5	2	2	1	0	1
2004	1	48	351	4,496	41,186	10,561	9,239	1,921	1,330	277	175	53	8	2	1	1	1	0
2005	4	113	145	3,178	13,613	30,847	4,009	2,893	628	492	116	80	25	4	1	0	0	0
2006	4	388	448	1,689	11,656	11,653	13,435	1,508	1,183	297	265	68	49	16	3	1	0	1
2007	8	540	2,241	7,346	7,529	10,445	5,107	5,422	717	686	201	198	53	39	13	2	1	1
2008	1	252	698	8,544	8,469	1,531	834	354	463	78	90	30	31	9	6	2	0	0
2009	0	79	527	4,351	17,469	3,992	342	151	68	102	20	25	8	9	3	2	1	0
2010	3	39	413	8,231	21,876	20,587	2,371	173	82	42	72	15	20	7	8	2	2	1
2011	4	160	86	2,714	17,182	10,254	4,629	453	37	20	12	23	5	7	2	3	1	1
2012	4	616	885	1,276	9,669	12,003	3,488	1,407	157	15	10	6	13	3	4	1	1	1
2013	2	396	2,580	10,352	3,474	3,242	1,343	363	200	31	4	3	2	4	1	1	0	1
2014	3	53	492	9,393	11,112	614	203	70	24	17	3	0	0	0	0	0	0	0
2015	3	234	147	3,949	22,544	5,624	143	39	15	6	5	1	0	0	0	0	0	0
2016	3	358	974	1,758	13,414	15,131	1,701	37	11	5	2	2	0	0	0	0	0	0
2017	2	220	955	7,576	4,002	5,752	2,837	286	7	3	2	1	1	0	0	0	0	0

**Table 3.11.** Predicted dead discards at age (numbers of fish) for the ARcomm fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	3	112	257	856	714	376	163	70	24	7	2	1	0	0	0	0	0	0
1992	3	88	135	411	1,462	348	87	29	12	4	1	0	0	0	0	0	0	0
1993	9	133	138	279	917	1,008	121	23	7	3	1	0	0	0	0	0	0	0
1994	19	442	232	321	682	665	359	33	6	2	1	0	0	0	0	0	0	0
1995	16	804	710	491	699	427	200	82	7	1	0	0	0	0	0	0	0	0
1996	30	557	1,055	1,226	876	349	99	35	14	1	0	0	0	0	0	0	0	0
1997	9	653	456	1,153	1,467	310	58	12	4	2	0	0	0	0	0	0	0	0
1998	11	257	713	665	1,872	764	82	11	2	1	0	0	0	0	0	0	0	0
1999	5	399	363	1,358	1,431	1,299	269	21	3	0	0	0	0	0	0	0	0	0
2000	16	190	596	724	2,998	1,020	476	74	5	1	0	0	0	0	0	0	0	0
2001	6	633	282	1,177	1,557	1,998	340	119	17	1	0	0	0	0	0	0	0	0
2002	4	235	961	572	2,576	1,047	670	86	27	4	0	0	0	0	0	0	0	0
2003	2	206	414	2,282	1,500	2,068	415	201	24	8	1	0	0	0	0	0	0	0
2004	5	76	263	704	4,238	870	596	89	40	5	2	0	0	0	0	0	0	0
2005	12	179	109	497	1,401	2,541	259	135	19	8	1	0	0	0	0	0	0	0
2006	12	612	336	264	1,200	960	866	70	35	5	2	0	0	0	0	0	0	0
2007	24	852	1,678	1,150	775	861	329	252	21	12	2	1	0	0	0	0	0	0
2008	5	398	522	1,337	872	126	54	16	14	1	1	0	0	0	0	0	0	0
2009	1	124	395	681	1,798	329	22	7	2	2	0	0	0	0	0	0	0	0
2010	9	61	309	1,288	2,252	1,696	153	8	2	1	1	0	0	0	0	0	0	0
2011	14	253	65	425	1,768	845	299	21	1	0	0	0	0	0	0	0	0	0
2012	12	973	663	200	996	990	225	65	5	0	0	0	0	0	0	0	0	0
2013	5	625	1,931	1,620	358	268	87	17	6	1	0	0	0	0	0	0	0	0
2014	11	84	368	1,470	1,144	51	13	3	1	0	0	0	0	0	0	0	0	0
2015	11	369	110	618	2,321	464	9	2	0	0	0	0	0	0	0	0	0	0
2016	10	566	729	275	1,381	1,248	110	2	0	0	0	0	0	0	0	0	0	0
2017	5	347	715	1,186	412	474	183	13	0	0	0	0	0	0	0	0	0	0

**Table 3.12.** Predicted harvest at age (numbers of fish) for the ASrec fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	0	0	76	3,143	3,292	2,256	1,548	1,232	876	576	385	311	236	156	108	75	50	79
1992	0	0	31	1,198	5,351	1,656	656	411	348	263	179	122	99	76	50	34	24	41
1993	0	0	33	834	3,448	4,933	928	328	211	187	145	100	69	56	43	28	19	37
1994	0	0	45	767	2,049	2,598	2,207	373	138	94	86	68	47	32	27	20	13	27
1995	0	0	130	1,120	2,002	1,592	1,172	897	161	64	45	42	34	24	16	13	10	20
1996	0	0	174	2,520	2,263	1,172	524	345	282	55	23	16	16	12	9	6	5	11
1997	0	0	66	2,072	3,312	911	267	104	71	62	12	5	4	4	3	2	1	4
1998	0	0	241	2,804	9,911	5,266	883	226	89	62	55	11	5	4	3	3	2	5
1999	0	0	80	3,742	4,953	5,854	1,908	281	72	29	21	19	4	2	1	1	1	2
2000	0	0	232	3,507	18,238	8,080	5,931	1,707	253	67	28	20	18	4	2	1	1	3
2001	0	0	113	5,851	9,724	16,241	4,349	2,823	827	127	34	14	10	9	2	1	1	2
2002	0	0	266	1,968	11,135	5,888	5,929	1,413	941	287	45	12	5	4	3	1	0	1
2003	0	0	50	3,423	2,827	5,071	1,602	1,442	356	249	79	13	3	1	1	1	0	0
2004	0	0	59	1,964	14,858	3,969	4,278	1,192	1,103	286	207	66	11	3	1	1	1	0
2005	0	0	19	1,089	3,854	9,097	1,457	1,409	409	399	107	79	25	4	1	0	0	1
2006	0	0	44	431	2,457	2,558	3,635	547	574	179	183	50	37	12	2	1	0	0
2007	0	0	150	1,281	1,084	1,566	944	1,346	238	283	95	100	28	21	7	1	0	0
2008	0	0	134	4,283	3,506	660	442	253	442	93	122	43	46	13	10	3	1	0
2009	0	0	104	2,230	7,394	1,759	186	110	66	124	27	36	13	14	4	3	1	0
2010	0	0	12	607	1,332	1,306	185	18	11	7	14	3	4	2	2	0	0	0
2011	0	0	14	1,147	5,995	3,726	2,072	272	29	20	14	28	6	9	3	3	1	1
2012	0	0	290	1,088	6,812	8,805	3,152	1,706	255	30	23	16	32	7	10	4	4	2
2013	0	0	219	2,285	633	615	314	114	84	16	2	2	1	2	1	1	0	0
2014	0	0	53	2,636	2,576	148	60	28	13	11	2	0	0	0	0	0	0	0
2015	0	0	47	3,310	15,606	4,053	127	46	23	11	11	2	0	0	0	0	0	0
2016	0	0	64	300	1,889	2,219	307	9	4	2	1	1	0	0	0	0	0	0
2017	0	0	79	1,627	710	1,062	645	87	3	1	1	0	0	0	0	0	0	0

**Table 3.13.** Predicted dead discards at age (numbers of fish) for the ASrec fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	0	0	42	789	457	175	63	23	7	2	1	0	0	0	0	0	0	0
1992	0	0	17	301	743	129	27	8	3	1	0	0	0	0	0	0	0	0
1993	0	0	18	210	479	384	38	6	2	1	0	0	0	0	0	0	0	0
1994	0	0	25	193	284	202	90	7	1	0	0	0	0	0	0	0	0	0
1995	0	0	72	281	278	124	48	17	1	0	0	0	0	0	0	0	0	0
1996	0	0	96	633	314	91	21	7	2	0	0	0	0	0	0	0	0	0
1997	0	0	36	521	460	71	11	2	1	0	0	0	0	0	0	0	0	0
1998	0	0	133	704	1,376	410	36	4	1	0	0	0	0	0	0	0	0	0
1999	0	0	44	940	687	455	77	5	1	0	0	0	0	0	0	0	0	0
2000	0	0	128	881	2,531	628	241	33	2	0	0	0	0	0	0	0	0	0
2001	0	0	62	1,470	1,350	1,263	176	54	7	0	0	0	0	0	0	0	0	0
2002	0	0	147	494	1,546	458	241	27	8	1	0	0	0	0	0	0	0	0
2003	0	0	28	860	392	395	65	28	3	1	0	0	0	0	0	0	0	0
2004	0	0	32	493	2,062	309	174	23	9	1	0	0	0	0	0	0	0	0
2005	0	0	11	274	535	708	59	27	3	1	0	0	0	0	0	0	0	0
2006	0	0	24	108	341	199	148	10	5	1	0	0	0	0	0	0	0	0
2007	0	0	83	322	151	122	38	26	2	1	0	0	0	0	0	0	0	0
2008	0	0	74	1,076	487	52	18	5	4	0	0	0	0	0	0	0	0	0
2009	0	0	57	560	1,027	137	8	2	1	0	0	0	0	0	0	0	0	0
2010	0	0	6	152	185	102	8	0	0	0	0	0	0	0	0	0	0	0
2011	0	0	8	288	832	290	84	5	0	0	0	0	0	0	0	0	0	0
2012	0	0	160	273	947	686	128	33	2	0	0	0	0	0	0	0	0	0
2013	0	0	121	574	88	48	13	2	1	0	0	0	0	0	0	0	0	0
2014	0	0	29	662	358	12	2	1	0	0	0	0	0	0	0	0	0	0
2015	0	0	26	832	2,167	316	5	1	0	0	0	0	0	0	0	0	0	0
2016	0	0	35	75	262	173	13	0	0	0	0	0	0	0	0	0	0	0
2017	0	0	43	409	99	83	26	2	0	0	0	0	0	0	0	0	0	0

**Table 3.14.** Predicted harvest at age (numbers of fish) for the RRrec fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	0	0	150	11,196	9,646	4,067	1,353	413	90	14	2	0	0	0	0	0	0	0
1992	0	0	35	2,402	8,825	1,683	323	77	20	4	0	0	0	0	0	0	0	0
1993	0	0	41	1,851	6,293	5,551	509	69	13	3	0	0	0	0	0	0	0	0
1994	0	0	47	1,449	3,186	2,491	1,031	67	7	1	0	0	0	0	0	0	0	0
1995	0	0	134	2,078	3,055	1,498	537	158	9	1	0	0	0	0	0	0	0	0
1996	0	0	154	4,022	2,971	950	207	52	13	1	0	0	0	0	0	0	0	0
1997	0	0	64	3,609	4,745	805	115	17	4	1	0	0	0	0	0	0	0	0
1998	0	0	221	4,628	13,454	4,405	361	36	4	1	0	0	0	0	0	0	0	0
1999	0	0	89	7,427	8,085	5,888	934	53	4	0	0	0	0	0	0	0	0	0
2000	0	0	202	5,501	23,526	6,421	2,294	254	12	1	0	0	0	0	0	0	0	0
2001	0	0	94	8,769	11,985	12,336	1,607	401	36	1	0	0	0	0	0	0	0	0
2002	0	0	338	4,512	20,998	6,843	3,355	307	62	5	0	0	0	0	0	0	0	0
2003	0	0	35	4,297	2,919	3,227	496	172	13	2	0	0	0	0	0	0	0	0
2004	0	0	50	2,987	18,583	3,060	1,607	172	48	3	0	0	0	0	0	0	0	0
2005	0	0	39	3,958	11,518	16,758	1,306	486	43	10	1	0	0	0	0	0	0	0
2006	0	0	131	2,306	10,811	6,941	4,797	277	88	7	1	0	0	0	0	0	0	0
2007	0	0	470	7,232	5,037	4,490	1,315	716	38	11	1	0	0	0	0	0	0	0
2008	0	0	102	5,843	3,936	458	150	33	17	1	0	0	0	0	0	0	0	0
2009	0	0	144	5,561	15,168	2,229	115	26	5	2	0	0	0	0	0	0	0	0
2010	0	0	60	5,631	10,168	6,147	425	16	3	0	0	0	0	0	0	0	0	0
2011	0	0	20	2,975	12,797	4,907	1,329	67	2	0	0	0	0	0	0	0	0	0
2012	0	0	376	2,545	13,113	10,458	1,823	378	17	1	0	0	0	0	0	0	0	0
2013	0	0	281	5,284	1,206	725	180	25	6	0	0	0	0	0	0	0	0	0
2014	0	0	67	5,976	4,805	171	34	6	1	0	0	0	0	0	0	0	0	0
2015	0	0	29	3,628	14,074	2,258	35	5	1	0	0	0	0	0	0	0	0	0
2016	0	0	244	2,061	10,685	7,749	524	6	1	0	0	0	0	0	0	0	0	0
2017	0	0	146	5,436	1,952	1,804	535	28	0	0	0	0	0	0	0	0	0	0

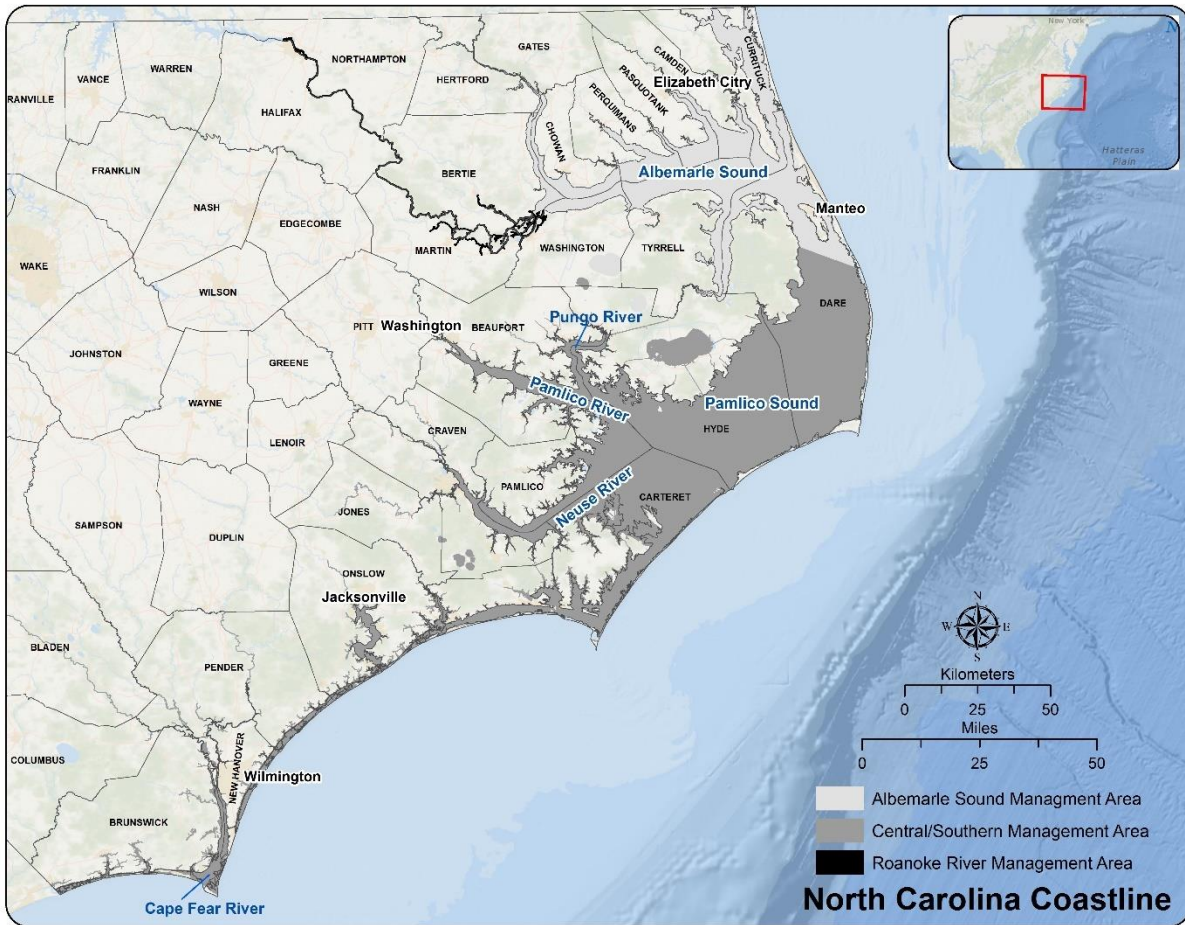
**Table 3.15.** Predicted dead discards at age (numbers of fish) for the RRrec fleet from the base run of the stock assessment model, 1991–2017.

Year	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17
1991	0	7	446	3,809	2,058	1,043	624	470	327	214	143	115	87	58	40	28	19	29
1992	0	3	132	1,034	2,383	546	189	112	93	69	47	32	26	20	13	9	6	11
1993	0	5	153	789	1,683	1,782	292	98	62	54	42	29	20	16	12	8	6	11
1994	0	11	156	551	760	713	529	85	31	21	19	15	10	7	6	4	3	6
1995	0	20	505	895	825	486	312	226	40	16	11	10	8	6	4	3	2	5
1996	0	31	1,636	4,868	2,255	865	338	210	168	32	13	10	9	7	5	4	3	7
1997	0	65	1,288	8,341	6,878	1,400	359	132	88	76	15	6	5	4	4	3	2	5
1998	0	16	1,235	2,951	5,381	2,116	310	75	29	20	18	4	2	1	1	1	1	2
1999	0	16	421	4,036	2,756	2,410	685	95	24	10	7	6	1	1	0	0	0	1
2000	0	4	339	1,057	2,836	930	596	162	24	6	3	2	2	0	0	0	0	0
2001	0	10	123	1,309	1,122	1,387	324	199	57	9	2	1	1	1	0	0	0	0
2002	0	3	327	499	1,456	570	501	113	74	22	4	1	0	0	0	0	0	0
2003	0	1	72	1,013	432	573	158	134	33	23	7	1	0	0	0	0	0	0
2004	0	3	250	1,713	6,684	1,321	1,243	327	296	76	55	18	3	1	0	0	0	0
2005	0	7	119	1,393	2,542	4,440	620	567	161	156	42	31	10	2	0	0	0	0
2006	0	13	195	393	1,155	890	1,103	157	161	50	51	14	10	3	1	0	0	0
2007	0	11	590	1,036	453	484	254	342	59	70	23	25	7	5	2	0	0	0
2008	0	29	1,060	6,951	2,937	409	239	129	221	46	60	21	23	6	5	2	0	0
2009	0	7	592	2,618	4,480	789	73	41	24	44	10	13	5	5	1	1	0	0
2010	0	2	234	2,492	2,823	2,047	253	23	14	9	18	4	5	2	2	1	0	0
2011	0	10	72	1,206	3,255	1,497	726	90	9	7	4	9	2	3	1	1	0	0
2012	0	26	507	392	1,266	1,211	378	193	28	3	2	2	4	1	1	0	0	0
2013	0	14	1,231	2,646	379	272	121	42	30	6	1	1	0	1	0	0	0	0
2014	0	5	632	6,463	3,260	139	49	21	10	9	2	0	0	0	0	0	0	0
2015	0	14	120	1,731	4,213	810	22	8	4	2	2	0	0	0	0	0	0	0
2016	0	11	410	396	1,289	1,121	135	4	1	1	0	0	0	0	0	0	0	0
2017	0	11	634	2,693	607	672	356	45	2	1	0	0	0	0	0	0	0	0

**Table 3.16.** Annual estimates of fishing mortality (numbers-weighted, ages 3–5) and associated standard deviations (SDs) from the base run of the stock assessment model, 1991–2017.

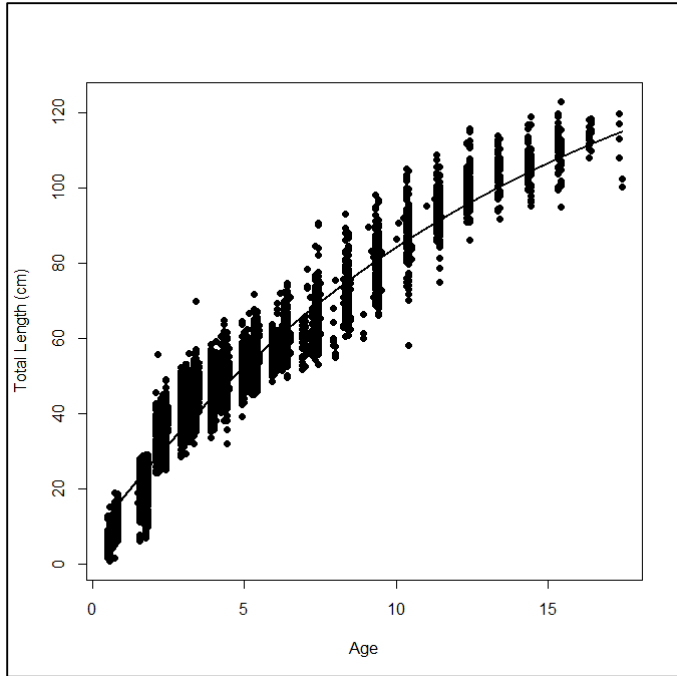
<b>Year</b>	<b>Fishing Mortality</b>	
	<b>Value</b>	<b>SD</b>
<b>1991</b>	0.25	0.015
<b>1992</b>	0.23	0.012
<b>1993</b>	0.35	0.021
<b>1994</b>	0.32	0.020
<b>1995</b>	0.28	0.019
<b>1996</b>	0.20	0.012
<b>1997</b>	0.15	0.0082
<b>1998</b>	0.21	0.012
<b>1999</b>	0.15	0.0071
<b>2000</b>	0.26	0.013
<b>2001</b>	0.24	0.012
<b>2002</b>	0.29	0.017
<b>2003</b>	0.15	0.0066
<b>2004</b>	0.30	0.0099
<b>2005</b>	0.42	0.011
<b>2006</b>	0.52	0.026
<b>2007</b>	0.48	0.030
<b>2008</b>	0.21	0.013
<b>2009</b>	0.28	0.015
<b>2010</b>	0.34	0.0094
<b>2011</b>	0.44	0.010
<b>2012</b>	1.3	0.057
<b>2013</b>	0.35	0.023
<b>2014</b>	0.23	0.0091
<b>2015</b>	0.50	0.017
<b>2016</b>	0.75	0.045
<b>2017</b>	0.27	0.025

9 FIGURES

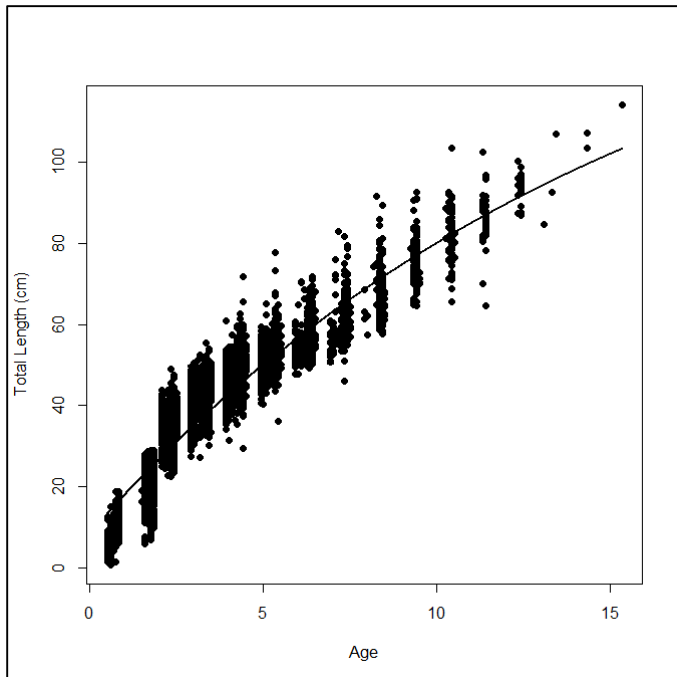


**Figure 1.1.** Boundary lines defining the Albemarle Sound Management Area, Central-Southern Management Area, and the Roanoke River Management Area.

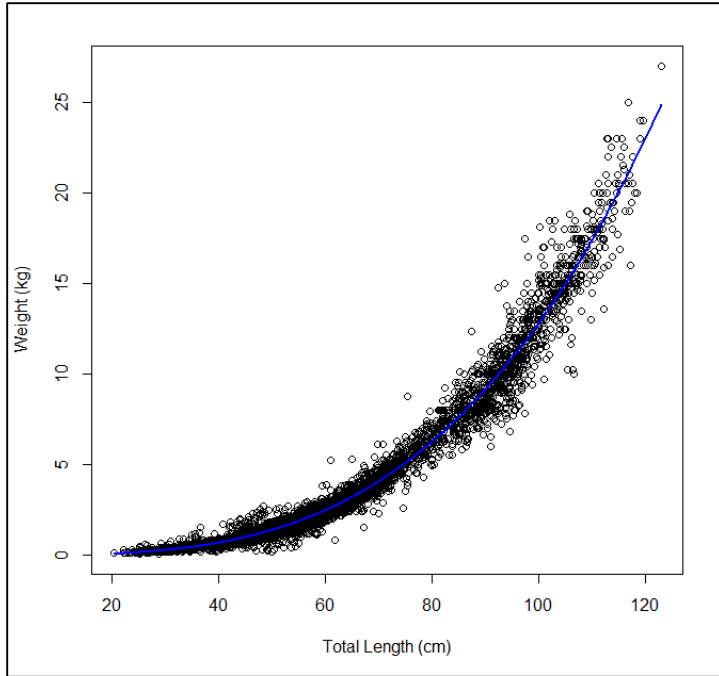




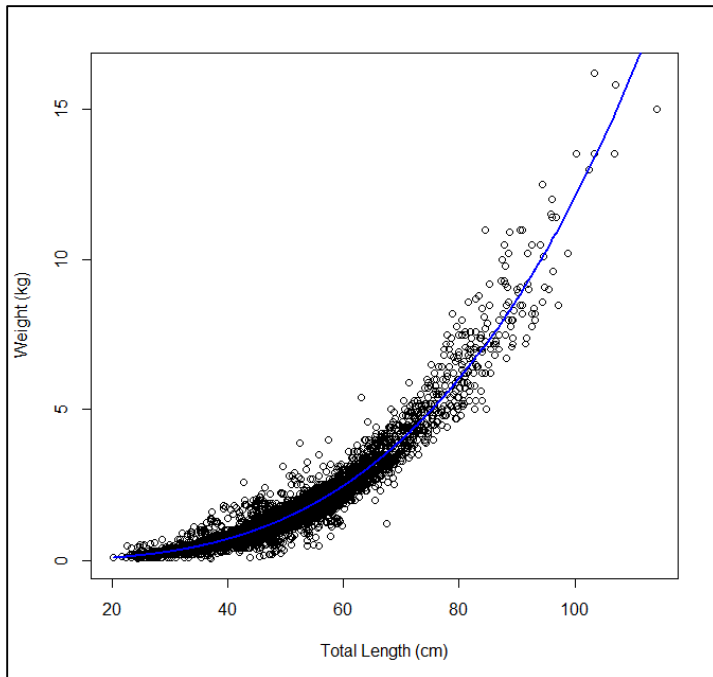
**Figure 1.2.** Fit of the age-length function to available age data for female striped bass.



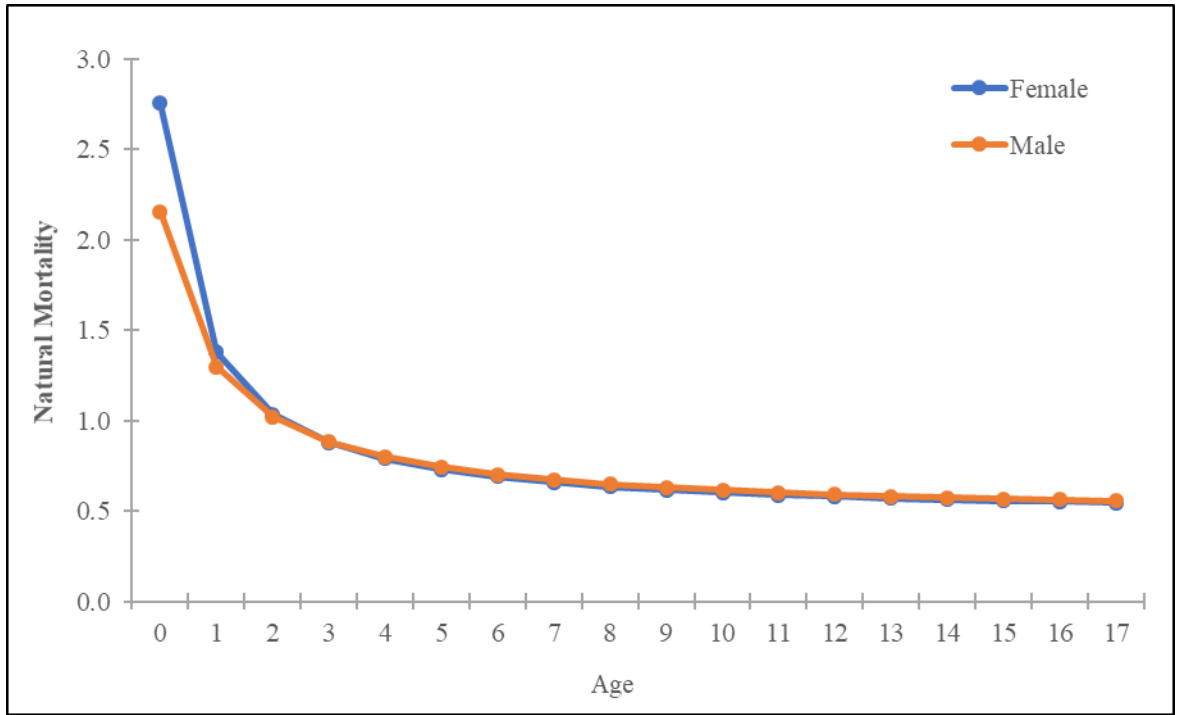
**Figure 1.3.** Fit of the age-length function to available age data for male striped bass.



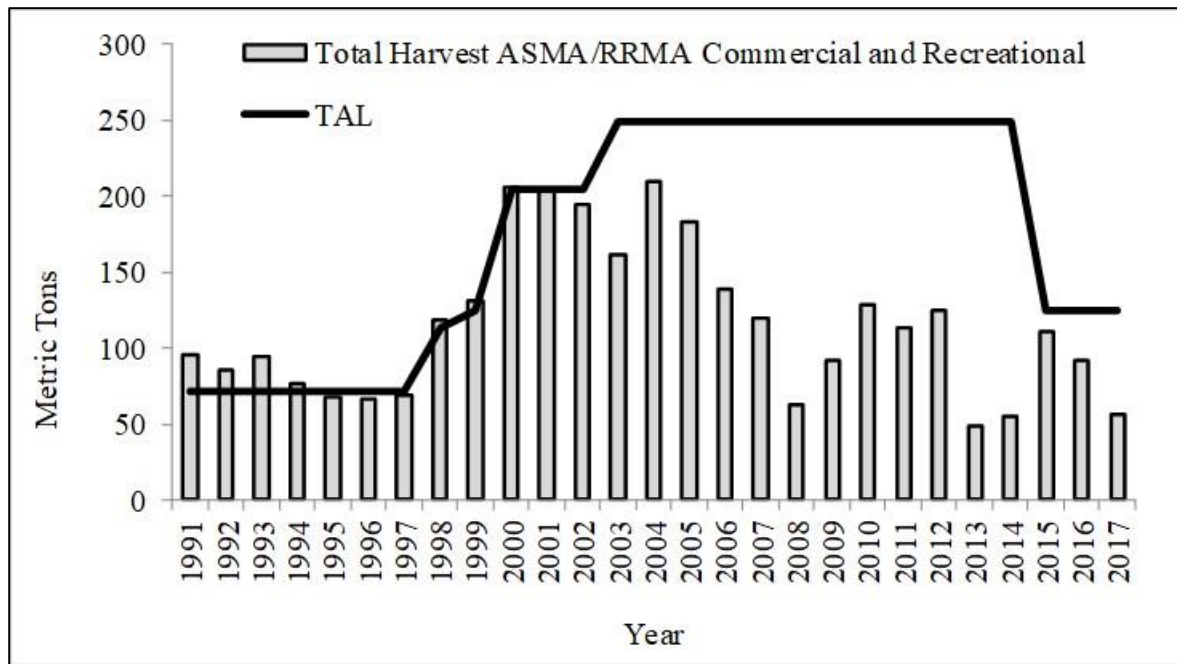
**Figure 1.4.** Fit of the length-weight function to available biological data for female striped bass.



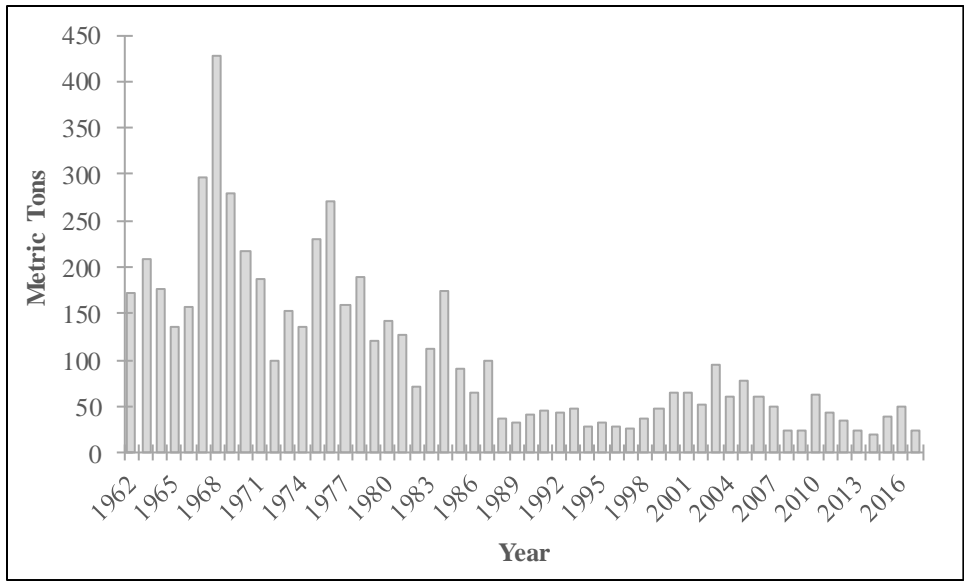
**Figure 1.5.** Fit of the length-weight function to available biological data for male striped bass.



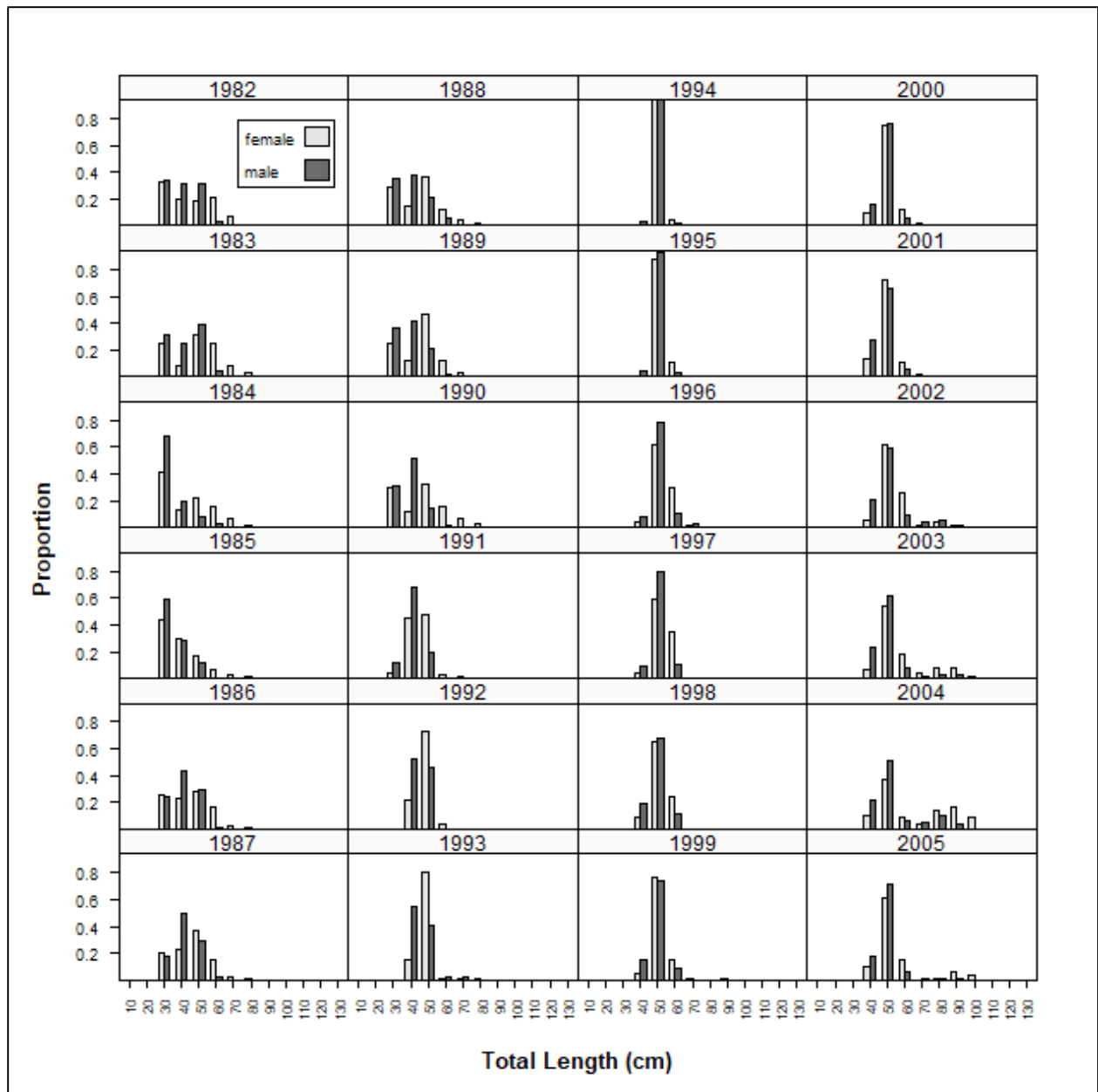
**Figure 1.6.** Estimates of natural mortality at age based on the method of Lorenzen (1996).



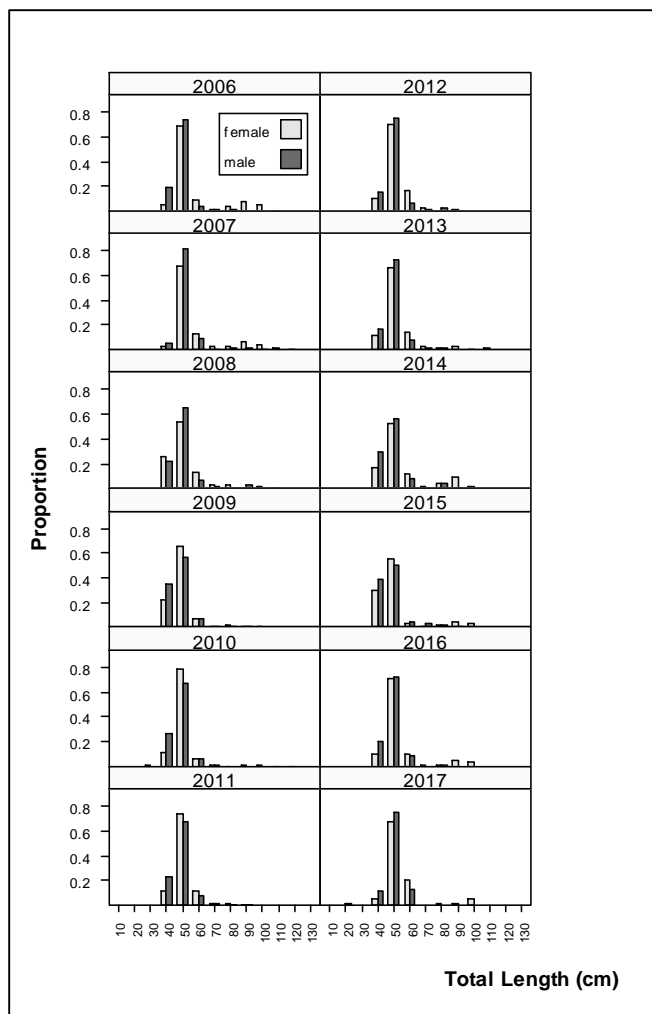
**Figure 1.7.** Annual total landings/harvest in metric tons of striped bass from the ASMA and RRMA commercial and recreational sectors combined compared to the TAL, 1991–2017.



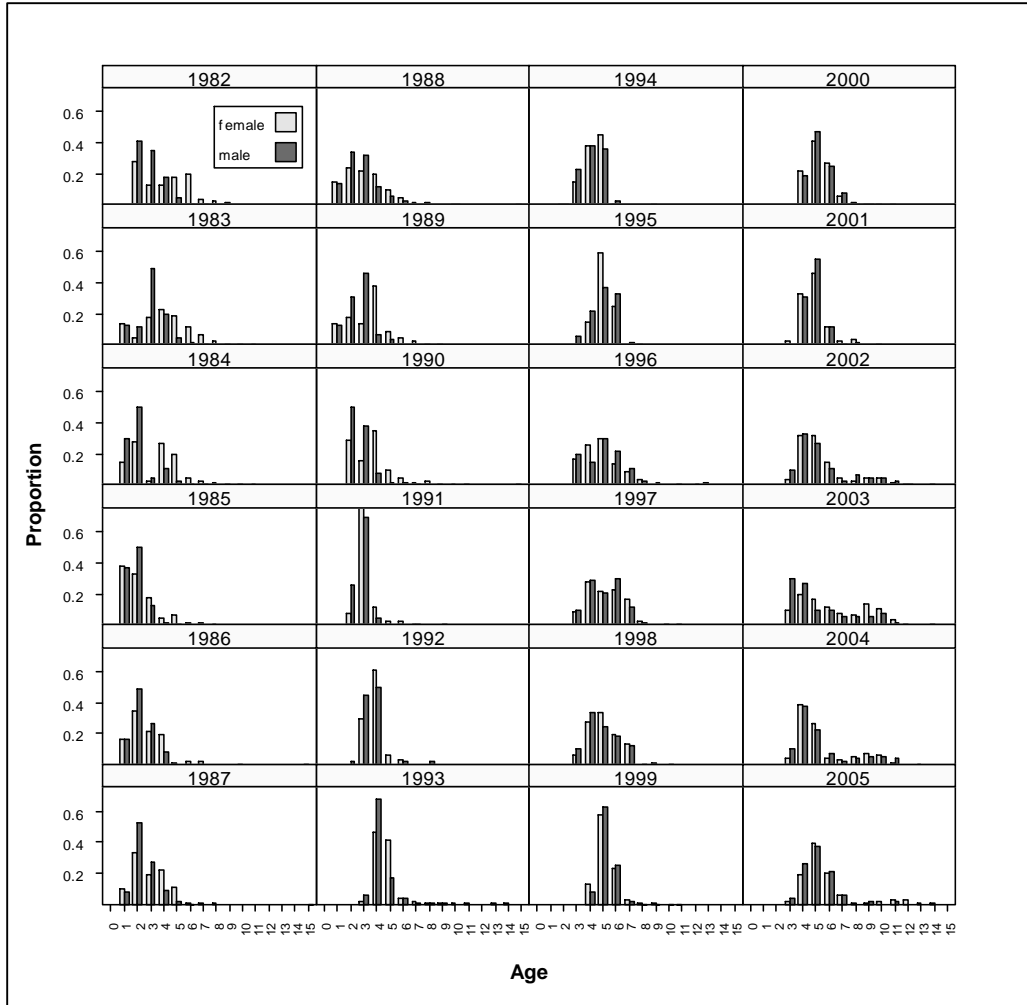
**Figure 2.1.** Annual commercial landings of striped bass in the ASMA-RRMA, 1962–2017.



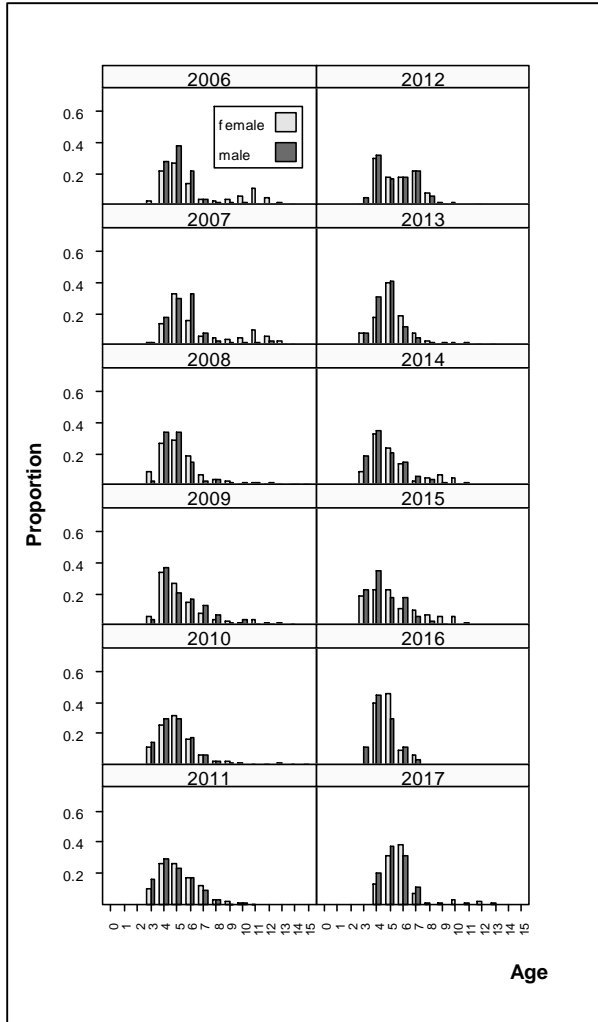
**Figure 2.2.** Annual length frequencies of striped bass commercial landings, 1982–2005.



**Figure 2.3.** Annual length frequencies of striped bass commercial landings, 2006–2017.



**Figure 2.4.** Annual age frequencies of striped bass commercial landings, 1982–2005. The age-15 bin represents a plus group.



**Figure 2.5.** Annual age frequencies of striped bass commercial landings, 2006–2017. The age-15 bin represents a plus group.



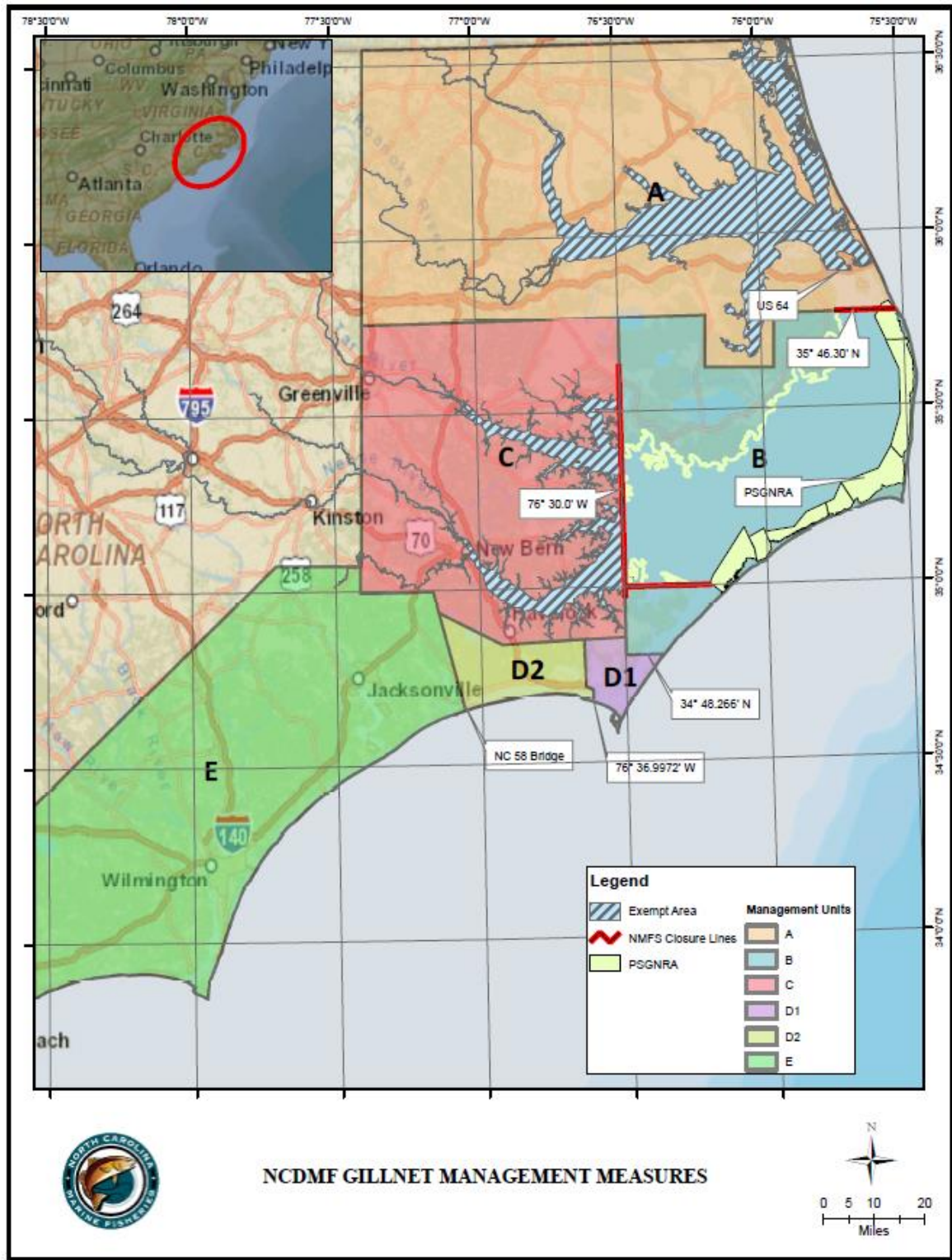
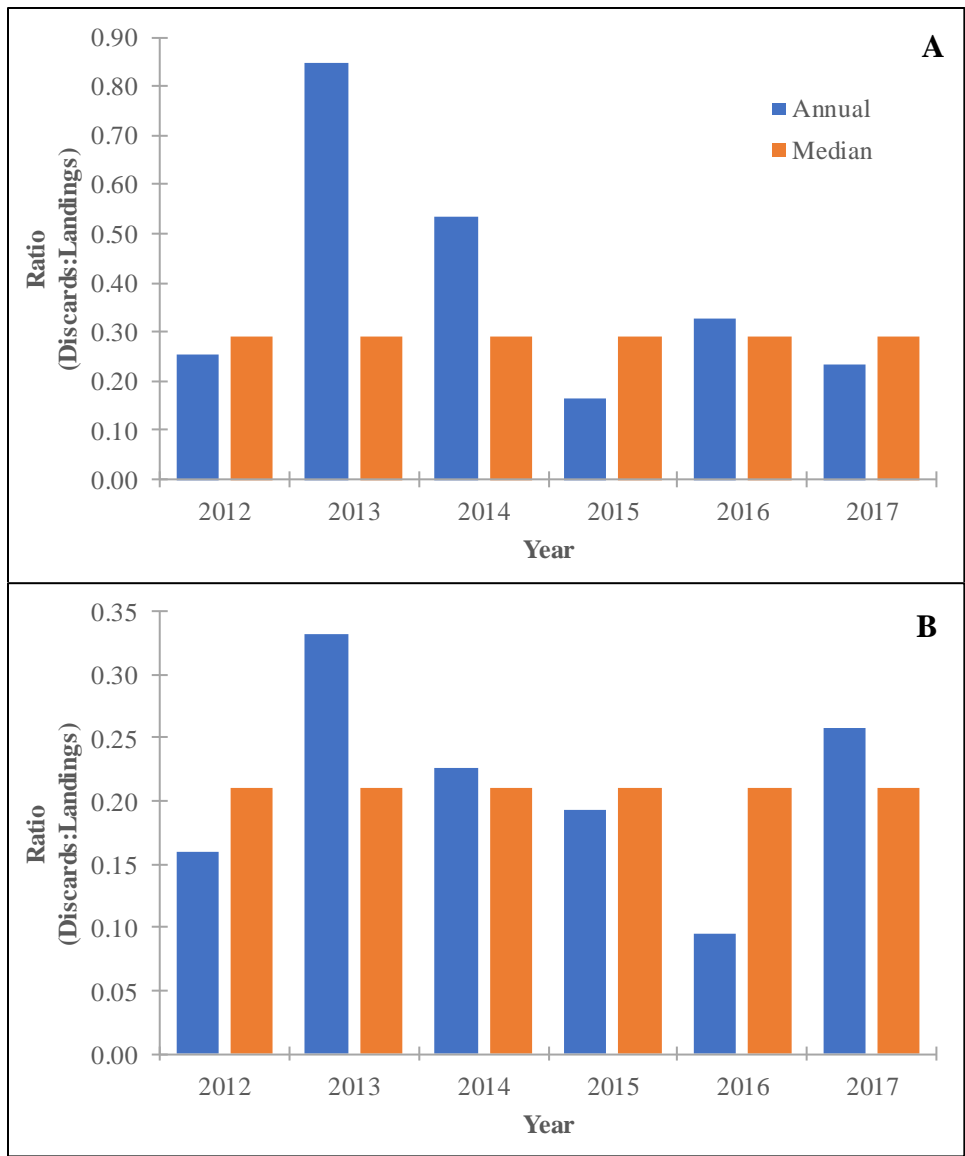
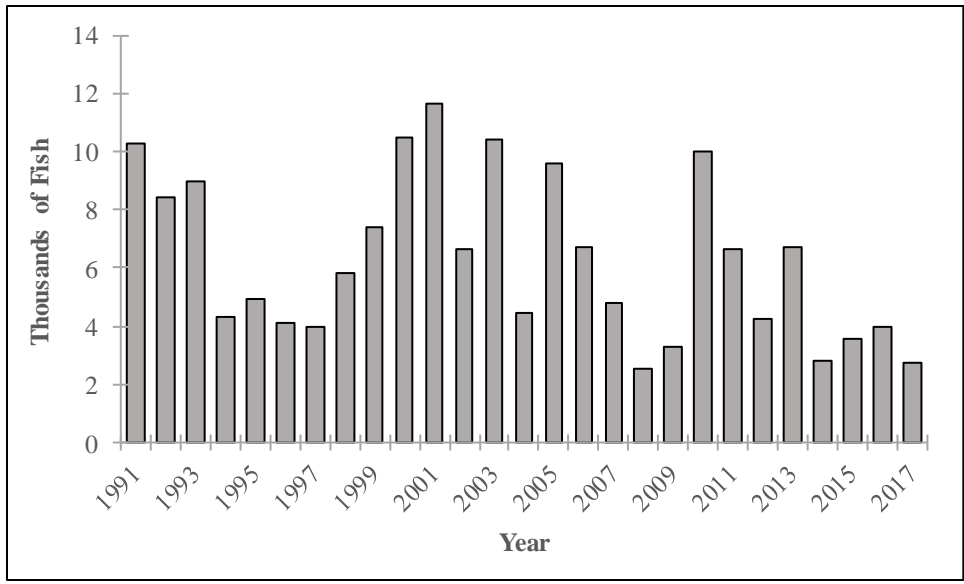


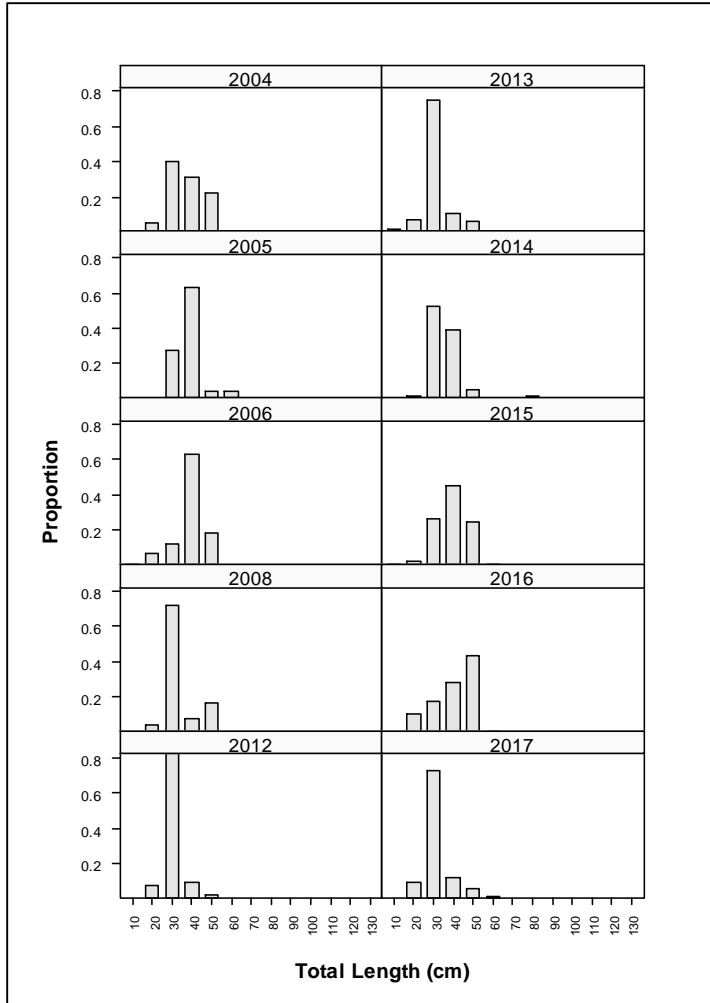
Figure 2.6. Management areas used in development of GLM for commercial gill-net discards.



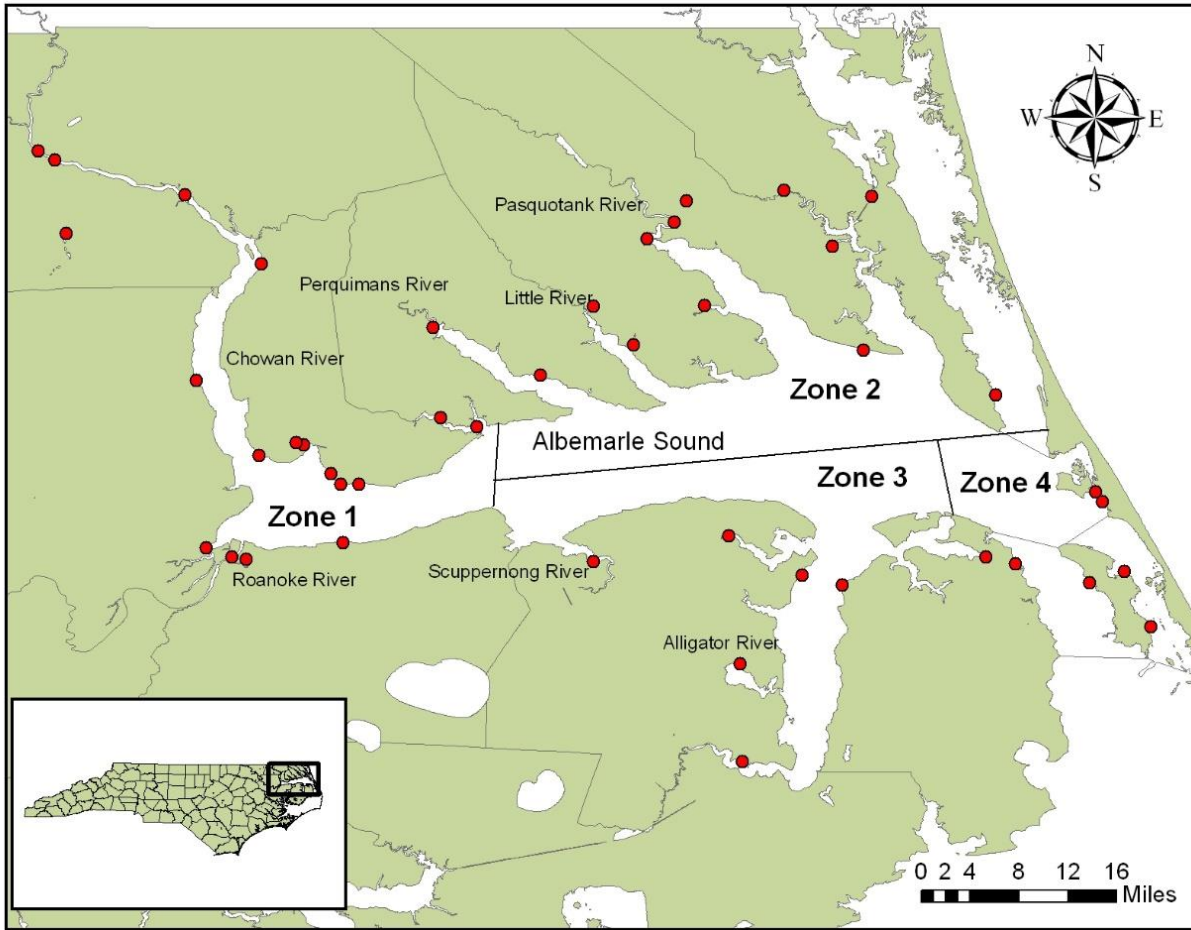
**Figure 2.7.** Ratio of commercial (A) live and (B) dead discards to commercial landings, 2012–2017.



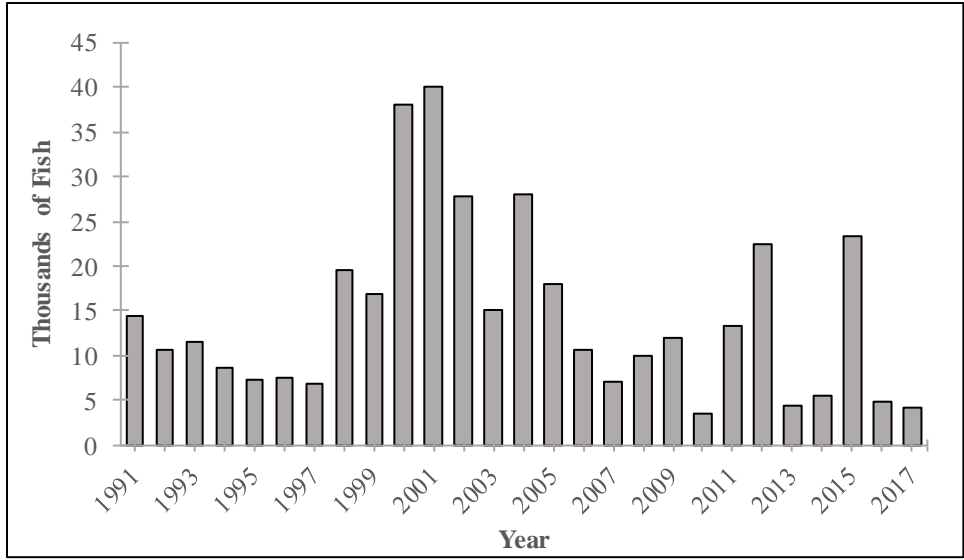
**Figure 2.8.** Annual estimates of commercial gill-net discards, 1991–2017. Note that values prior to 2012 were estimated using a hindcasting approach.



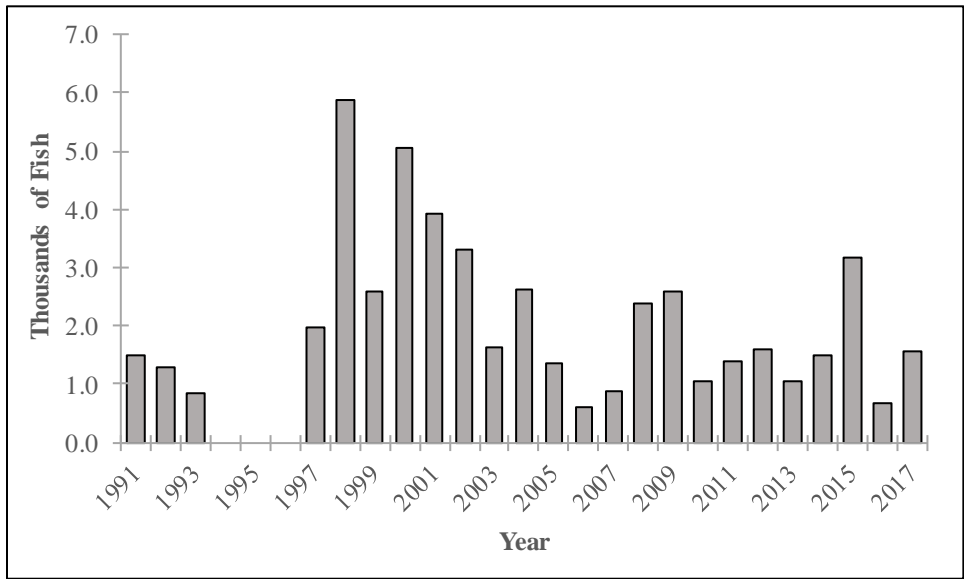
**Figure 2.9.** Annual length frequencies of striped bass commercial gill-net discards, 2004–2017.



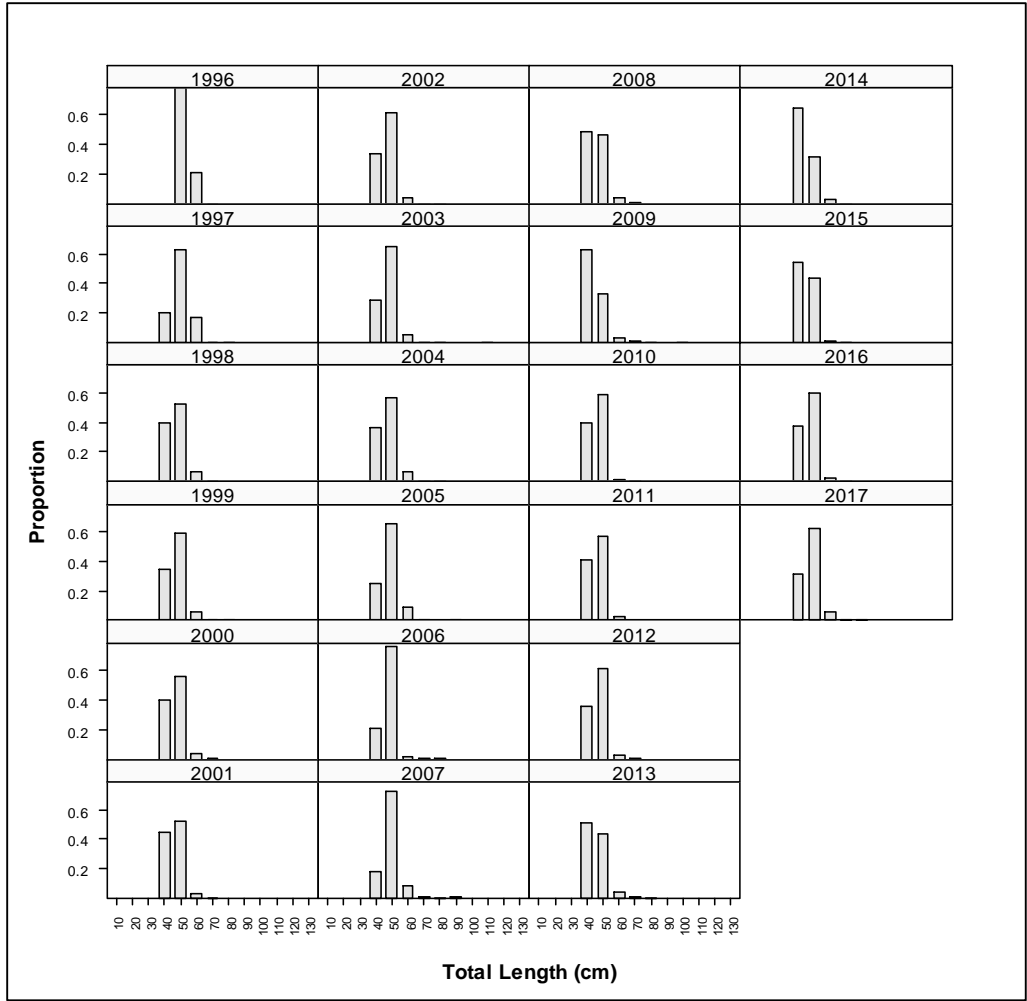
**Figure 2.10.** Sampling zones and access sites of the striped bass recreational creel survey in the ASMA.



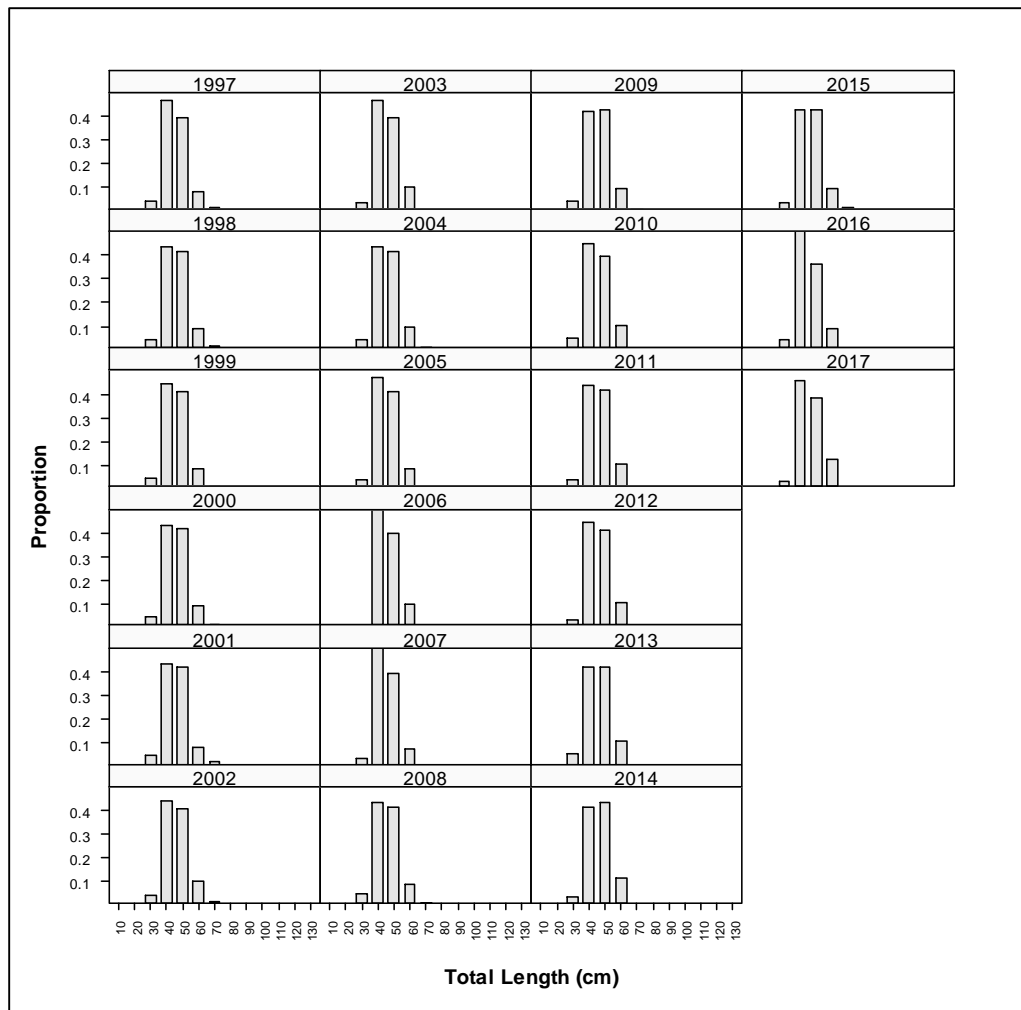
**Figure 2.11.** Annual estimates of recreational harvest for the Albemarle Sound, 1991–2017.



**Figure 2.12.** Annual estimates of recreational dead discards for the Albemarle Sound, 1991–2017.

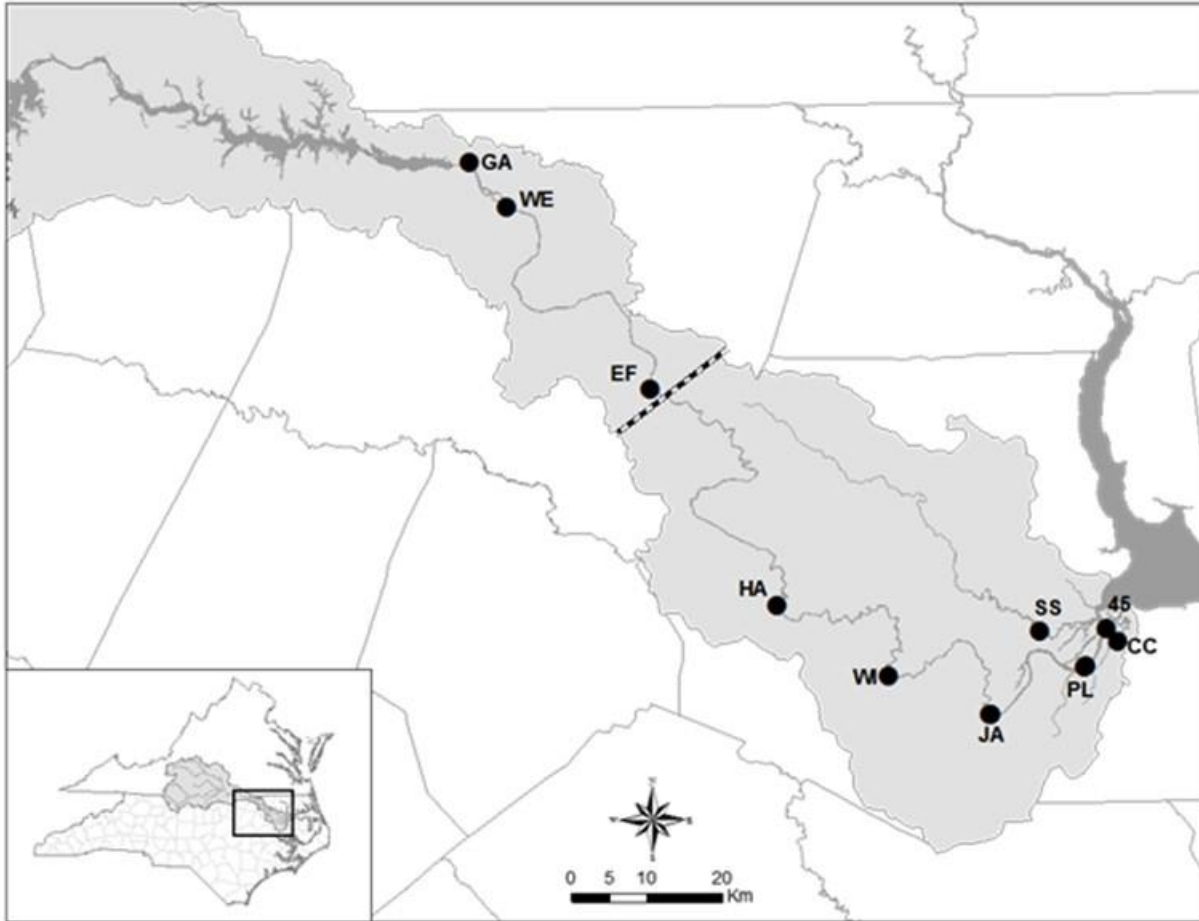


**Figure 2.13.** Annual length frequencies of striped bass recreational harvest in the Albemarle Sound, 1996–2017.

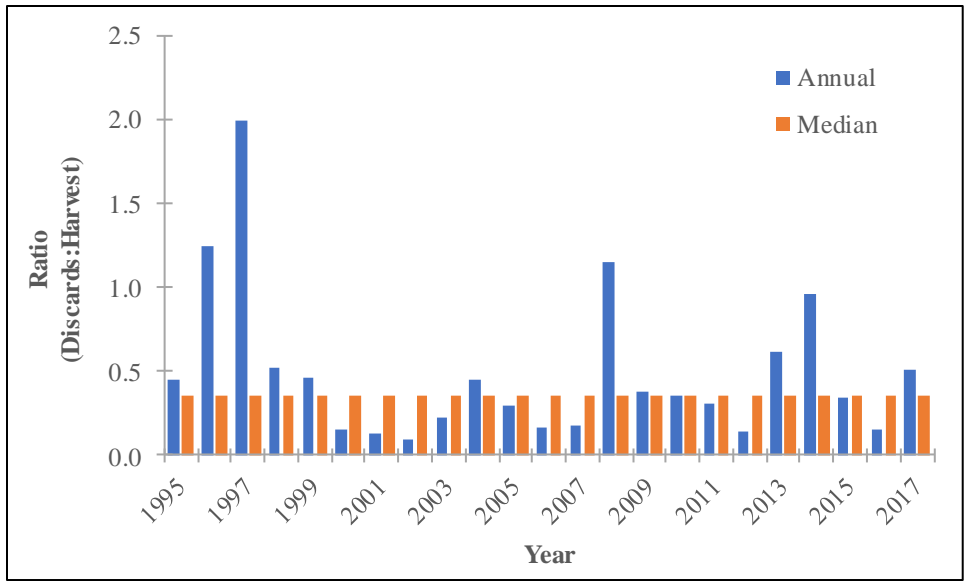


**Figure 2.14.** Annual length frequencies of striped bass recreational discards in the Albemarle Sound, 1997–2017.

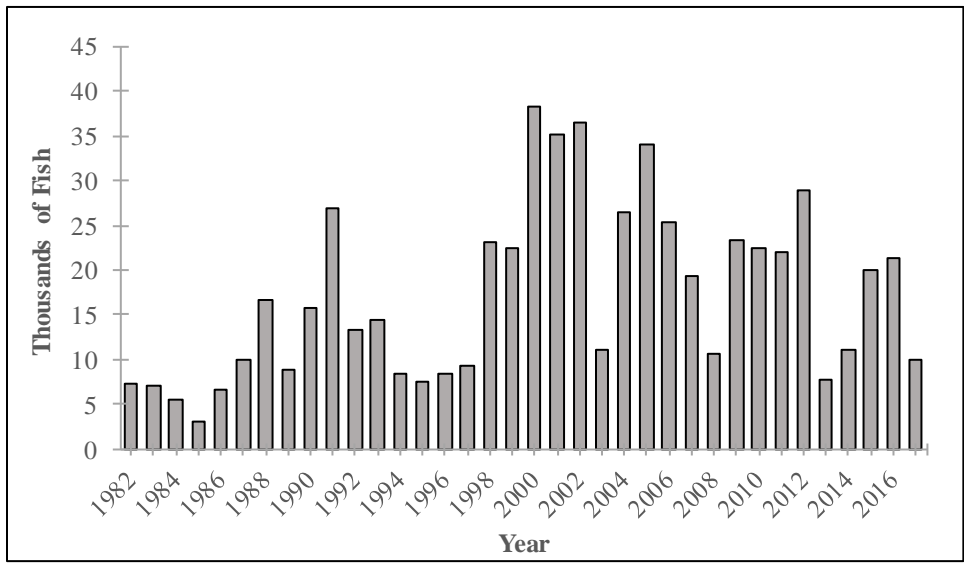




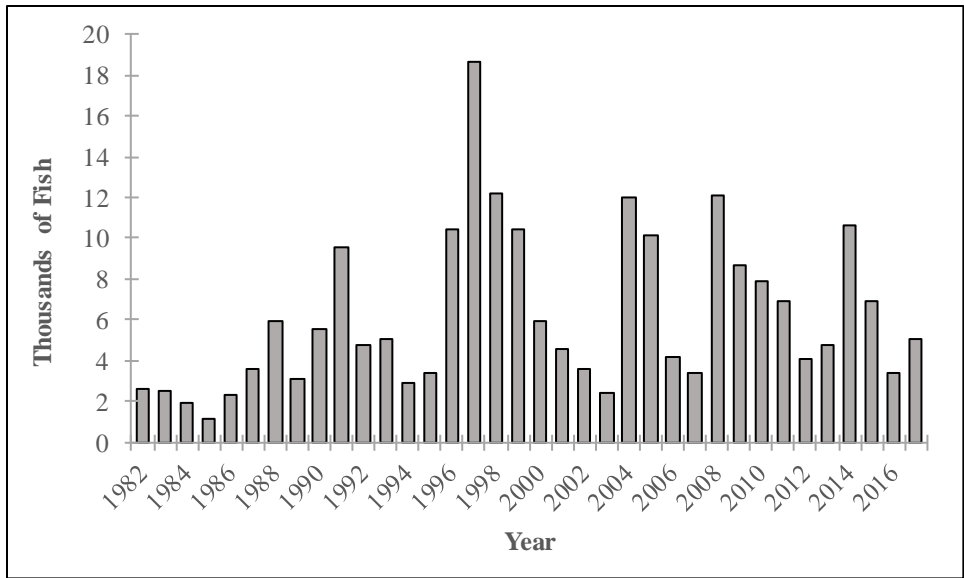
**Figure 2.15.** Map of angler creel survey interview locations in the RRMA, NC. The dashed line indicates the demarcation point between the upper and lower zones. Zone 1 access areas include (GA) Gaston (US HWY 48), (WE) Weldon, and (EF) Scotland Neck (Edwards Ferry US HWY 258). Zone 2 access areas include (HA) Hamilton, (WI) Williamston, (JA) Jamesville, (PL) Plymouth, (45) US HWY 45, (CC) Conaby Creek, and (SS) Sans Souci (Cashie River).



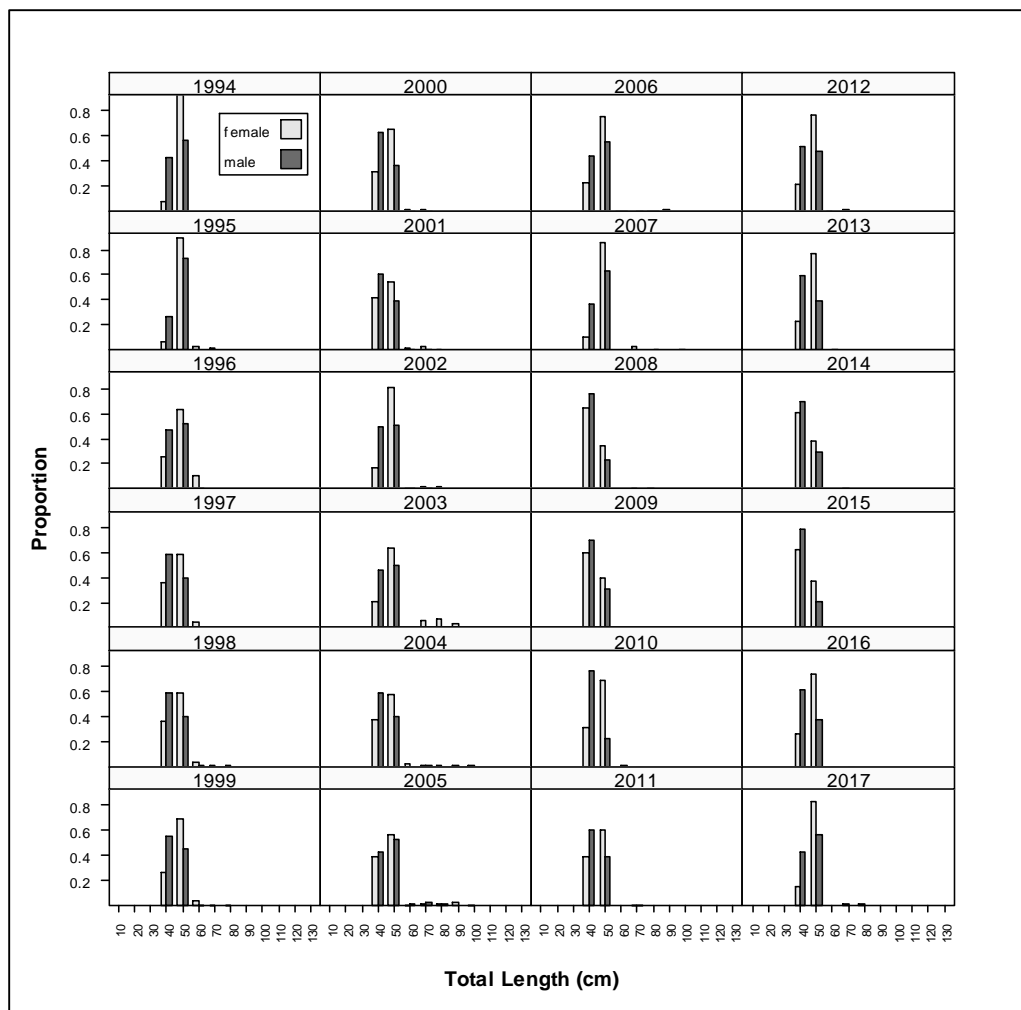
**Figure 2.16.** Ratio of recreational dead discards to recreational harvest in the Roanoke River, 1995–2017.



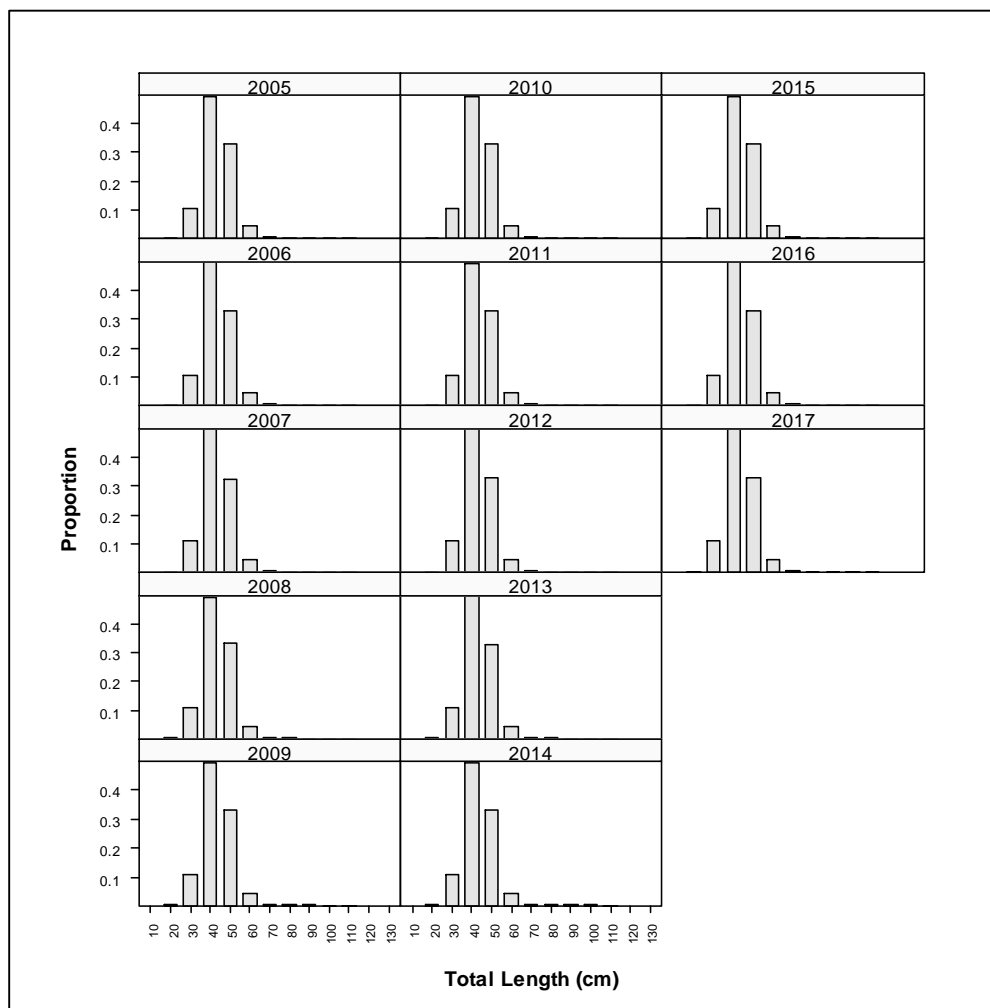
**Figure 2.17.** Annual estimates of recreational harvest for the Roanoke River, 1982–2017.



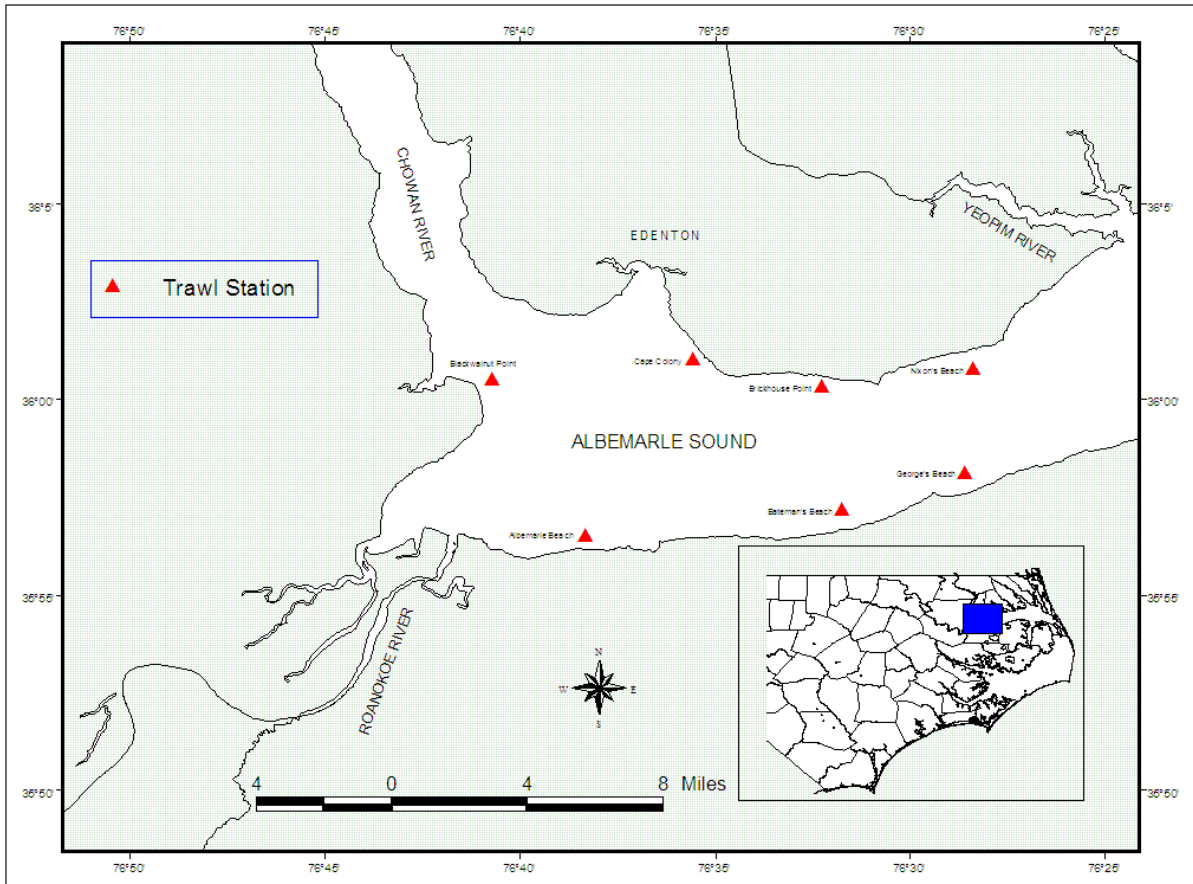
**Figure 2.18.** Annual estimates of recreational dead discards for the Roanoke River, 1982–2017. Note that discard values prior to 1995 were estimated using a hindcasting approach.



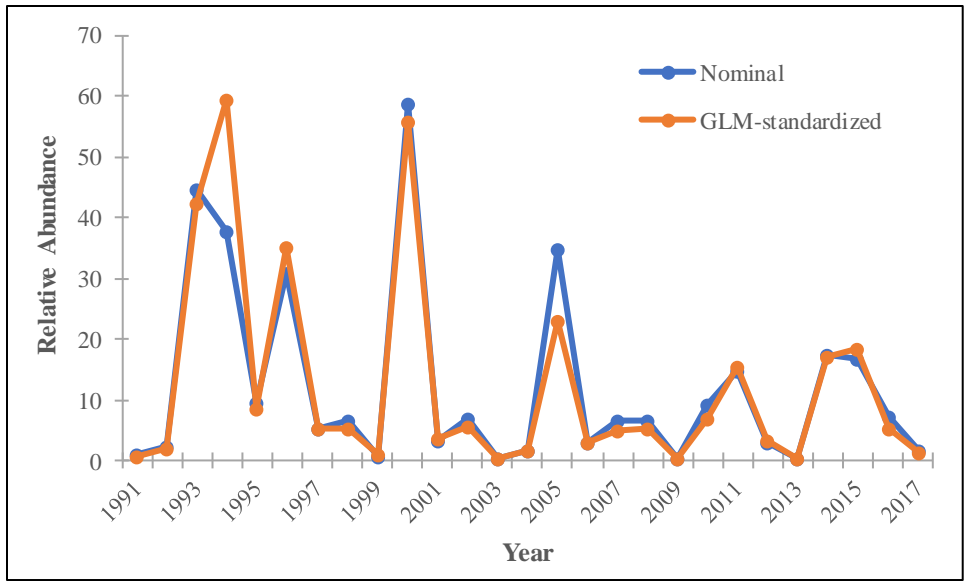
**Figure 2.19.** Annual length frequencies of striped bass recreational harvest in the Roanoke River, 1994–2017.



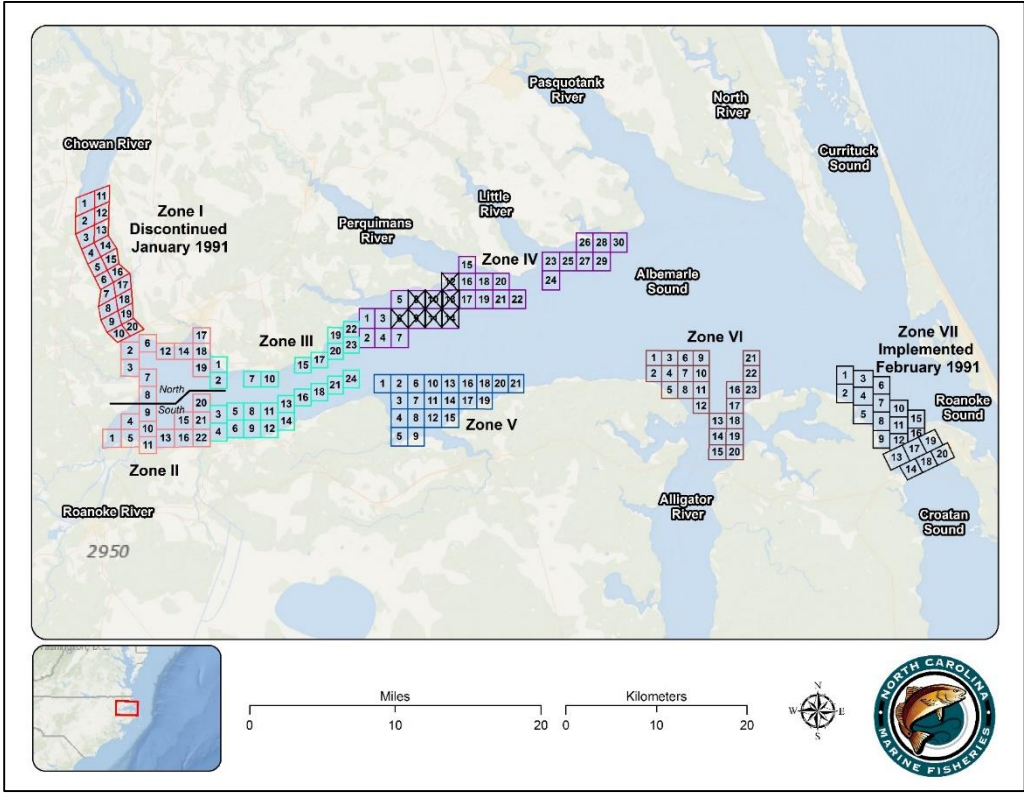
**Figure 2.20.** Annual length frequencies of striped bass recreational discards in the Roanoke River, 2005–2017.



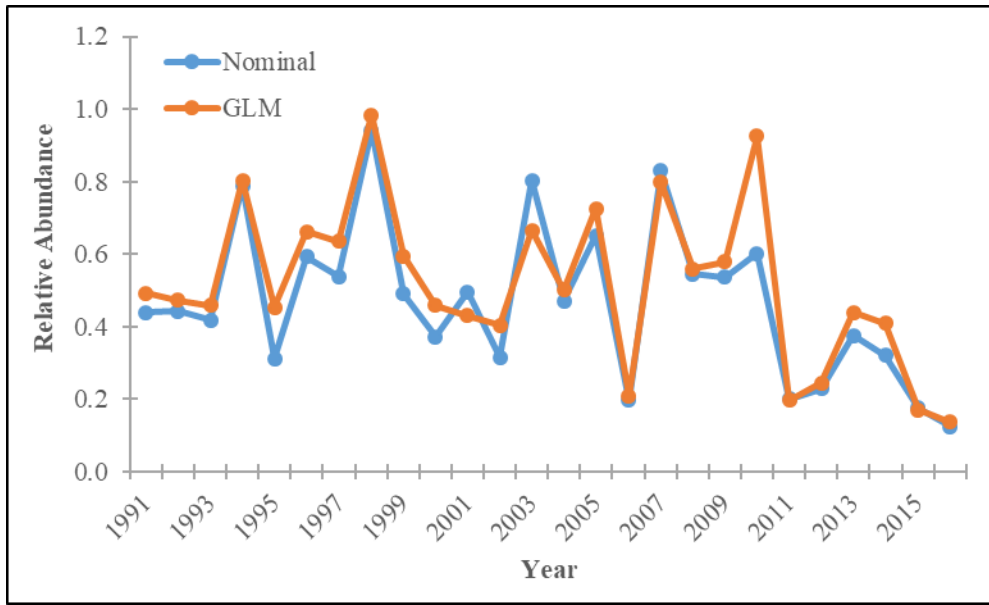
**Figure 2.21.** Map of NCDMF Juvenile Abundance Survey (Program 100) sampling sites.



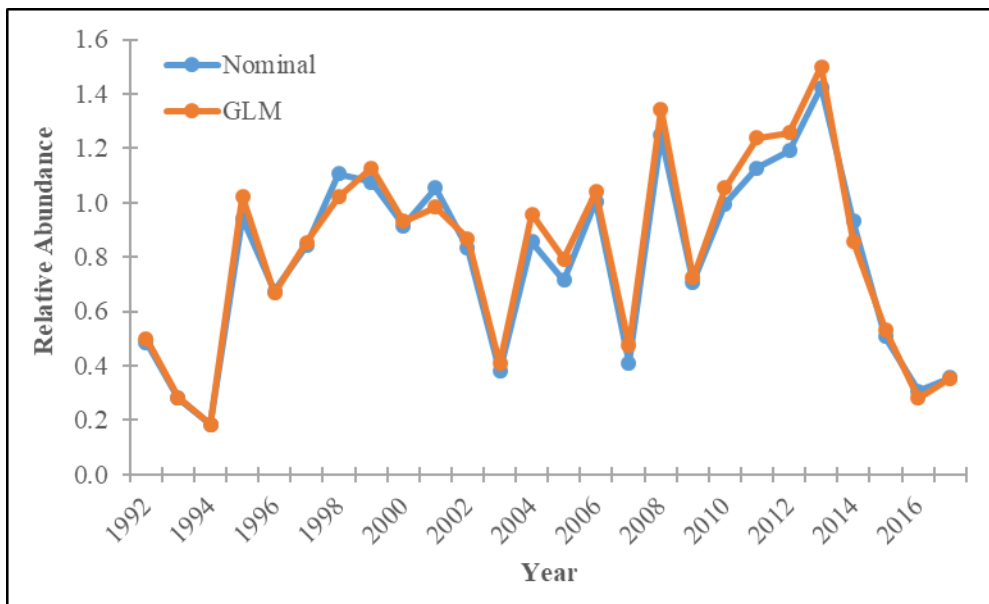
**Figure 2.22.** Nominal and GLM-standardized indices of relative age-0 abundance derived from the Juvenile Abundance Survey (P100), 1991–2017.



**Figure 2.23.** Map of sampling grids and zones for the NCDMF Independent Gill-Net Survey (Program 135).

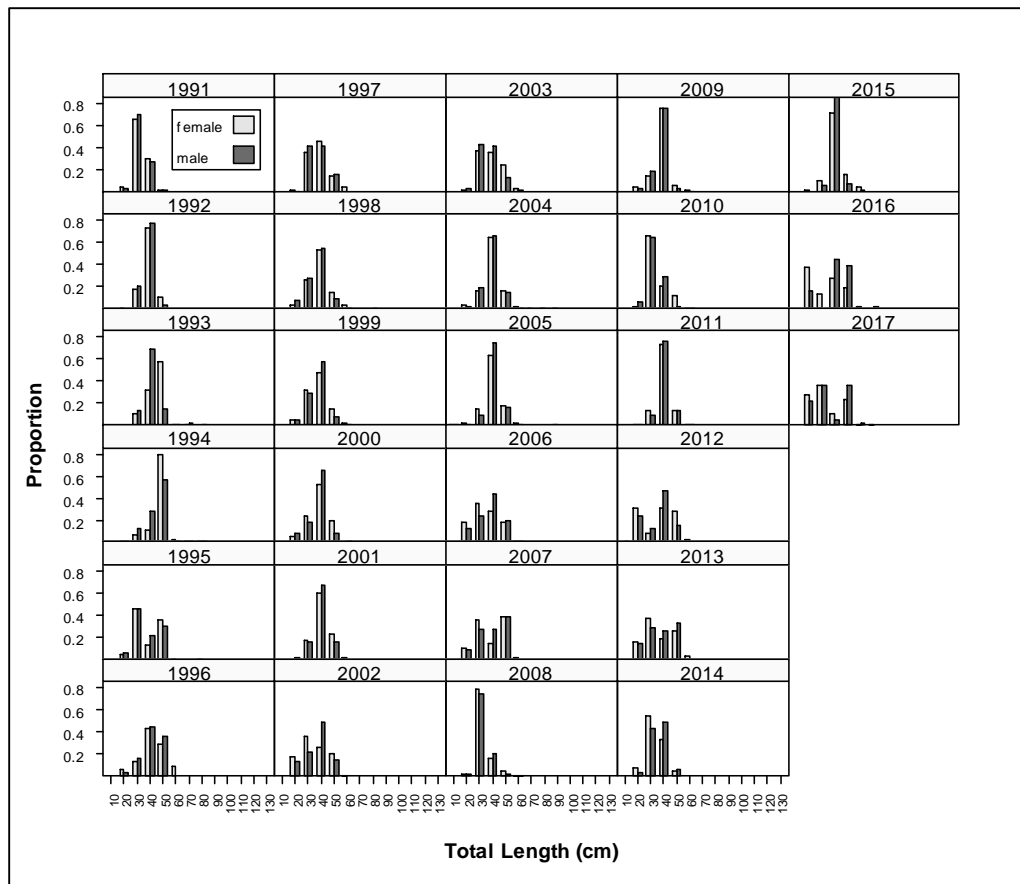


**Figure 2.24.** Nominal and GLM-standardized indices of relative abundance derived from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2016.

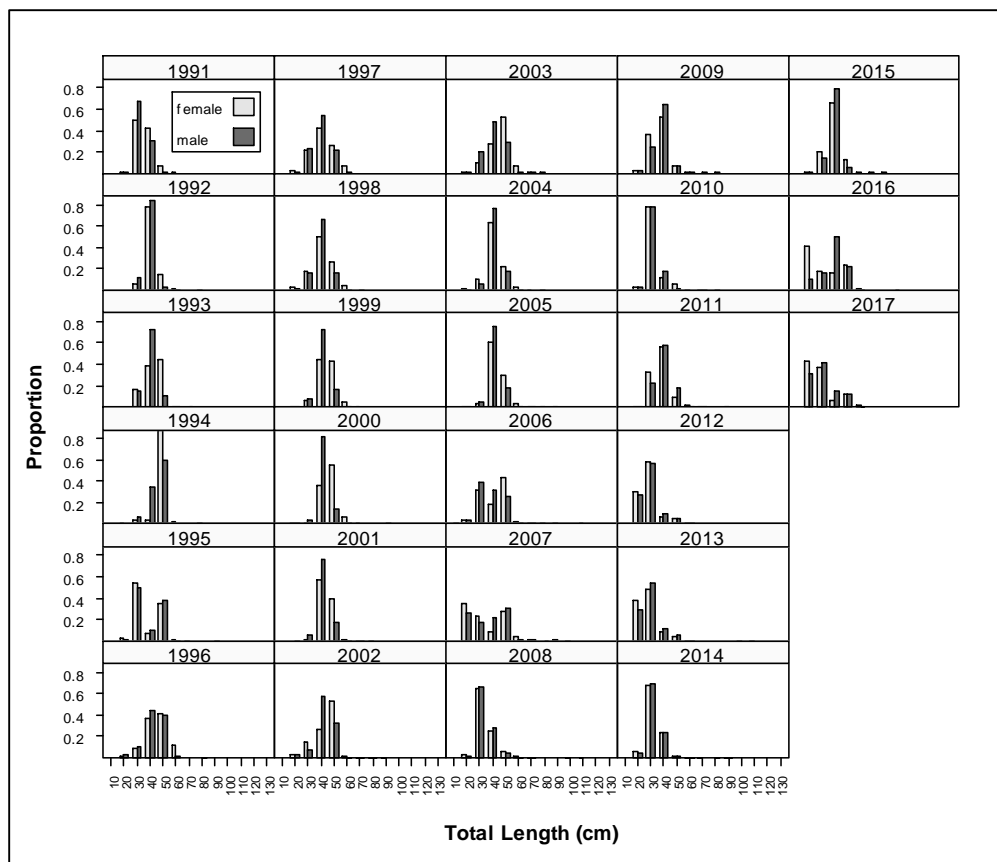


**Figure 2.25.** Nominal and GLM-standardized indices of relative abundance derived from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1992–2017.

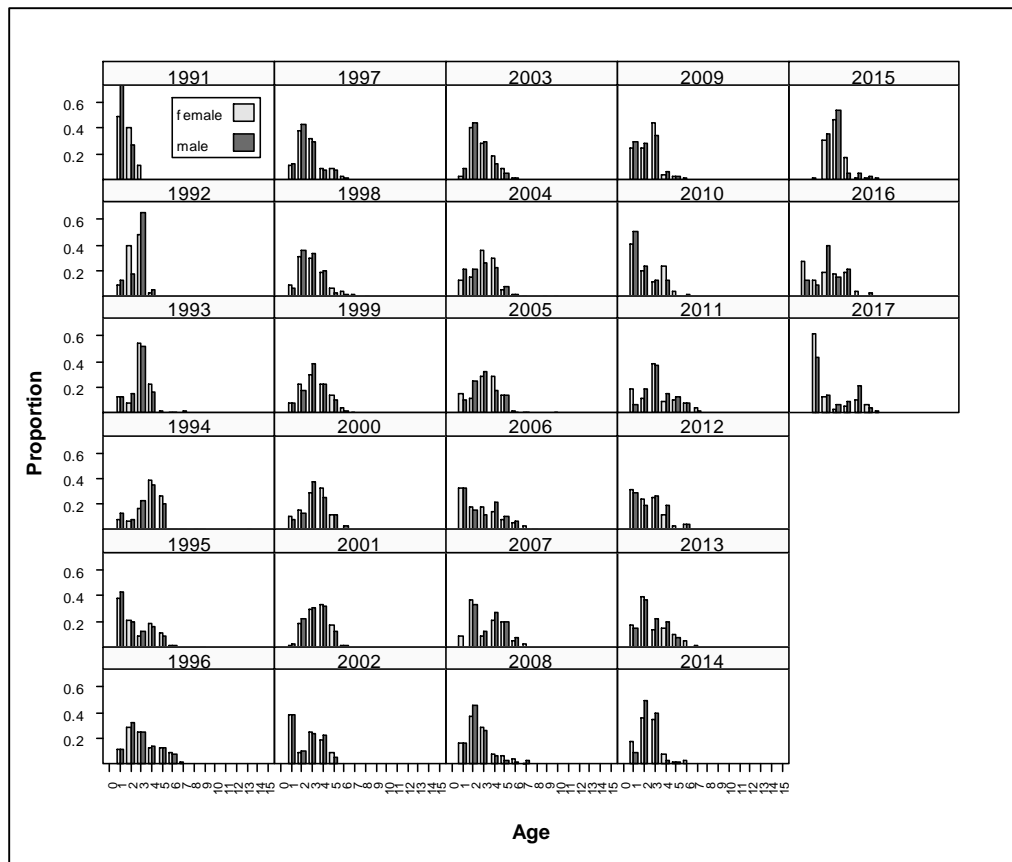




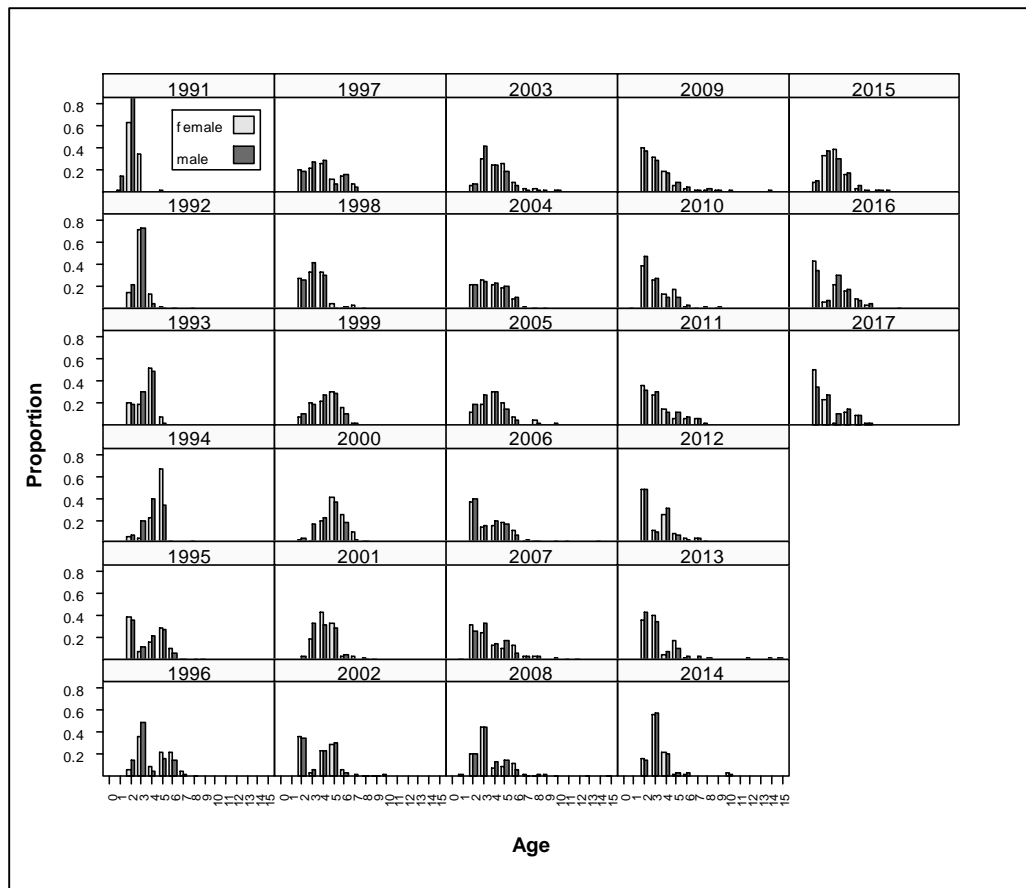
**Figure 2.26.** Annual length frequencies of striped bass sampled from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017.



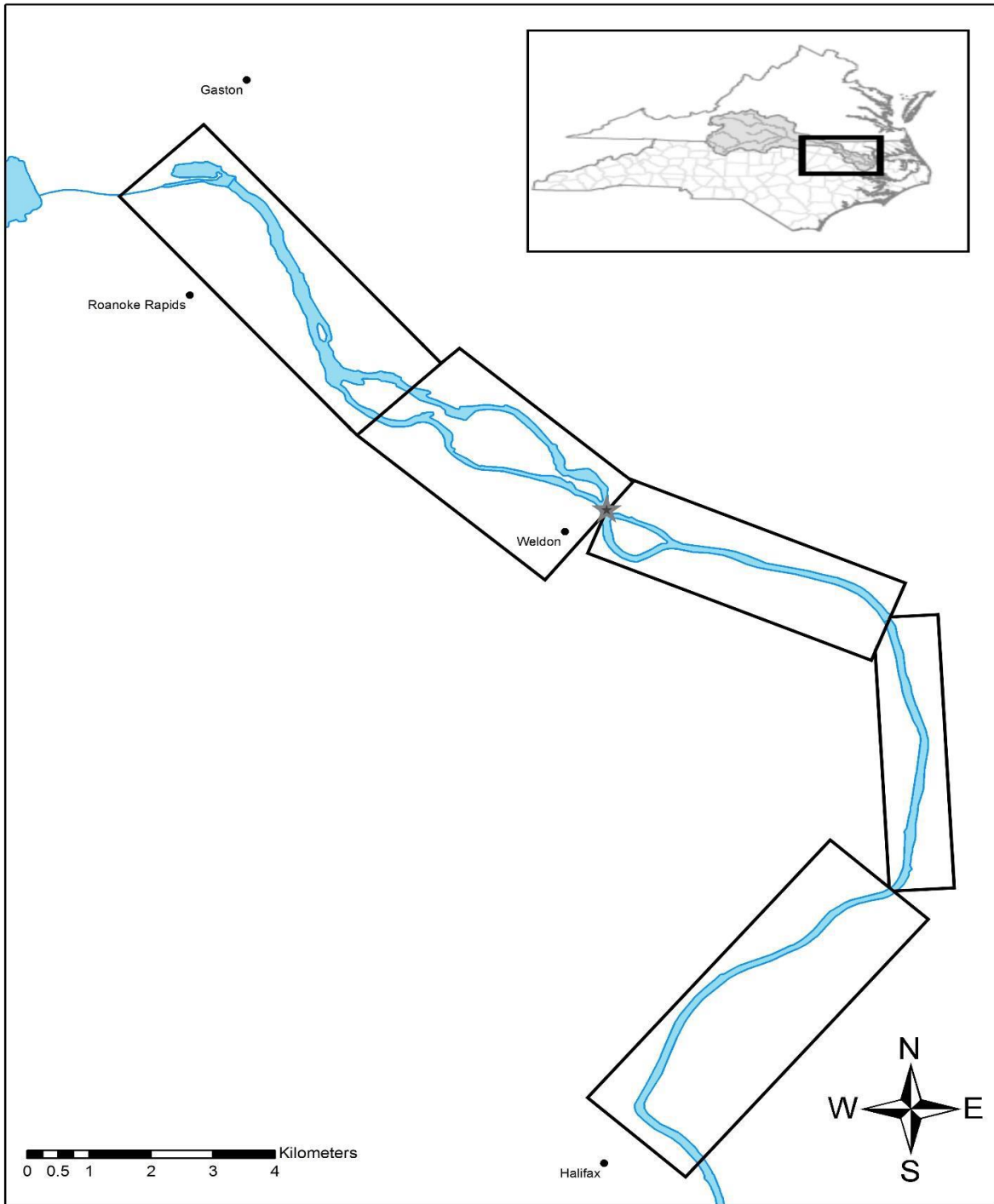
**Figure 2.27.** Annual length frequencies of striped bass sampled from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017.



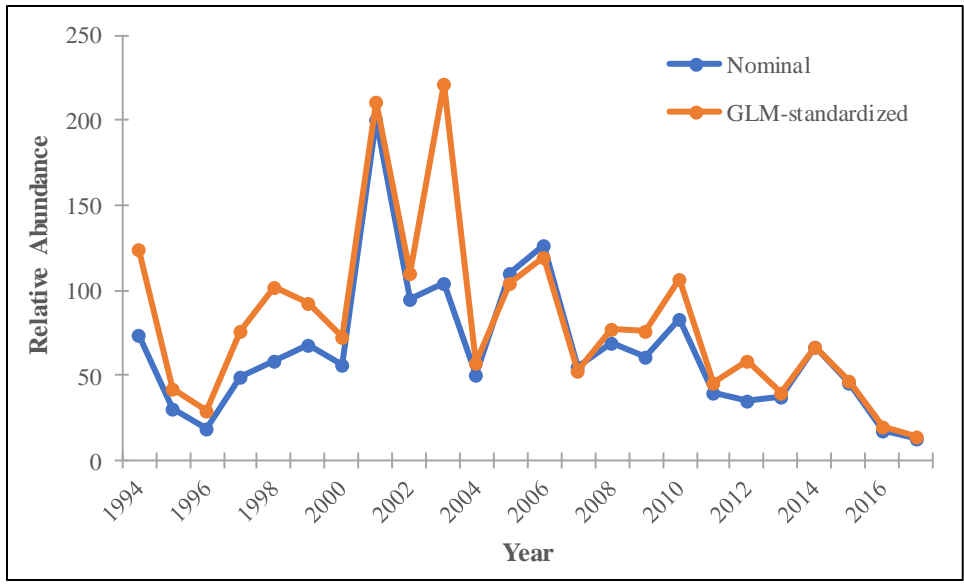
**Figure 2.28.** Annual age frequencies of striped bass sampled from the fall/winter component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. The age-15 bin represents a plus group.



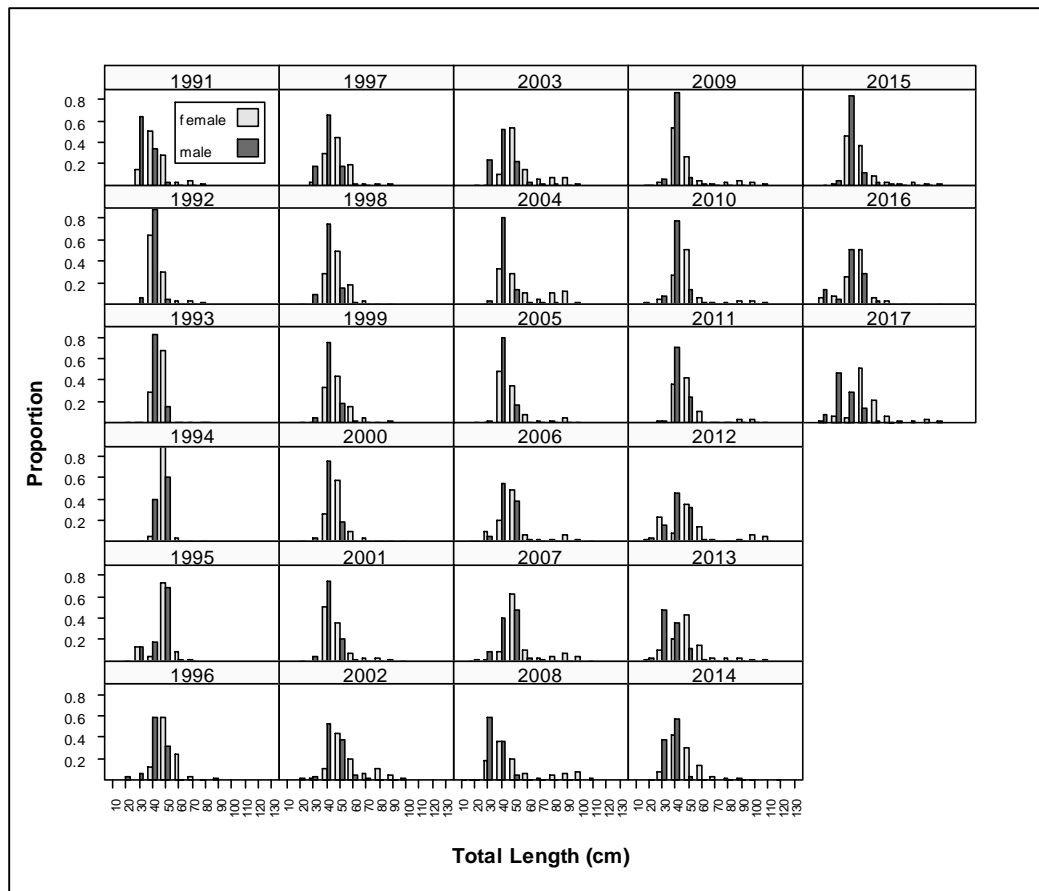
**Figure 2.29.** Annual age frequencies of striped bass sampled from the spring component of the NCDMF Independent Gill-Net Survey (P135), 1991–2017. The age-15 bin represents a plus group.



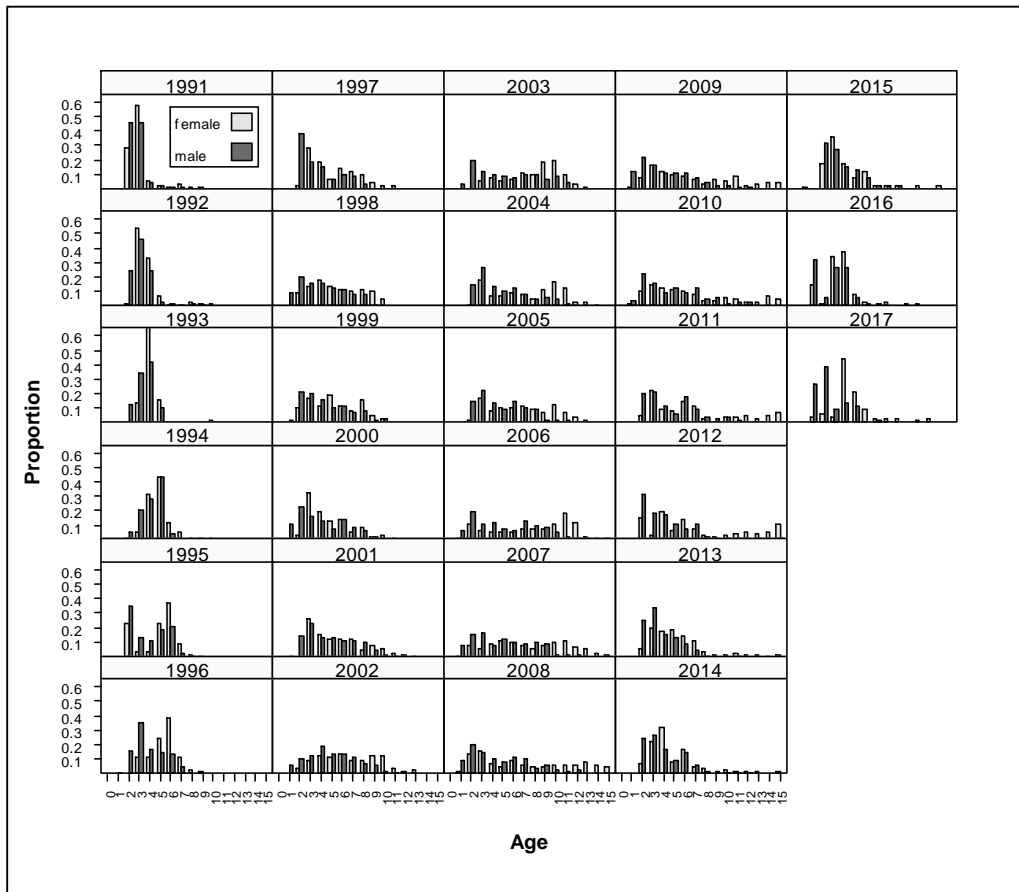
**Figure 2.30.** Striped Bass spawning grounds on the Roanoke River, near the vicinity of Weldon, North Carolina. Black boxes represent relative locations of river strata. The gray star indicates location of rapids near the Weldon boating access area; flows less than 7,000 cfs restrict access to the strata above this location.



**Figure 2.31.** Nominal and GLM-standardized indices of relative abundance derived from the NCWRC Roanoke River Electrofishing Survey, 1994–2017.

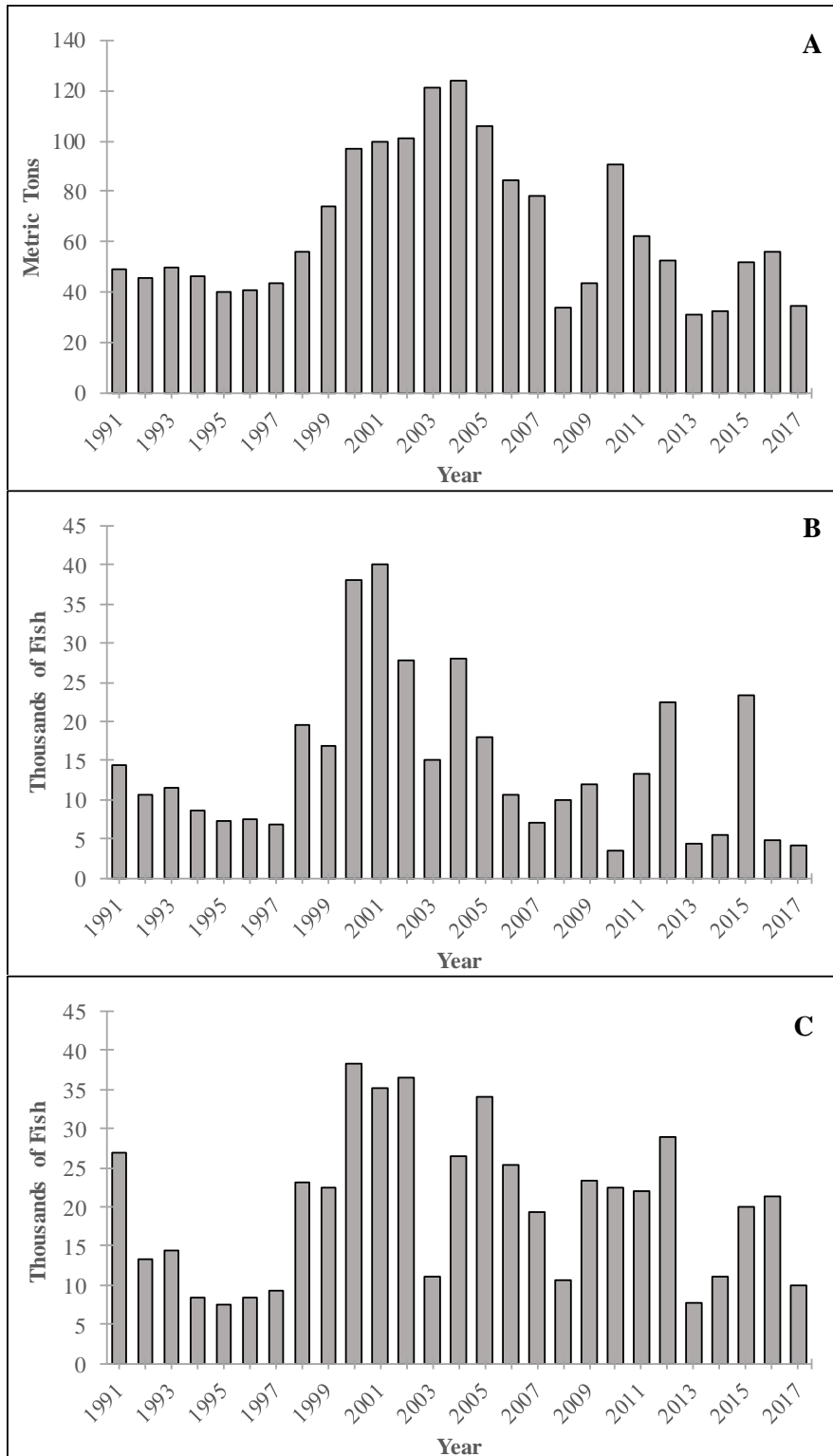


**Figure 2.32.** Annual length frequencies of striped bass sampled from the NCWRC Roanoke River Electrofishing Survey, 1991–2017.

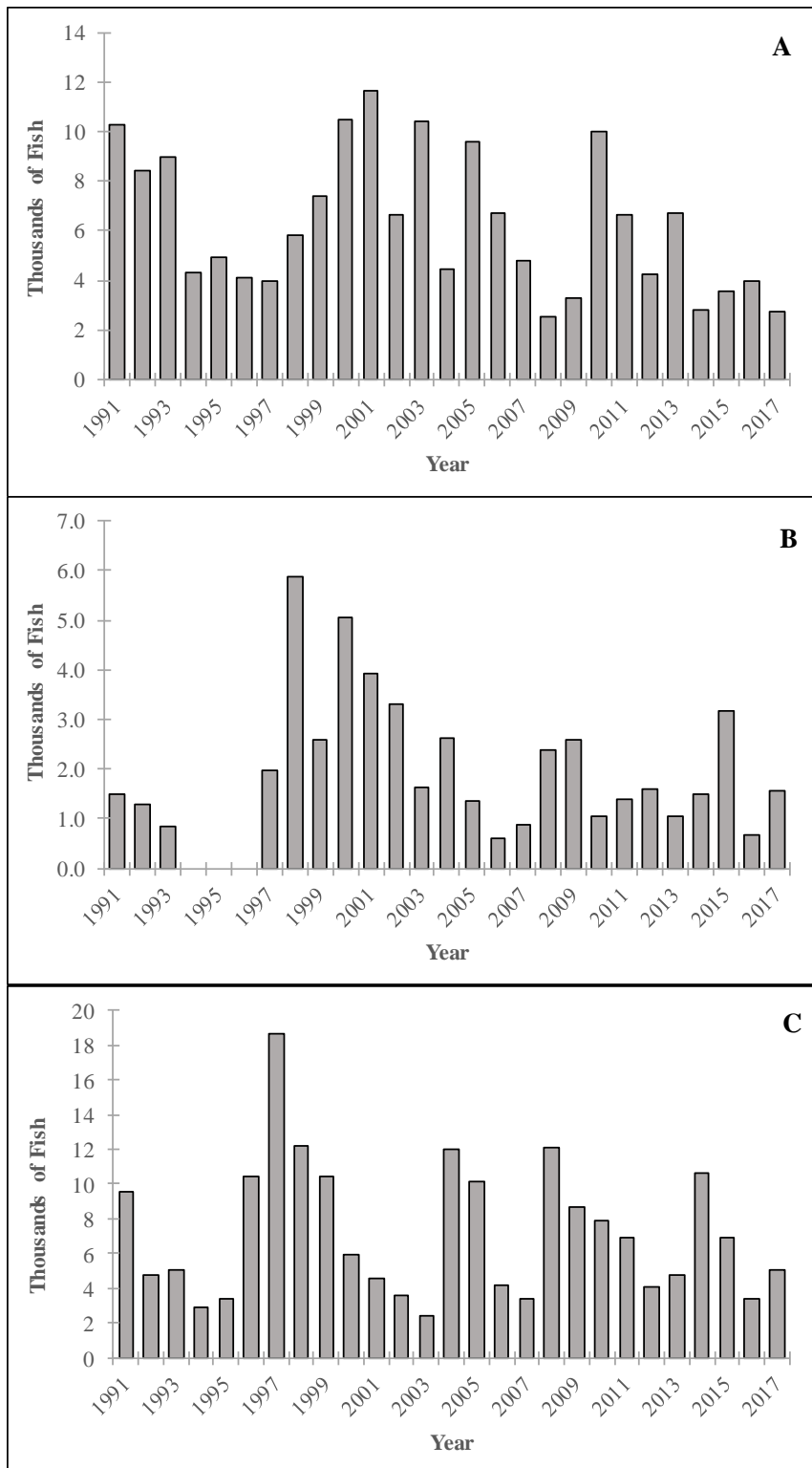


**Figure 2.33.** Annual age frequencies of striped bass sampled from the NCWRC Roanoke River Electrofishing Survey, 1991–2017. The age-15 bin represents a plus group.

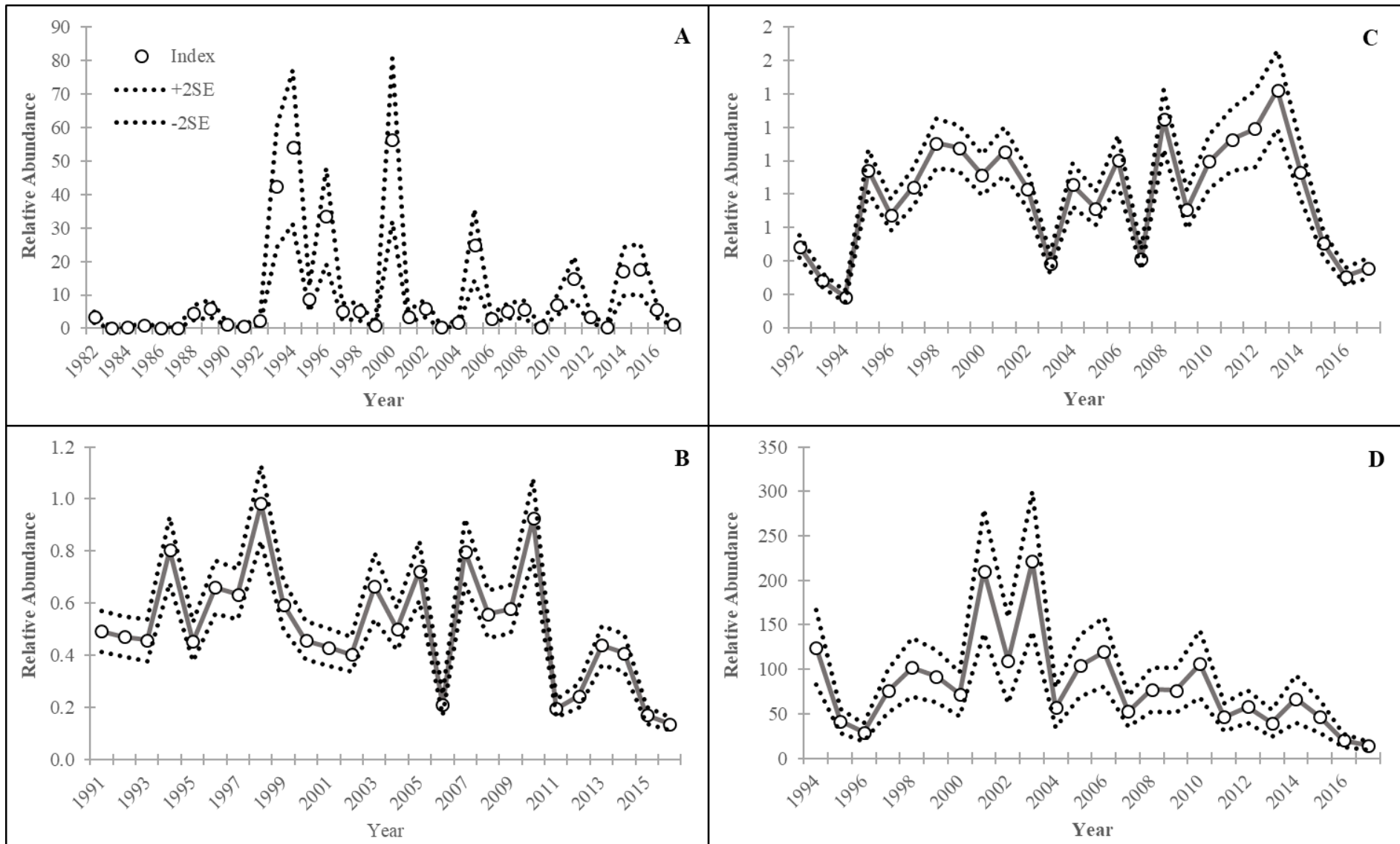




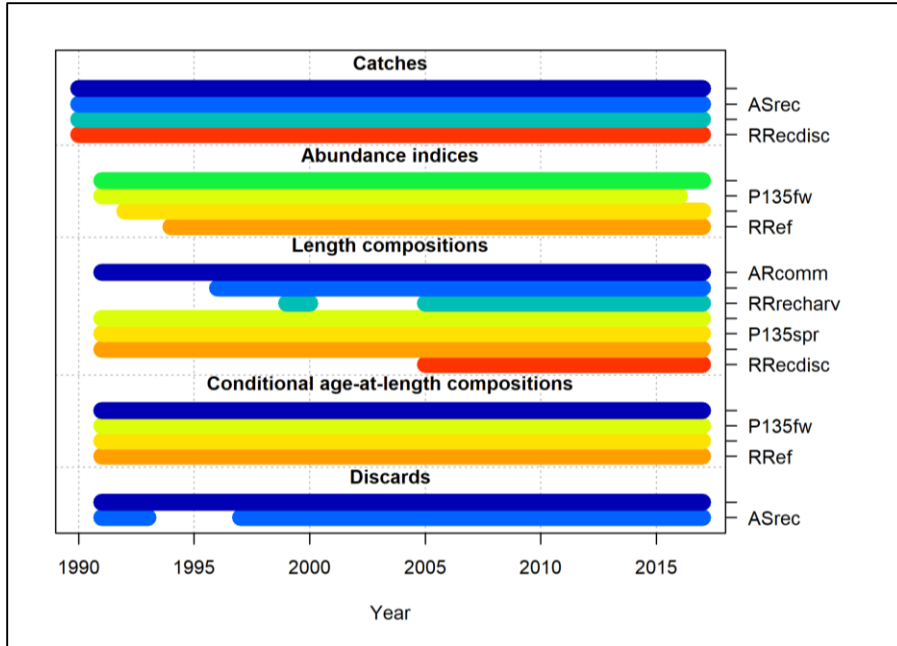
**Figure 3.1.** Annual (A) ARcomm landings, (B) ASrec harvest, and (C) RRrec harvest values that were input into the SS model, 1991–2017.



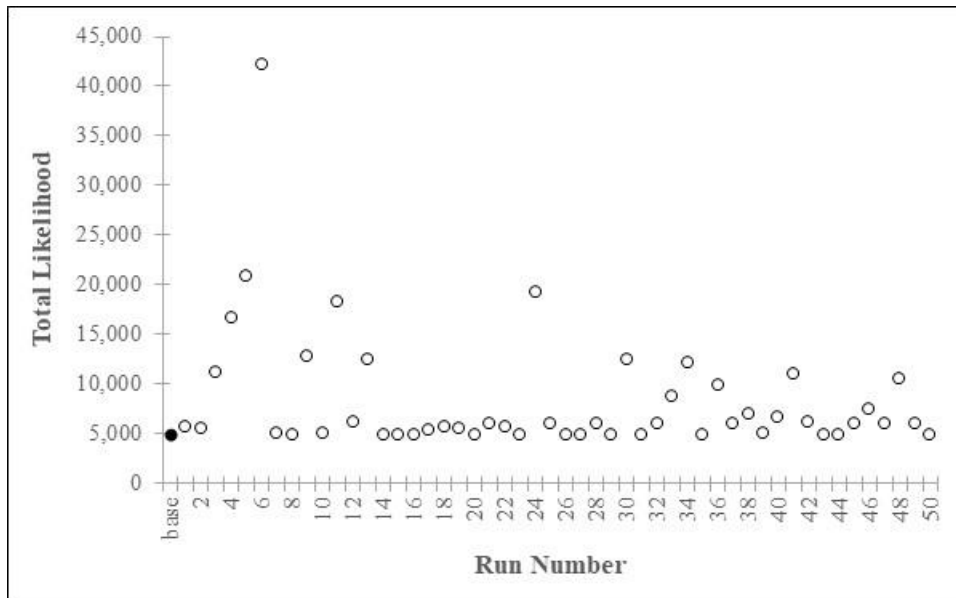
**Figure 3.2.** Annual (A) ARcomm, (B) ASrec, and (C) RRrec dead discards that were input into the SS model, 1991–2017.



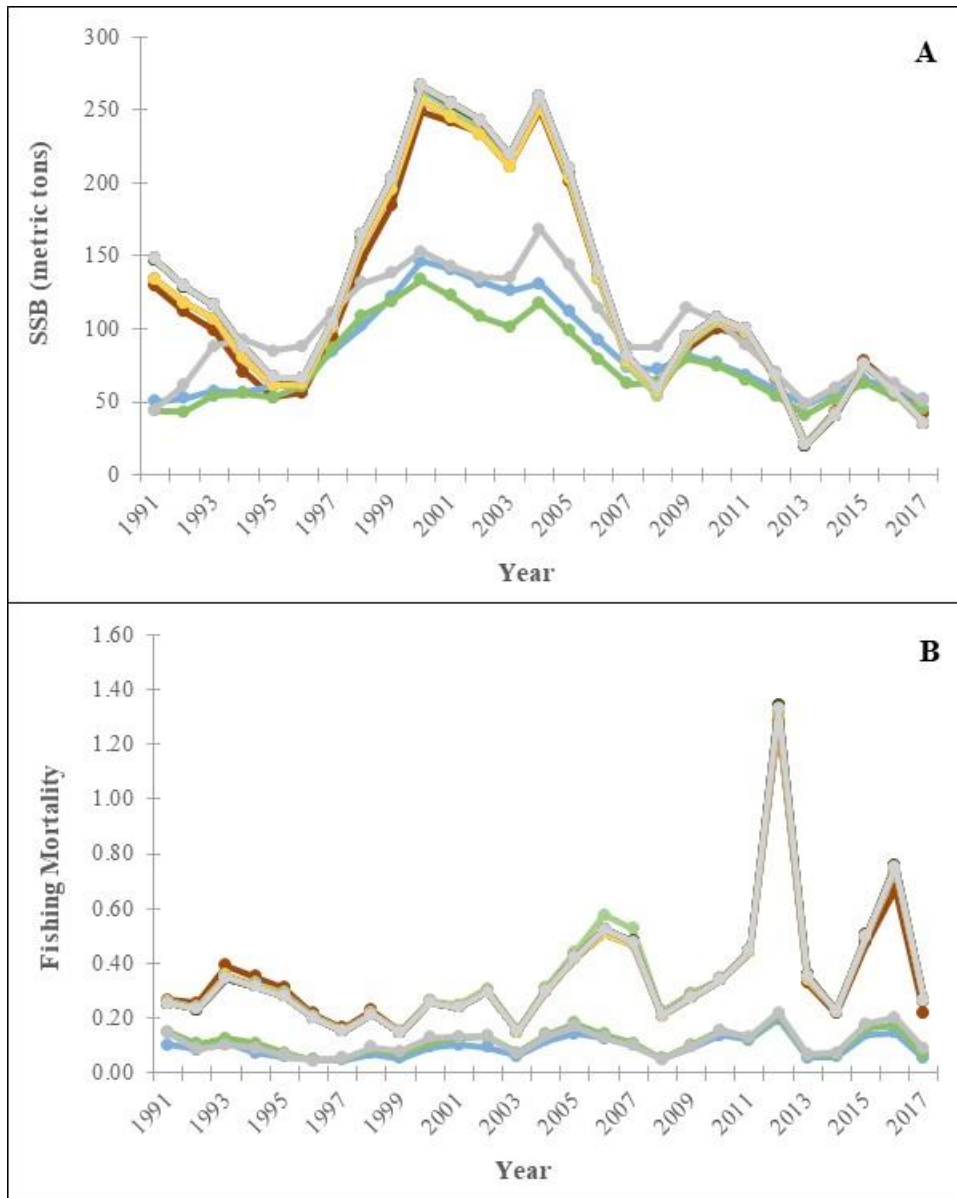
**Figure 3.3.** GLM-standardized indices of abundance derived from the (A) P100juv, (B) P135fw, (C) P135spr, and (D) RRef surveys that were input into the SS model, 1991–2017.



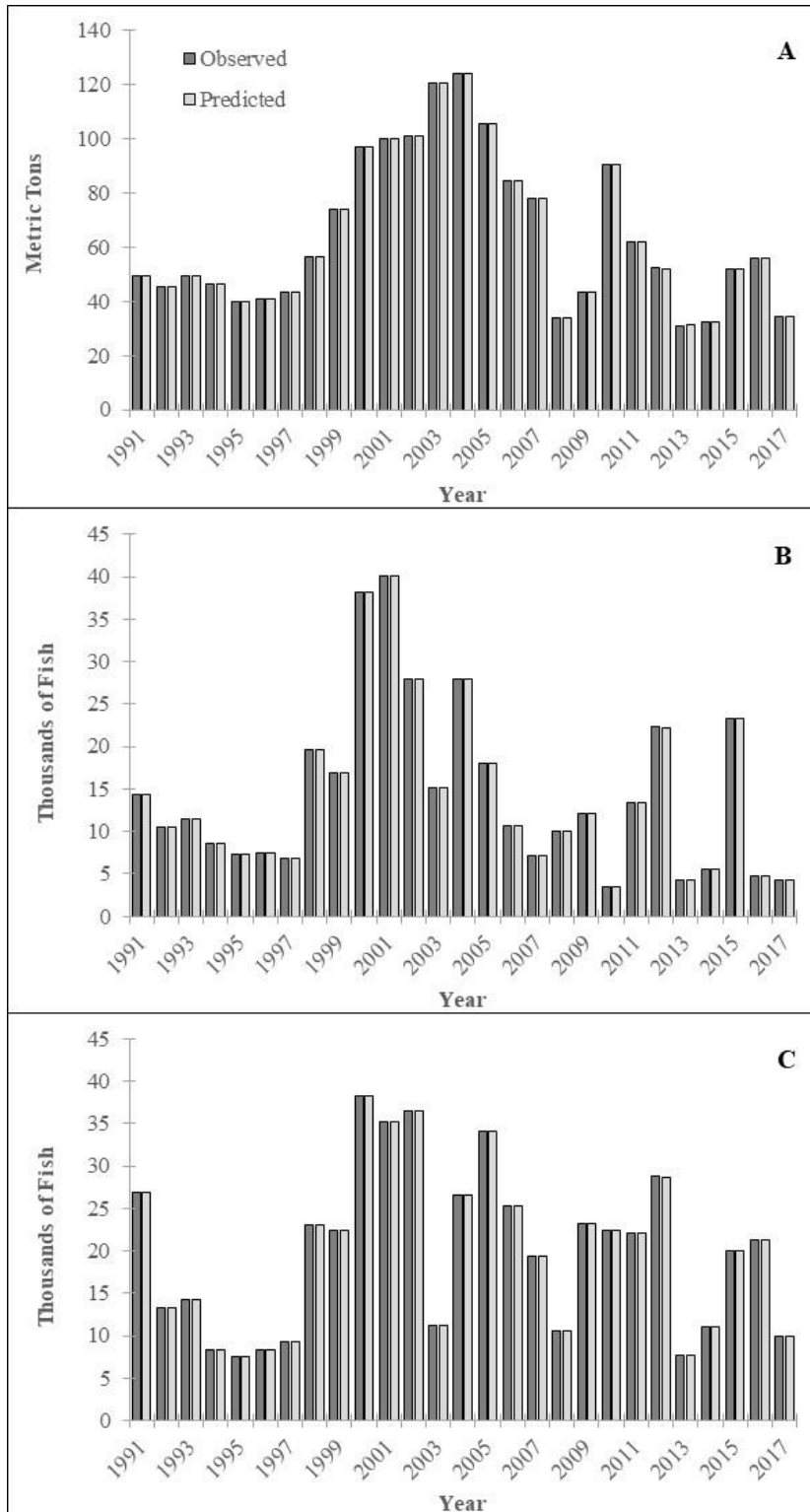
**Figure 3.4.** Summary of the data sources and types used in the stock assessment model for striped bass.



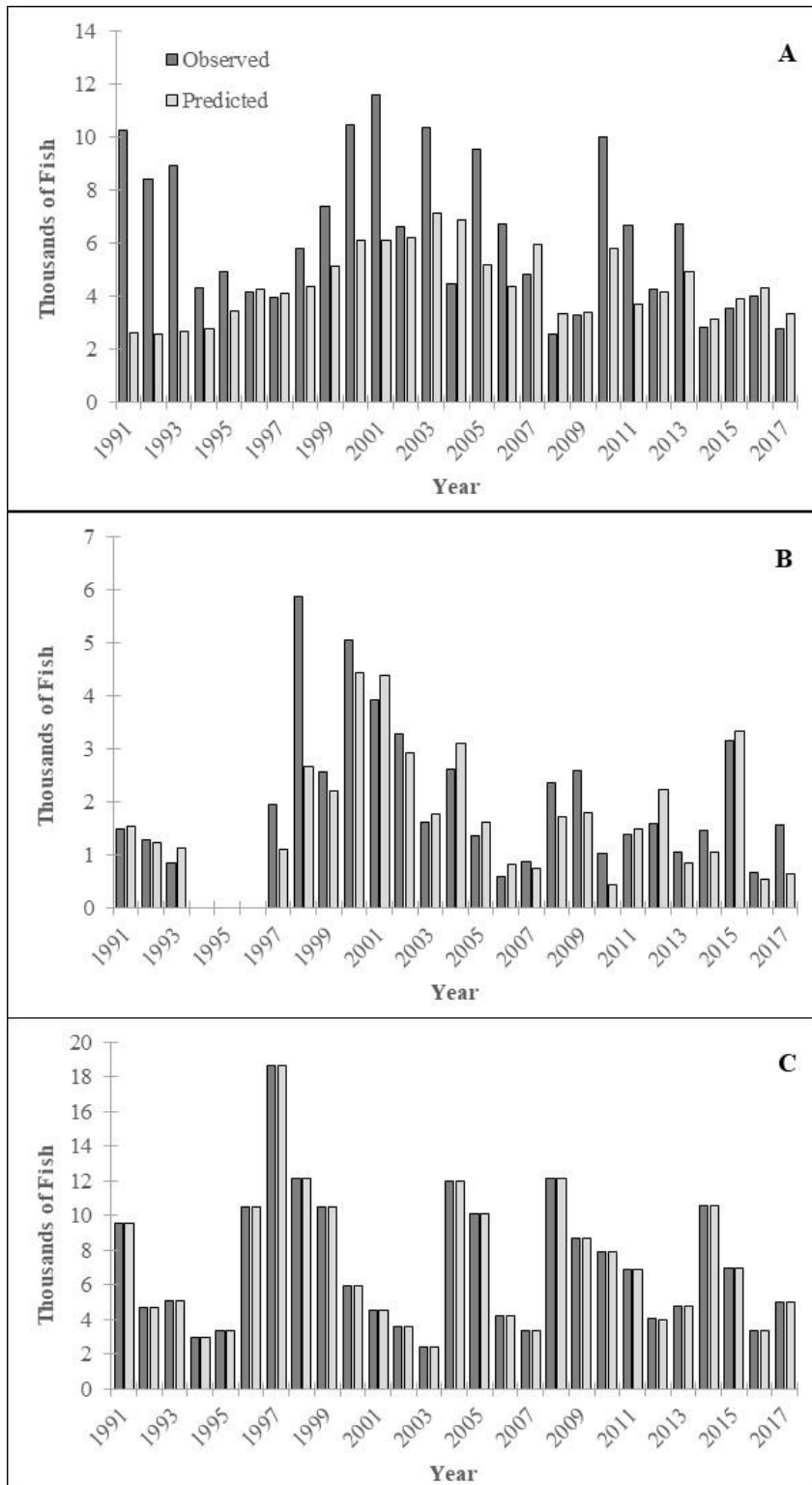
**Figure 3.5.** Negative log-likelihood values produced from the 50 jitter trials in which initial parameter values were jittered by 10%. The solid black circle is the value from the base run.



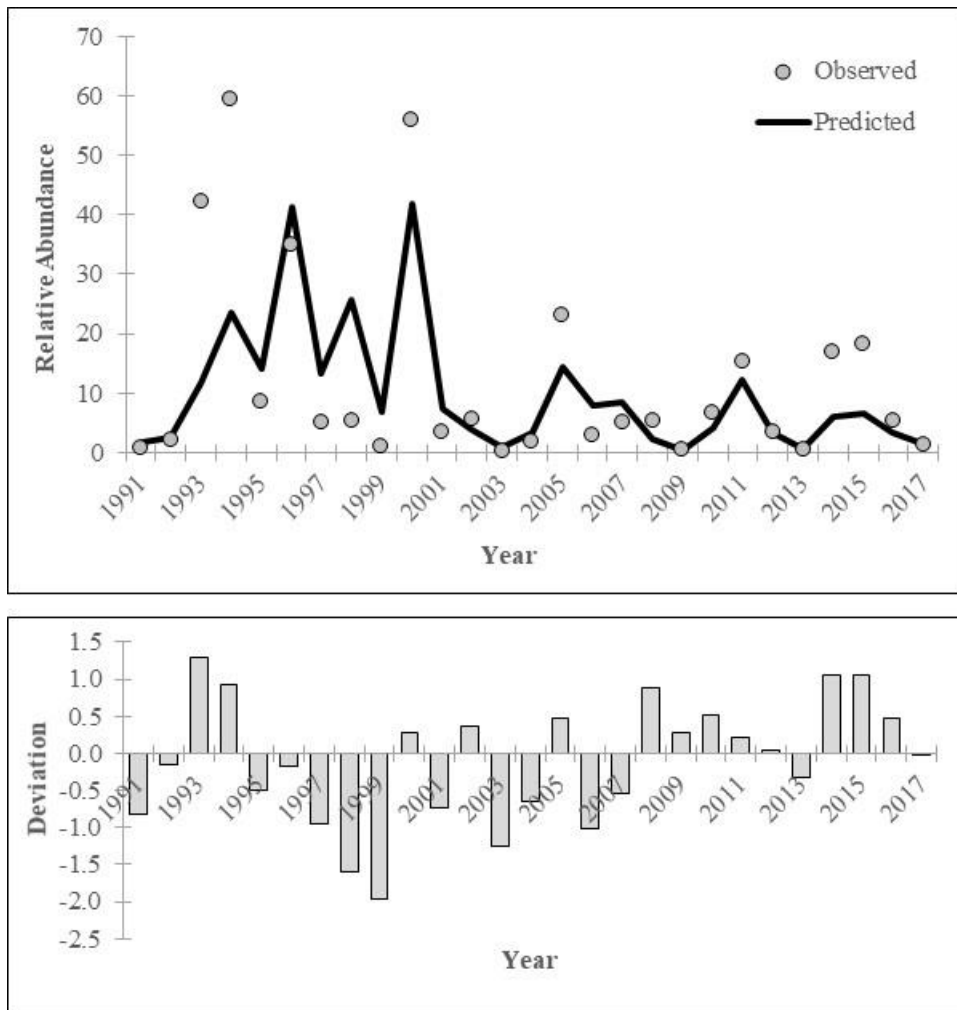
**Figure 3.6.** Predicted (A) female SSB and (B)  $F$  (numbers-weighted, ages 3–5) from the converged jitter trials (run 46 removed) in which initial parameter values were jittered by 10%, 1991–2017.



**Figure 3.7.** Observed and predicted (A) ARcomm landings, (B) ASrec harvest, and (C) RRrec harvest from the base run of the stock assessment model, 1991–2017.

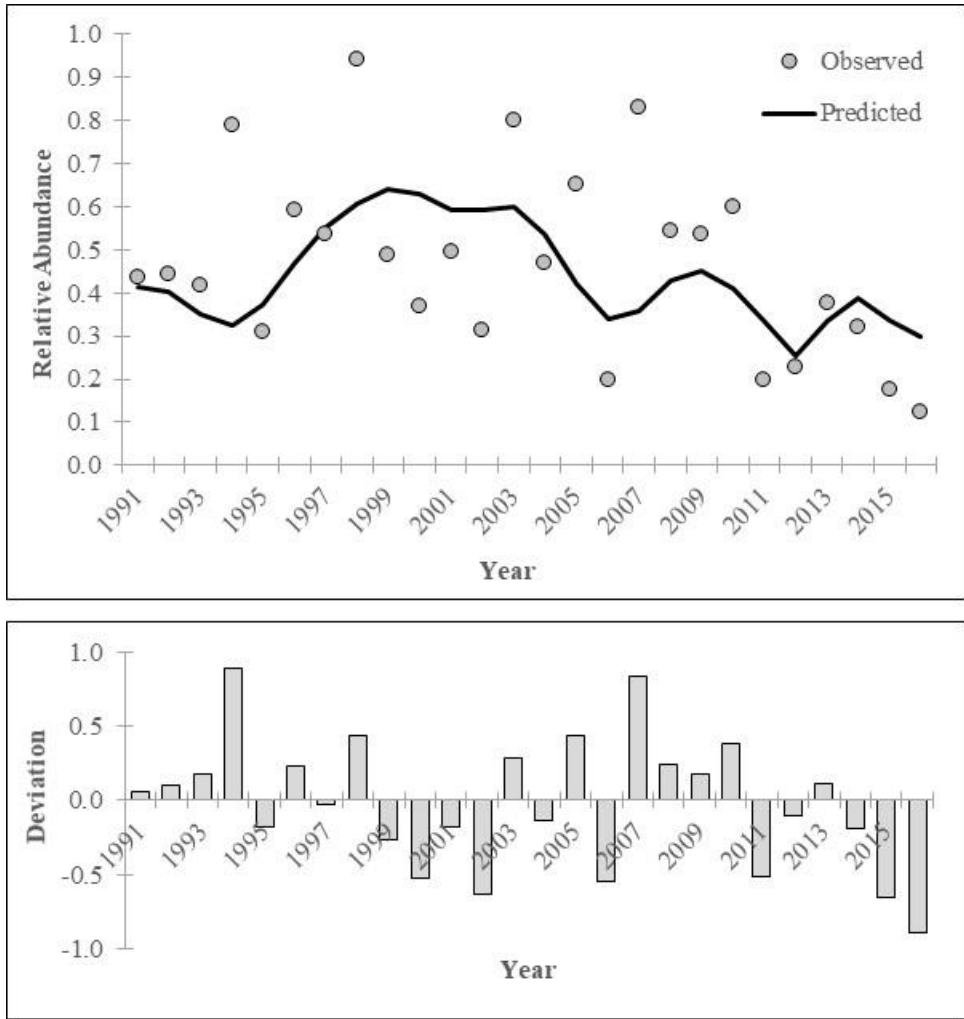


**Figure 3.8.** Observed and predicted (A) ARcomm, (B) ASrec, and (C) RRrec dead discards from the base run of the stock assessment model, 1991–2017.

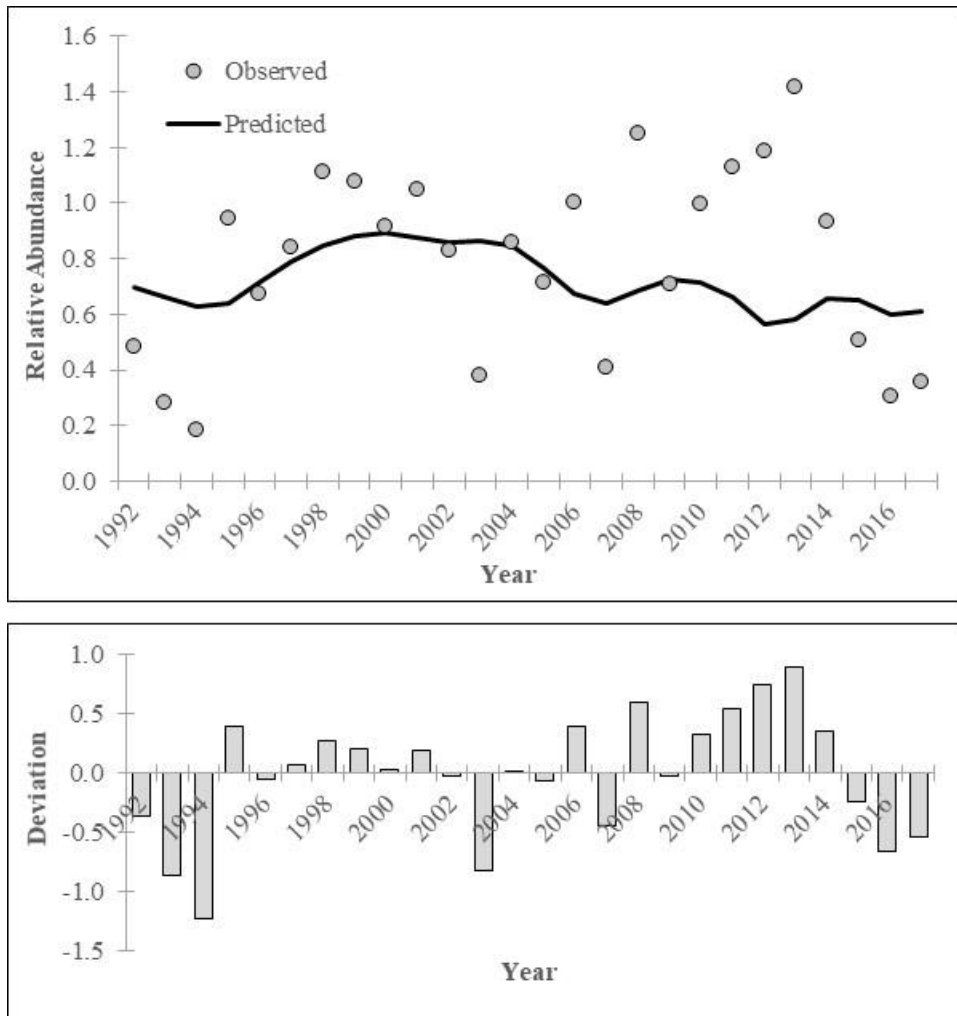


**Figure 3.9.** Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P100juv survey from the base run of the stock assessment model, 1991–2017.

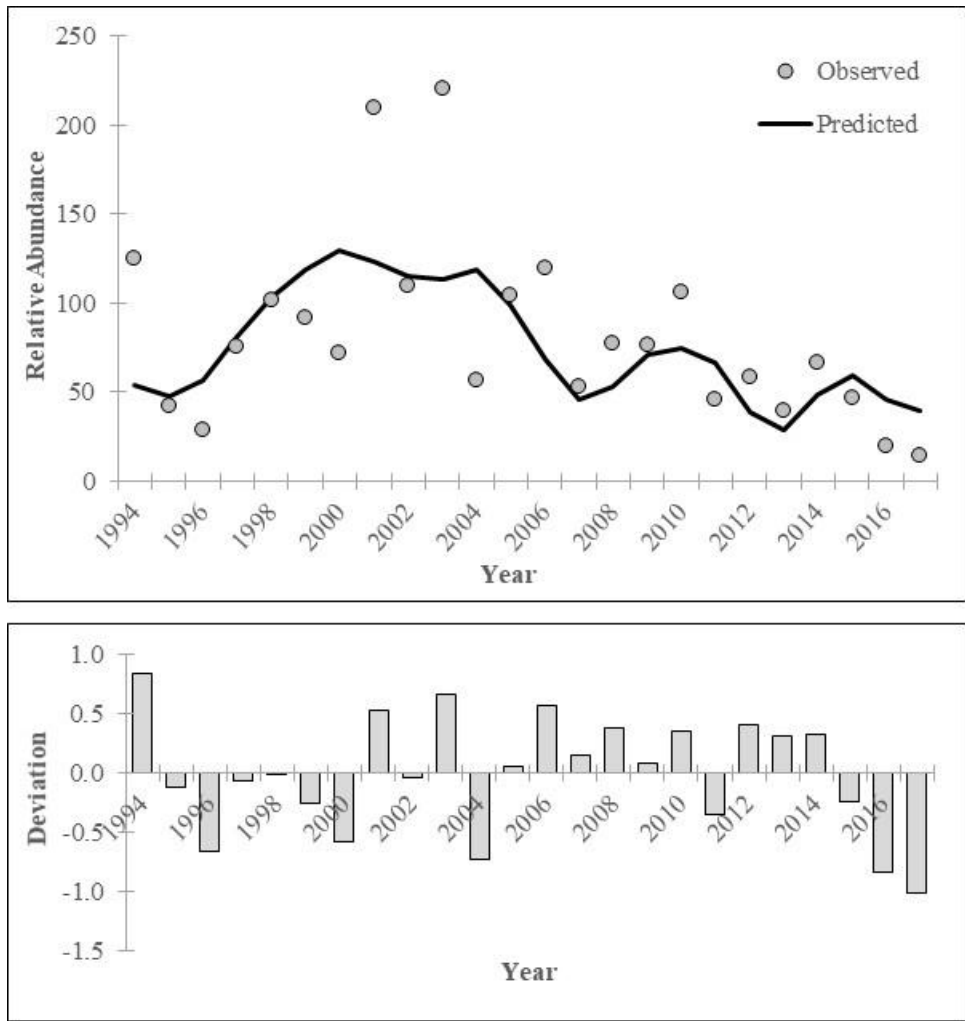




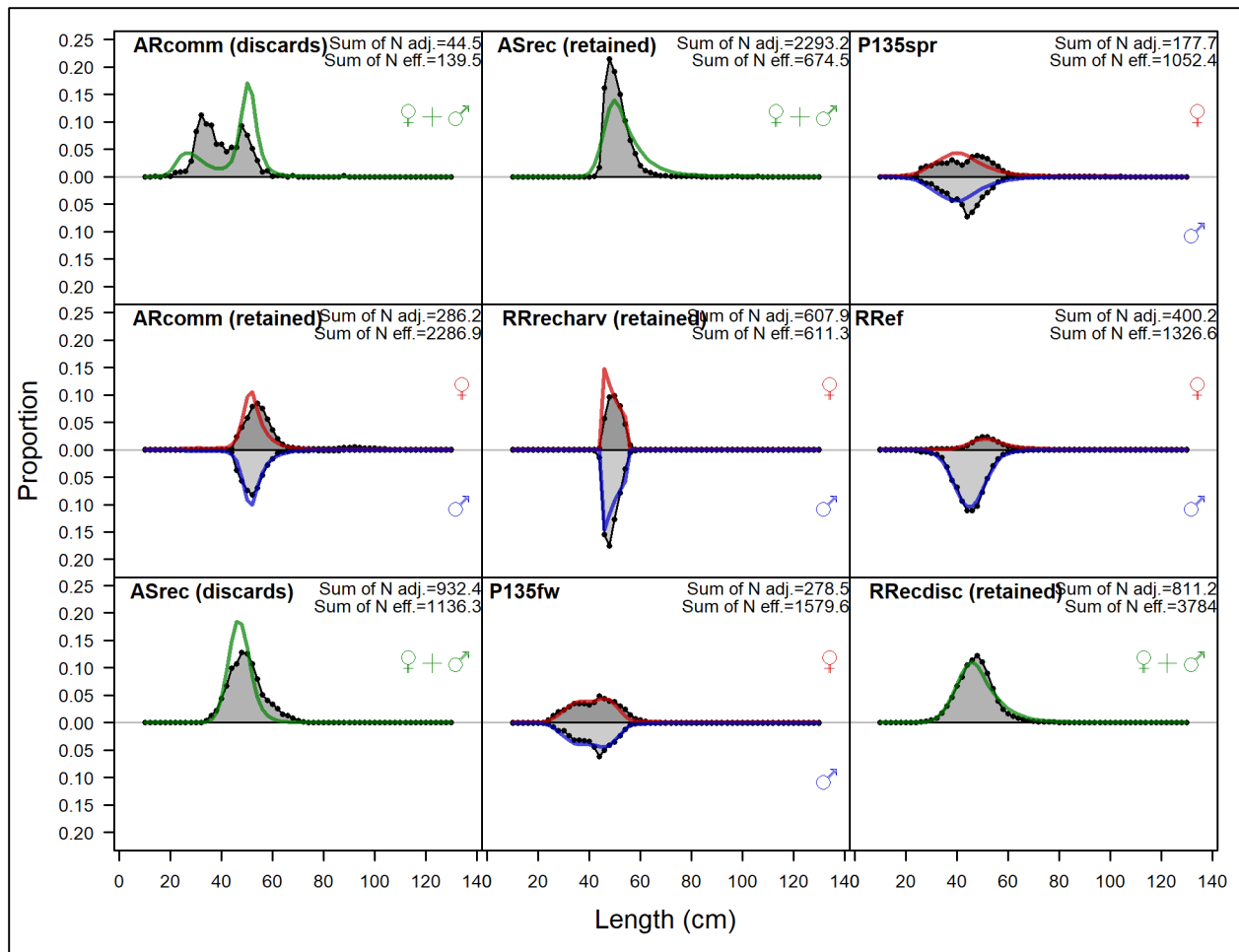
**Figure 3.10.** Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P135fw survey from the base run of the stock assessment model, 1991–2017.



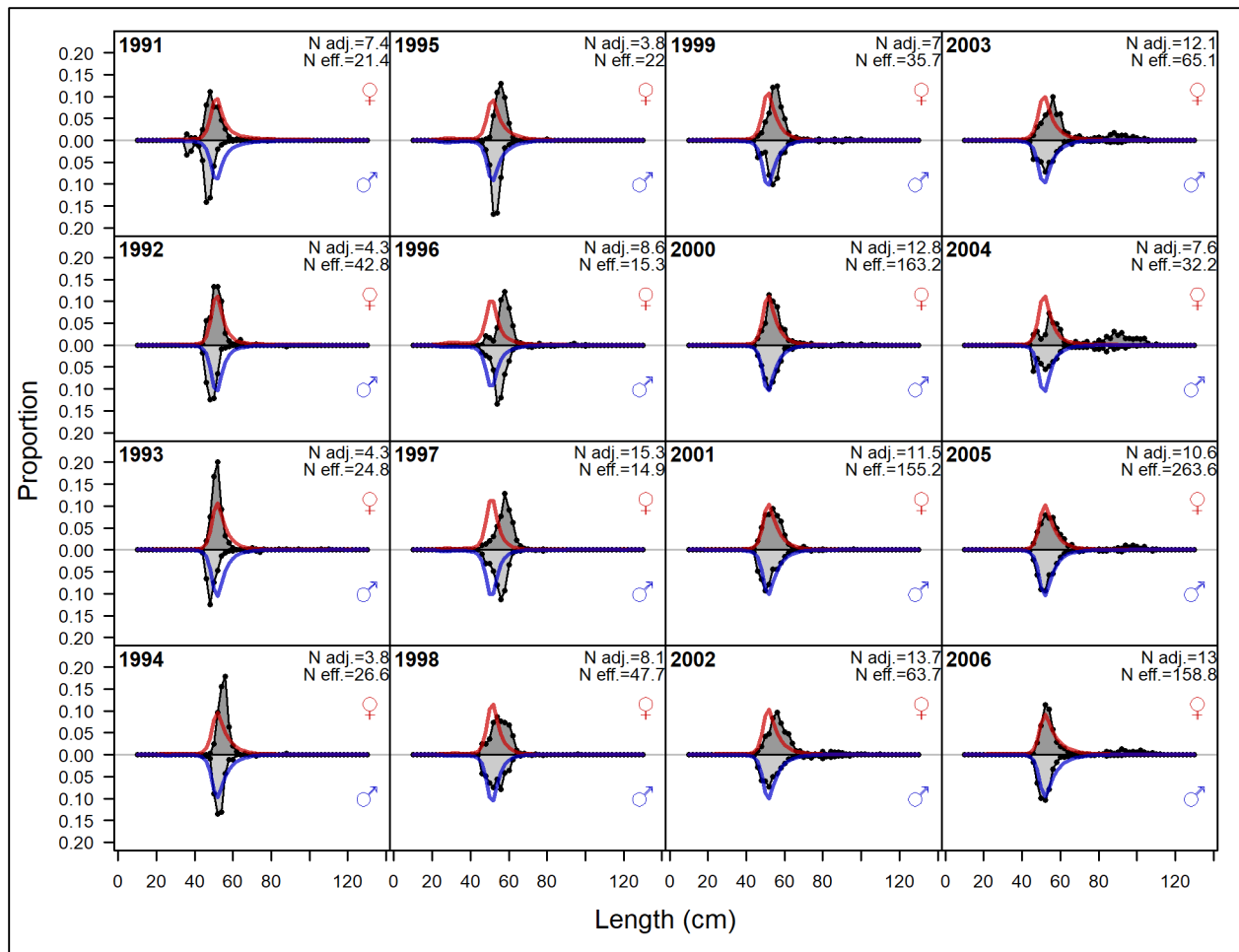
**Figure 3.11.** Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the P135spr survey from the base run of the stock assessment model, 1992–2017.



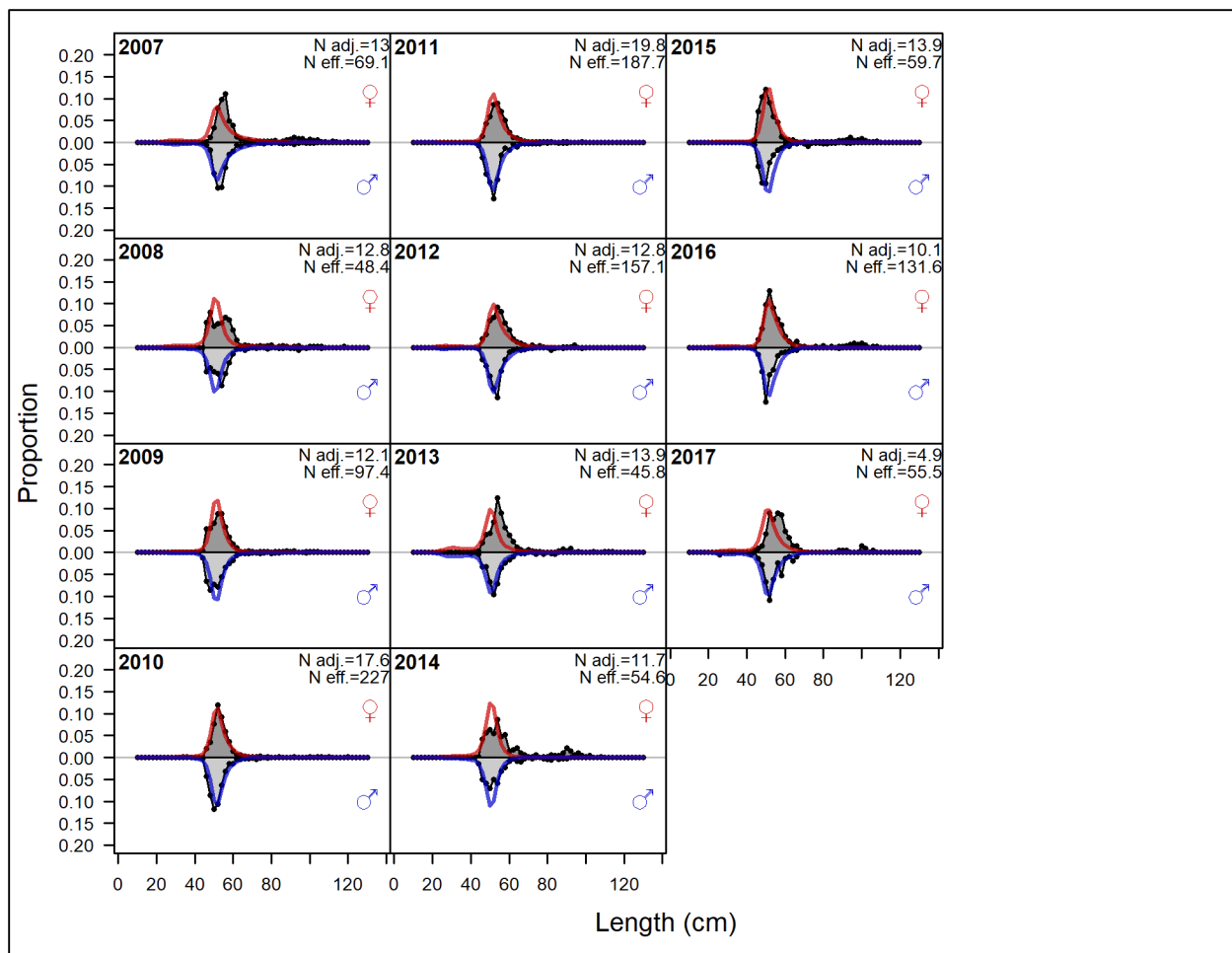
**Figure 3.12.** Observed and predicted relative abundance (top graph) and standardized residuals (bottom graph) for the RRef survey from the base run of the stock assessment model, 1994–2017.



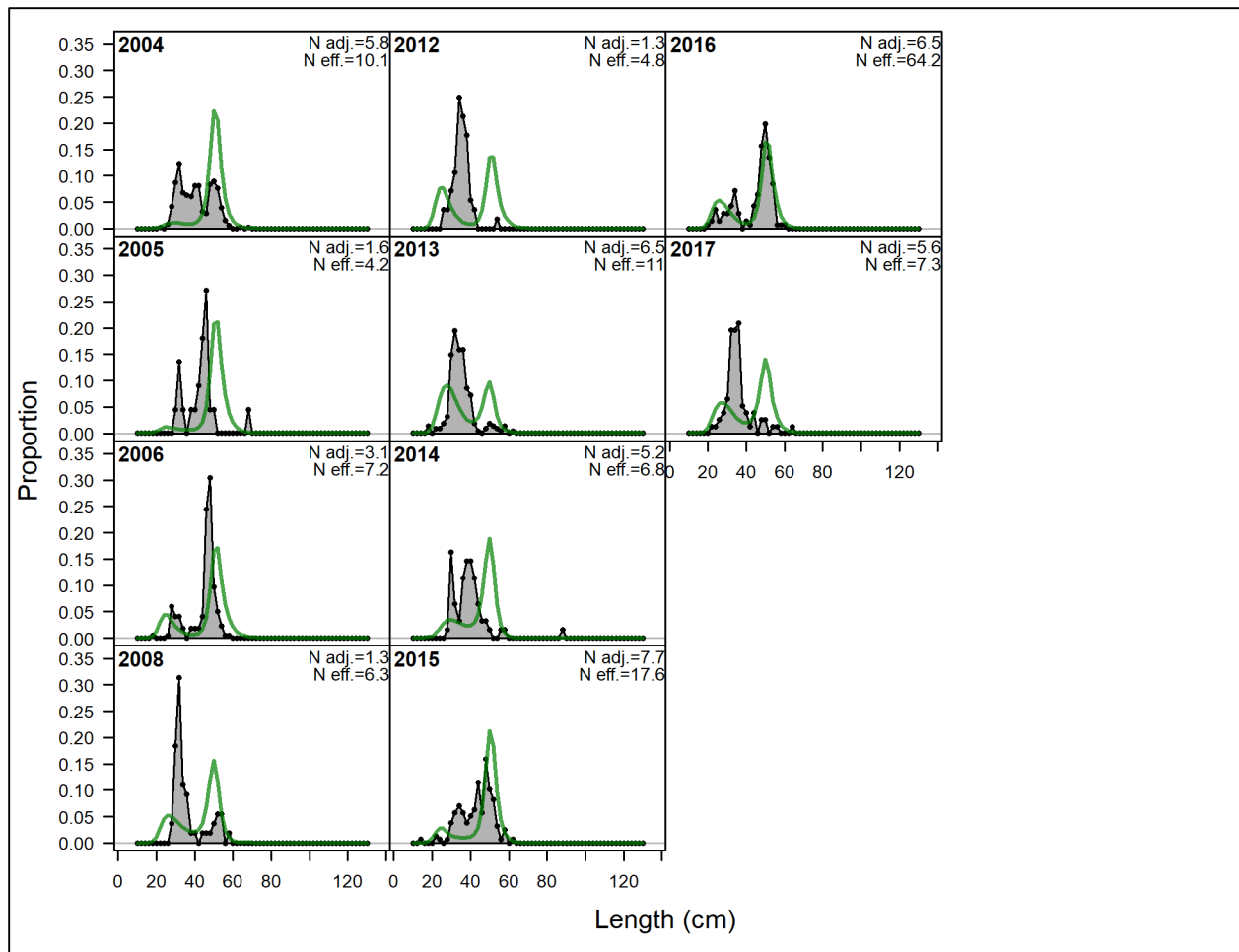
**Figure 3.13.** Observed and predicted length compositions for each data source from the base run of the stock assessment model aggregated across time. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



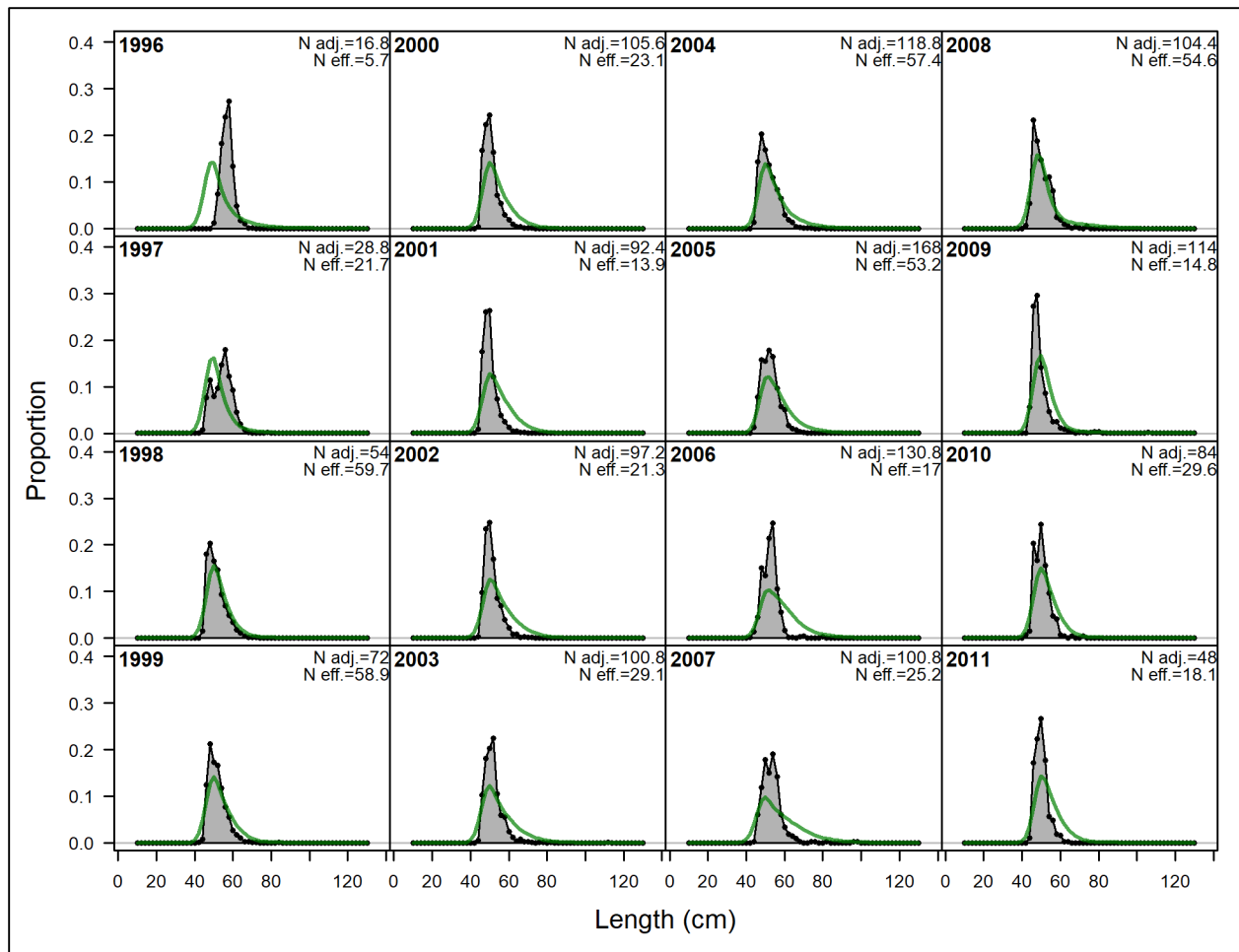
**Figure 3.14.** Observed and predicted length compositions for the ARcomm landings from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



**Figure 3.15.** Observed and predicted length compositions for the ARcomm landings from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

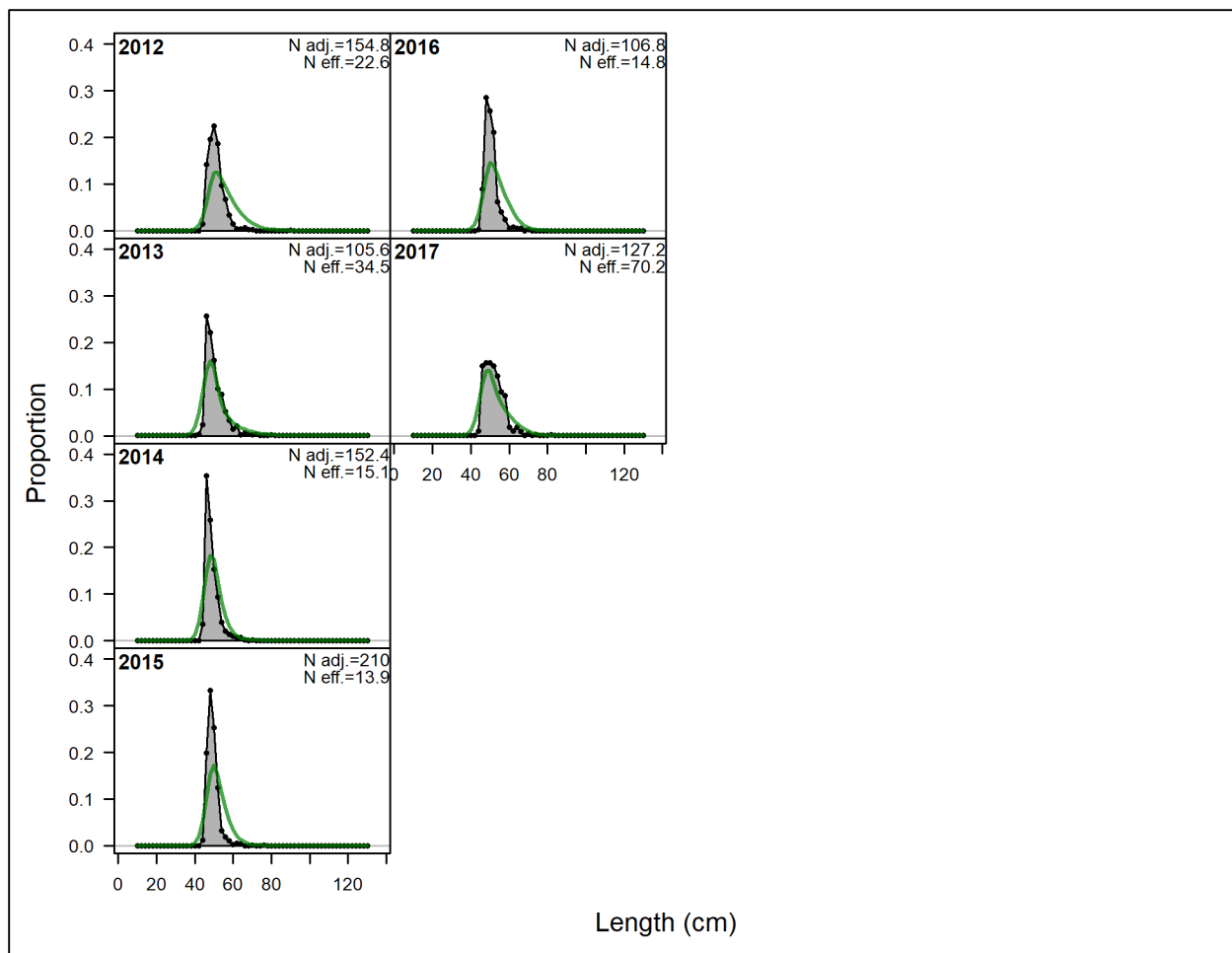


**Figure 3.16.** Observed and predicted length compositions for the ARcomm discards from the base run of the stock assessment model, 2004–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

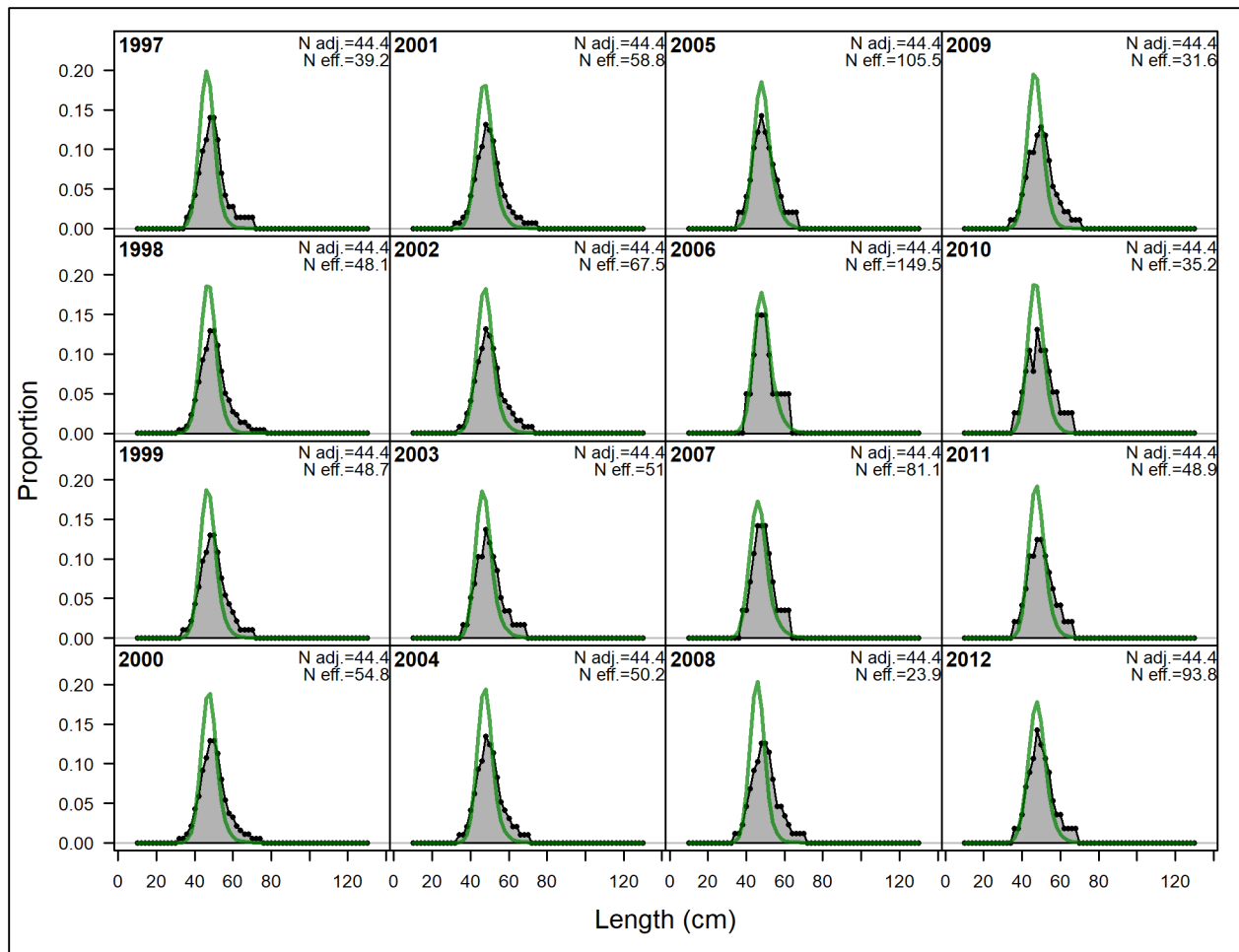


**Figure 3.17.** Observed and predicted length compositions for the ASrec harvest from the base run of the stock assessment model, 1996–2011. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

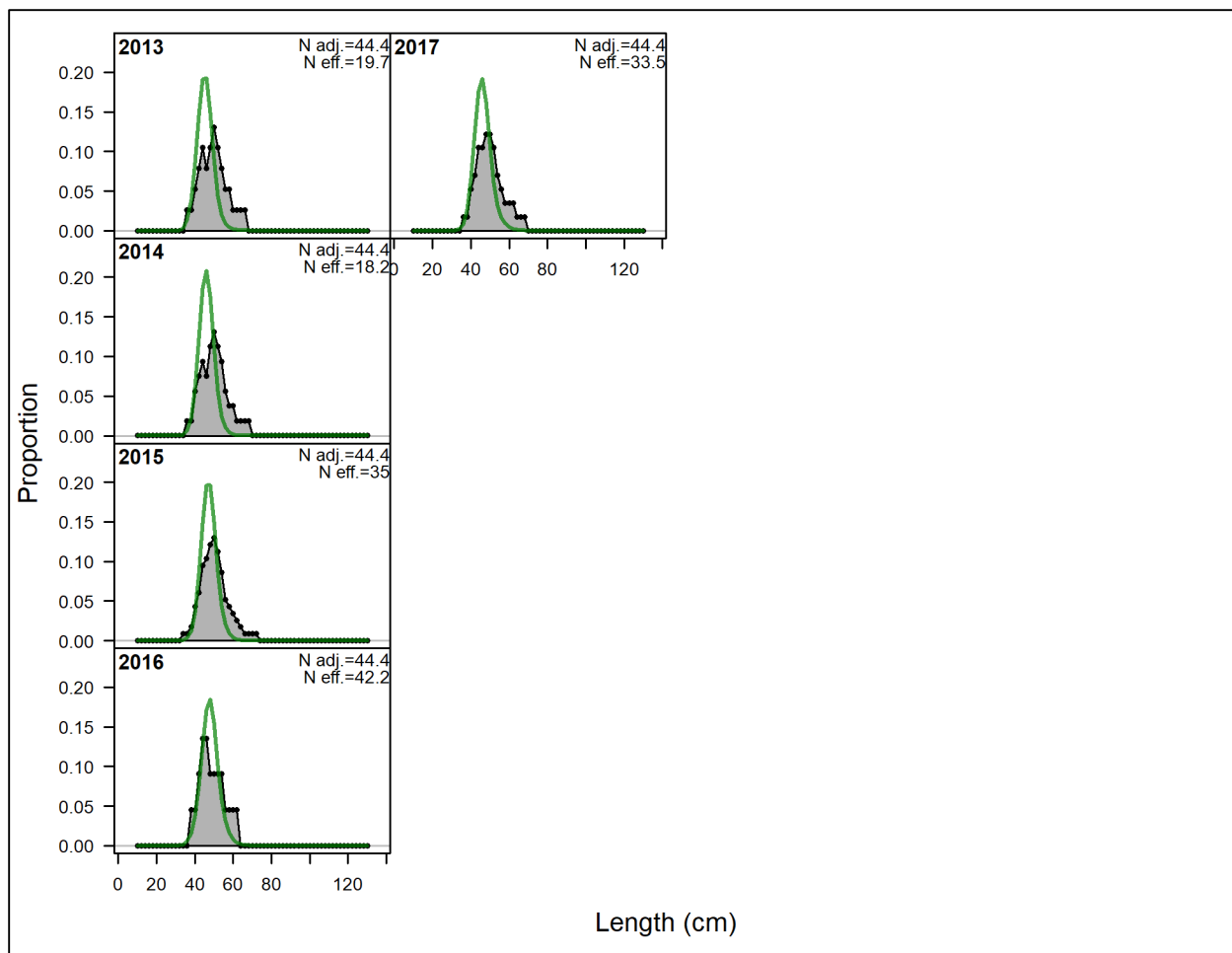




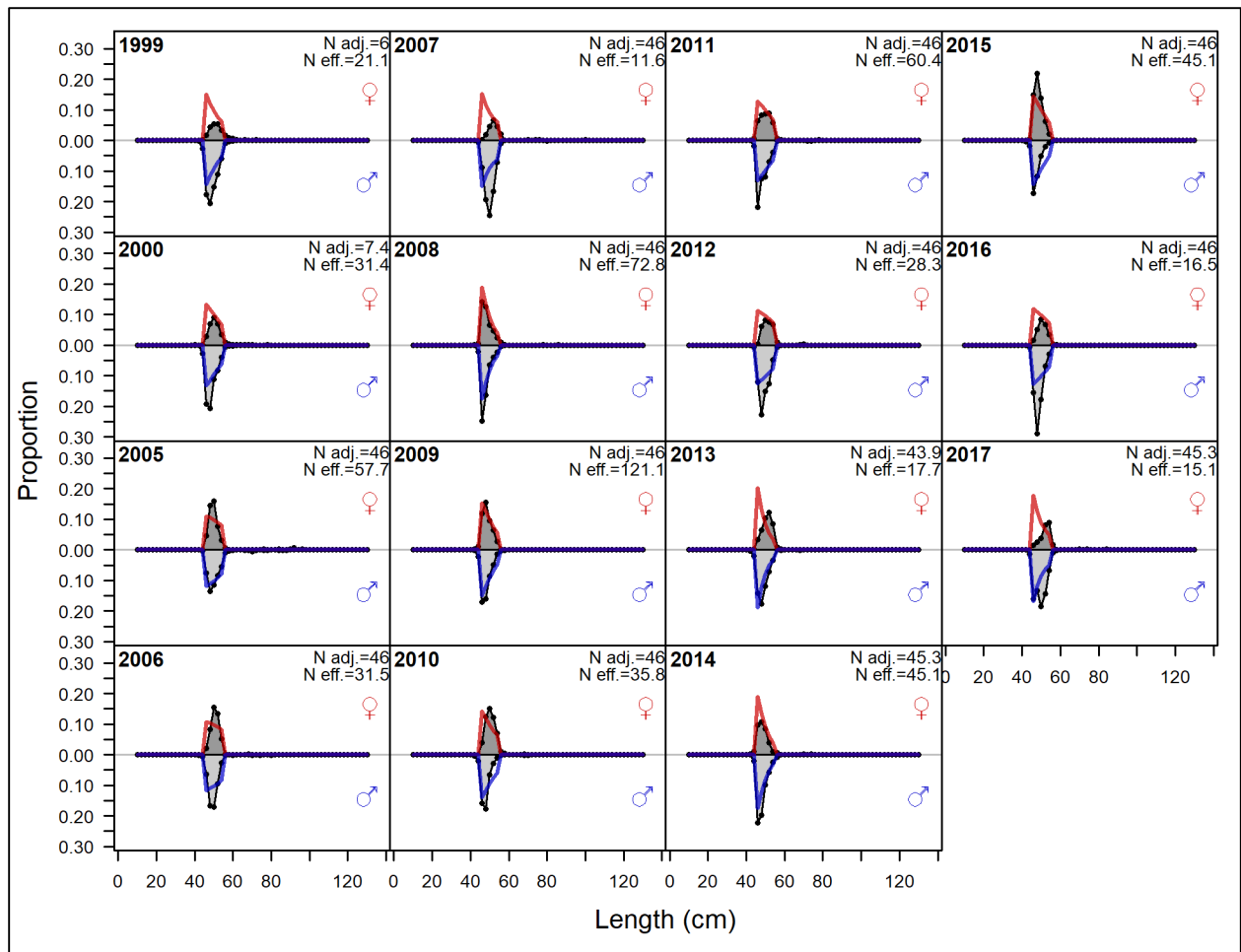
**Figure 3.18.** Observed and predicted length compositions for the ASrec harvest from the base run of the stock assessment model, 2012–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



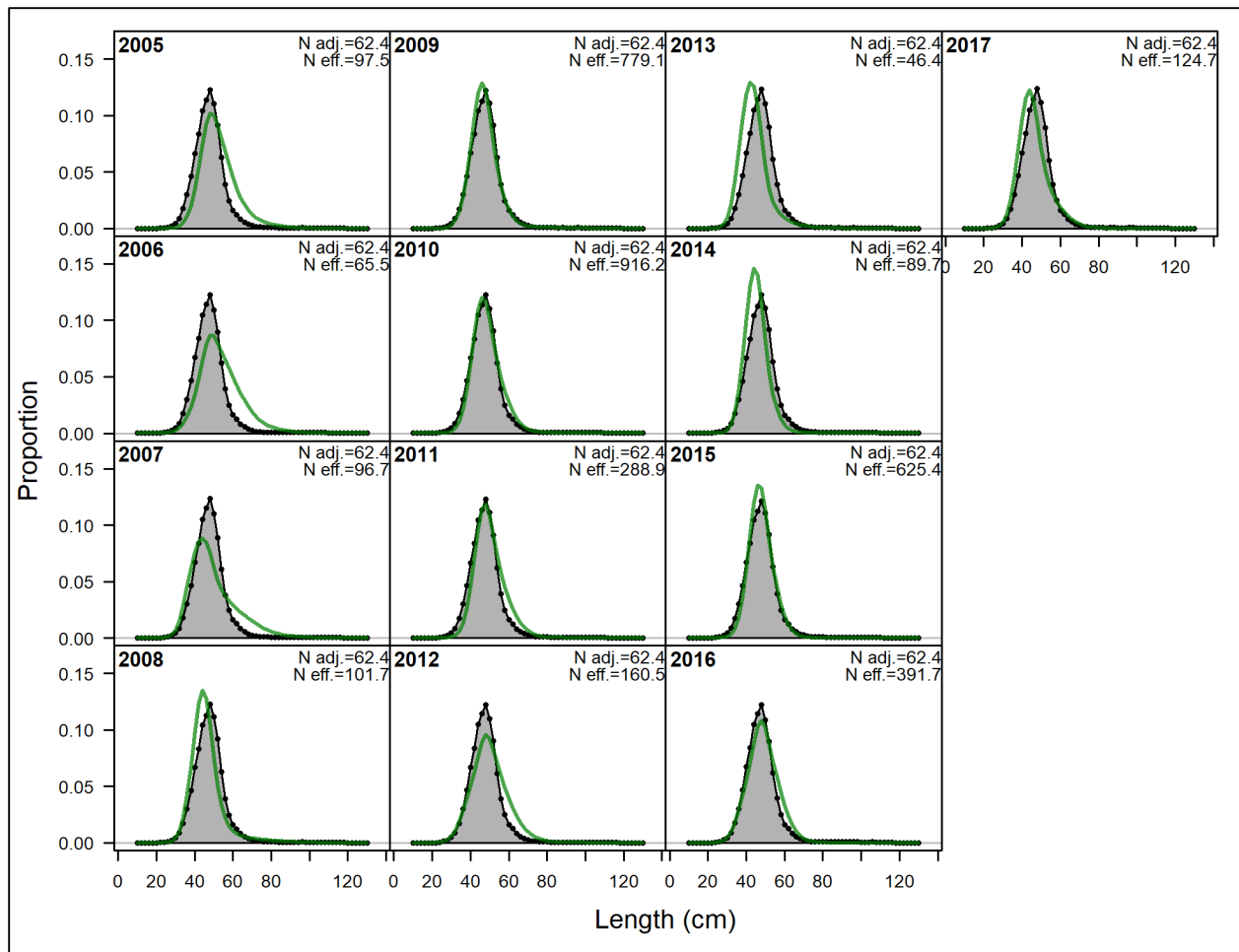
**Figure 3.19.** Observed and predicted length compositions for the ASrec discards from the base run of the stock assessment model, 1997–2012. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



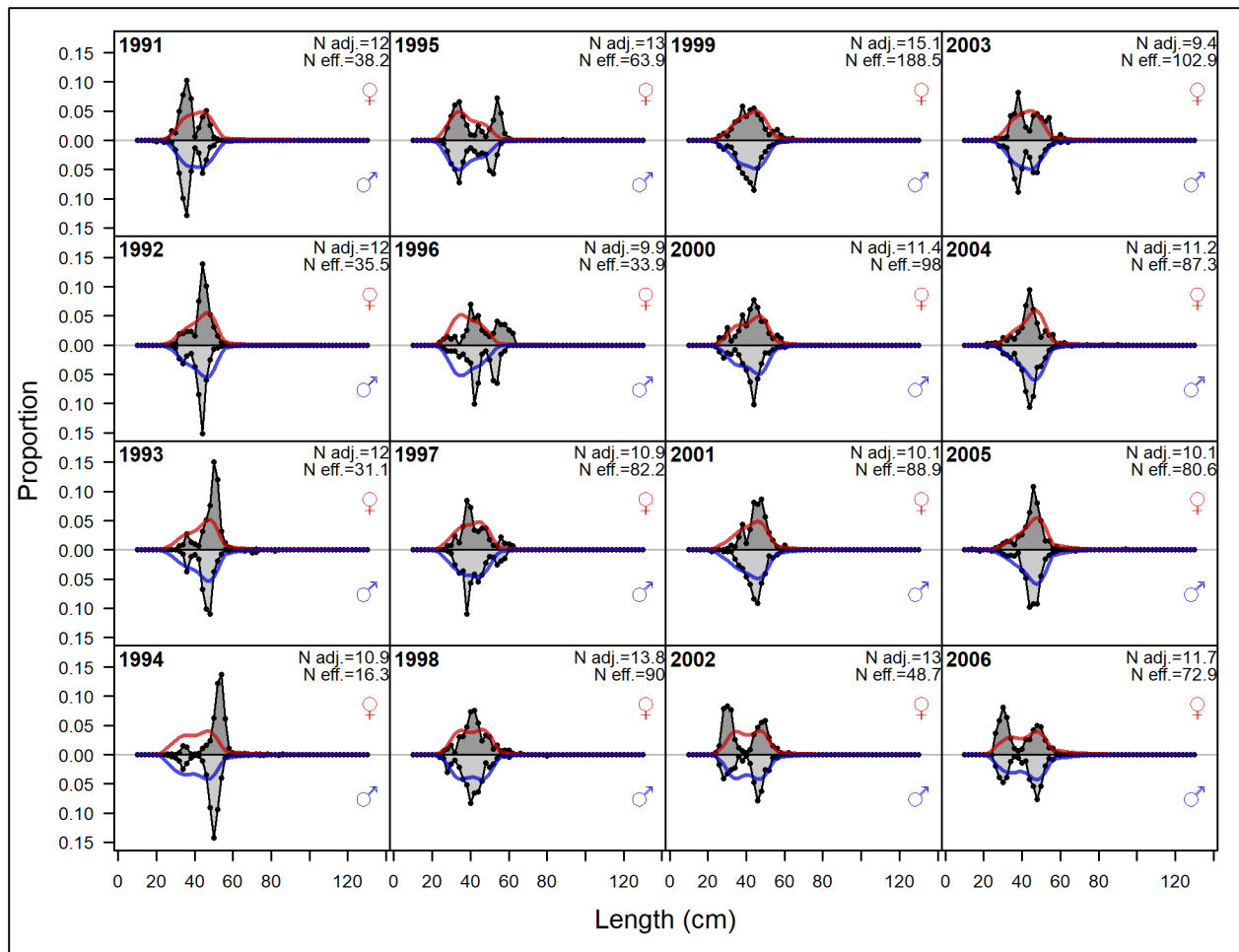
**Figure 3.20.** Observed and predicted length compositions for the ASrec discards from the base run of the stock assessment model, 2013–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



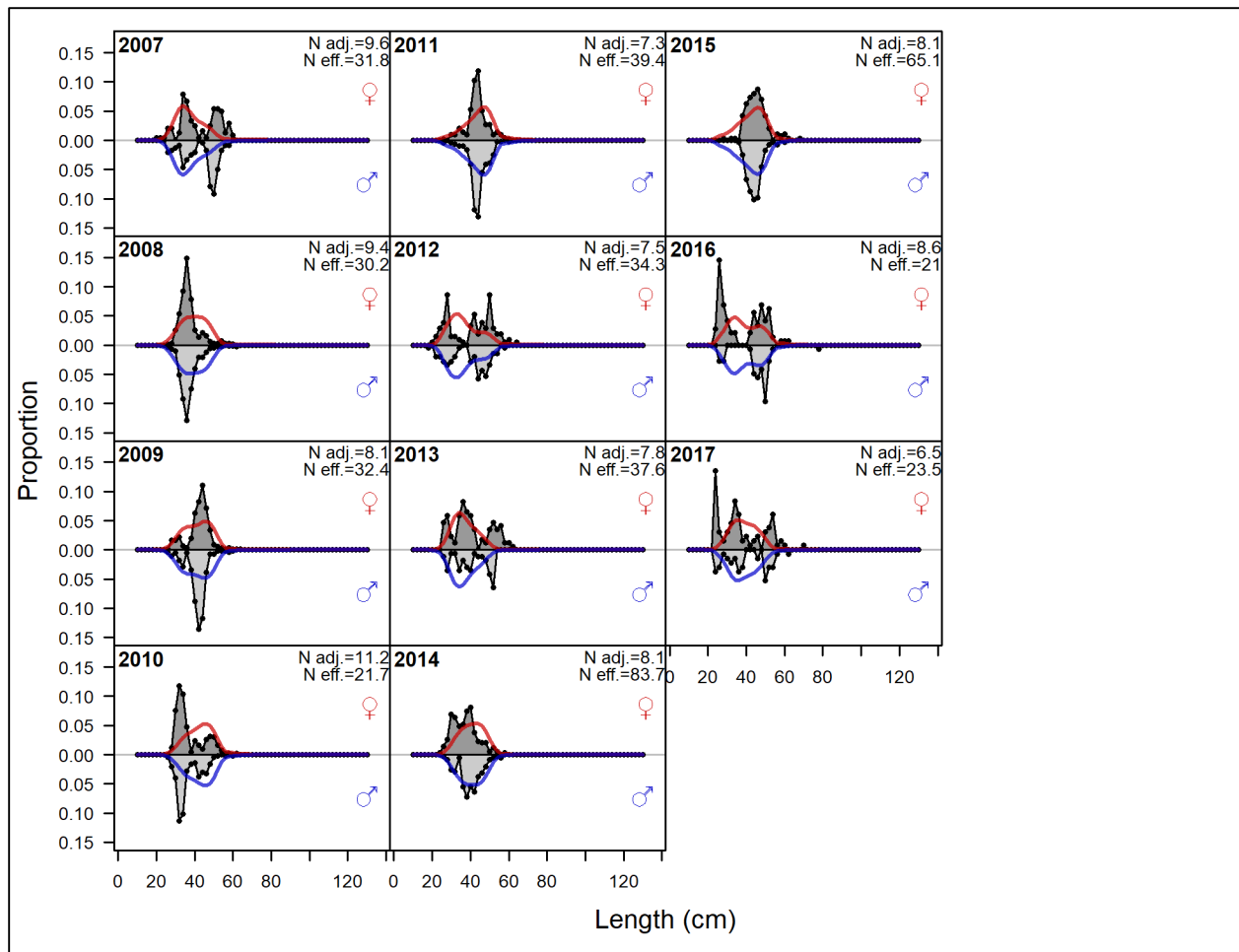
**Figure 3.21.** Observed and predicted length compositions for the RRrec harvest from the base run of the stock assessment model, 1999–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



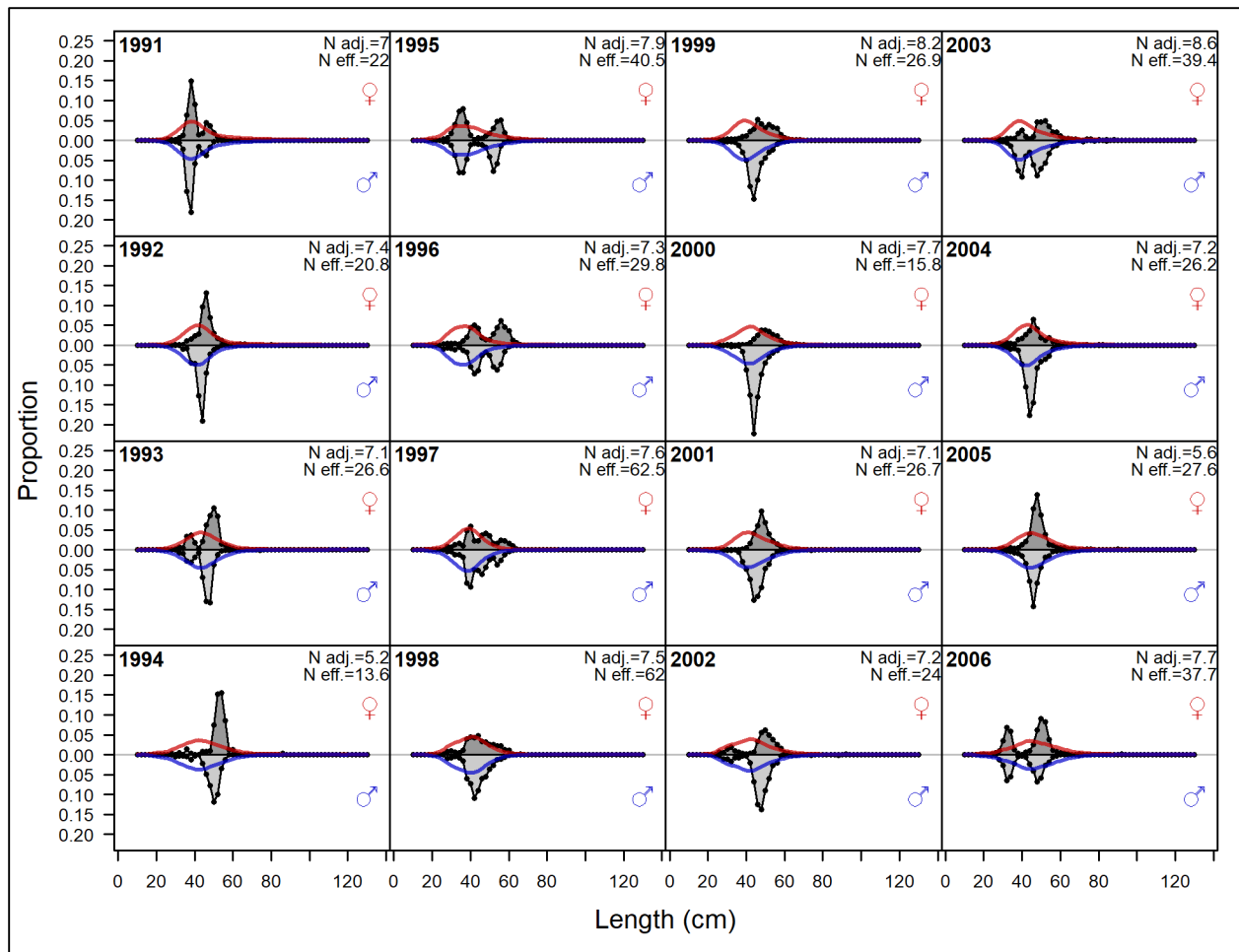
**Figure 3.22.** Observed and predicted length compositions for the RRrec discards from the base run of the stock assessment model, 2005–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



**Figure 3.23.** Observed and predicted length compositions for the P135fw survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

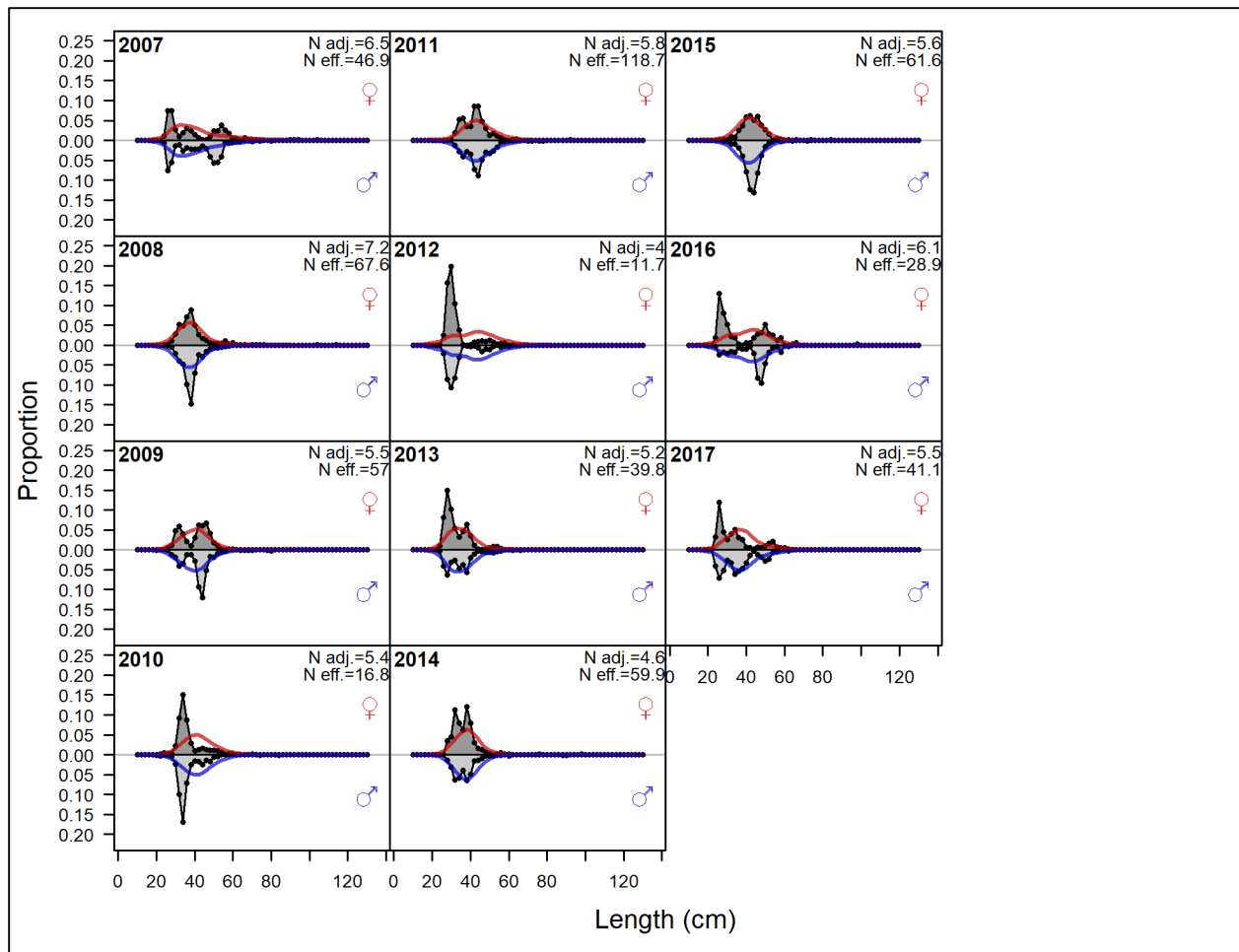


**Figure 3.24.** Observed and predicted length compositions for the P135fw survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

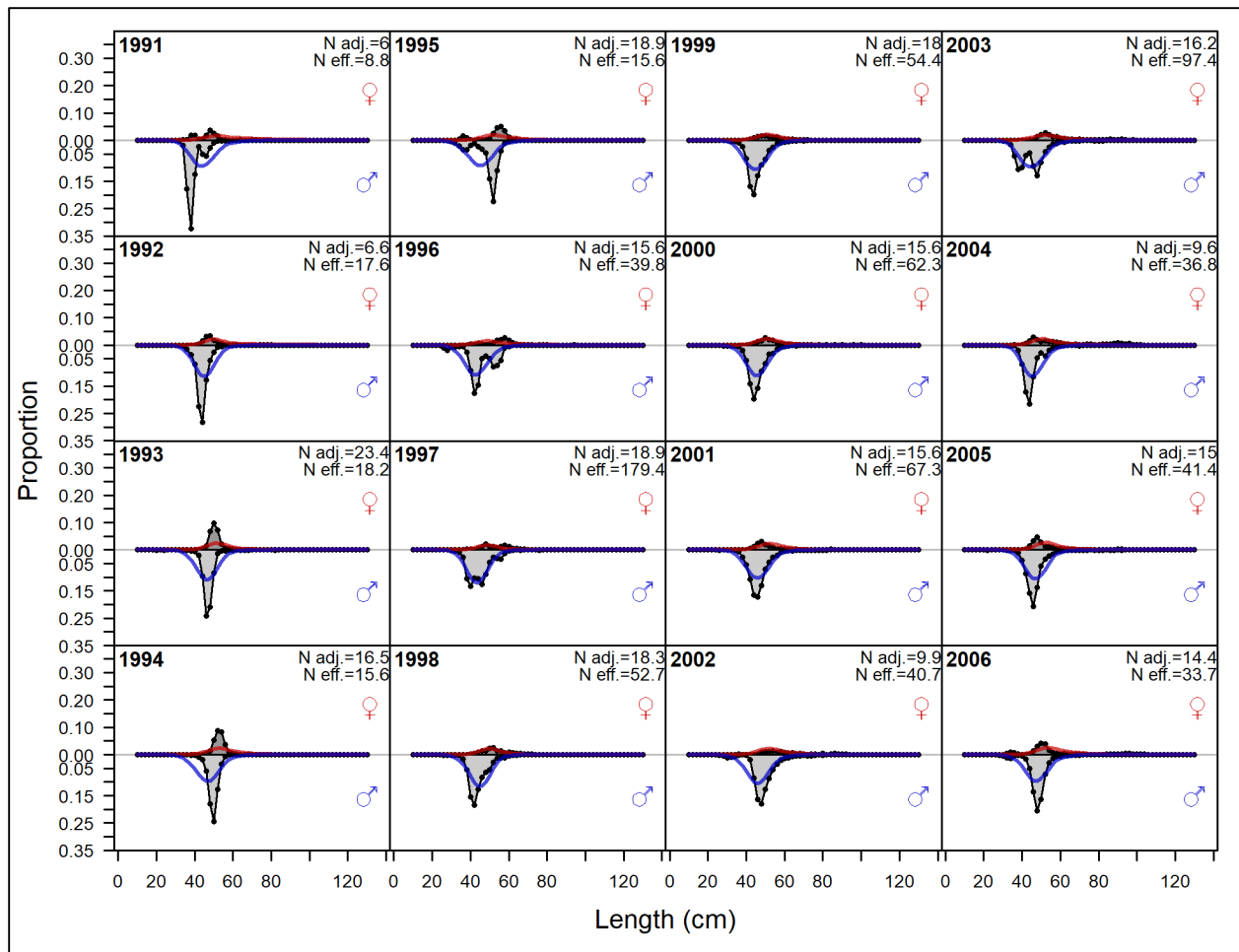


**Figure 3.25.** Observed and predicted length compositions for the P135spr survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.

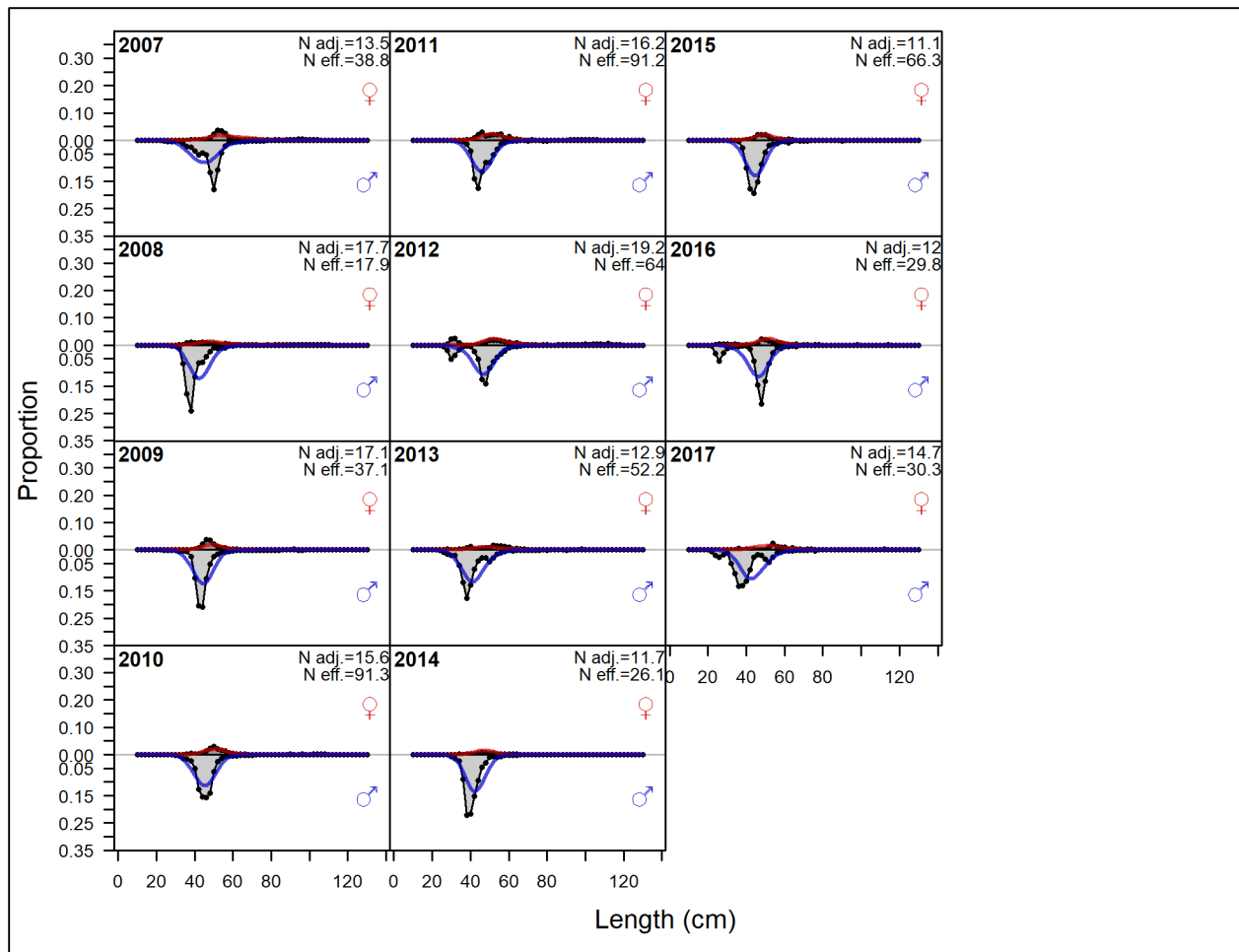




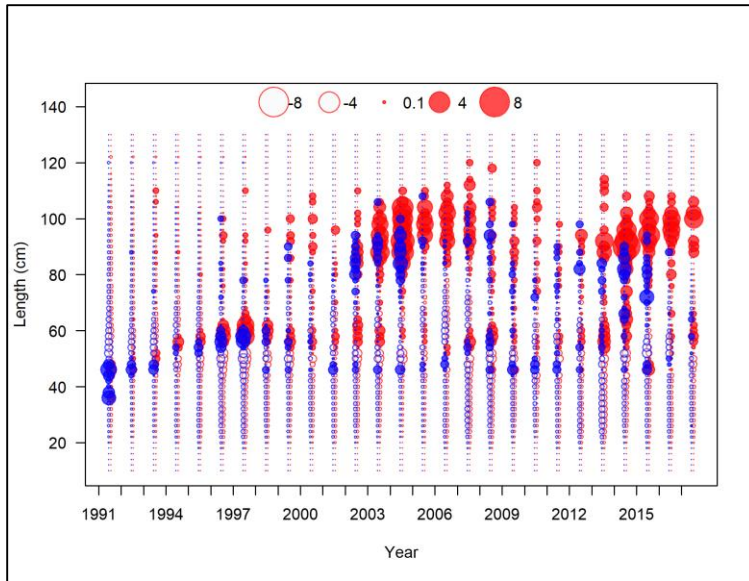
**Figure 3.26.** Observed and predicted length compositions for the P135spr survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



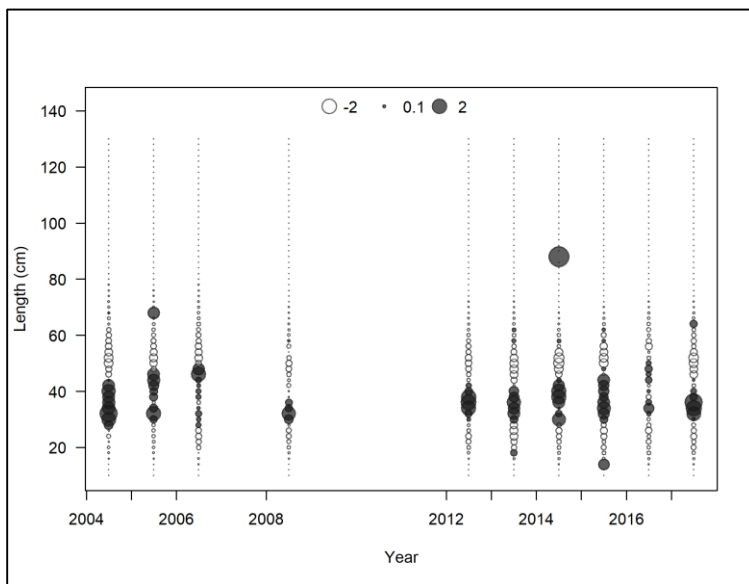
**Figure 3.27.** Observed and predicted length compositions for the RRef survey from the base run of the stock assessment model, 1991–2006. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



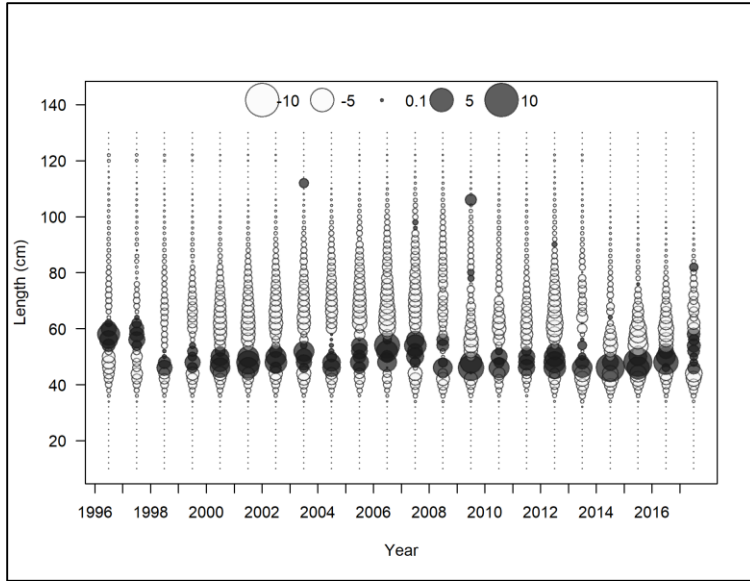
**Figure 3.28.** Observed and predicted length compositions for the RRef survey from the base run of the stock assessment model, 2007–2017. N adj. represents the input effective sample size (number of trips sampled) and N eff. represents the model estimate of effective sample size.



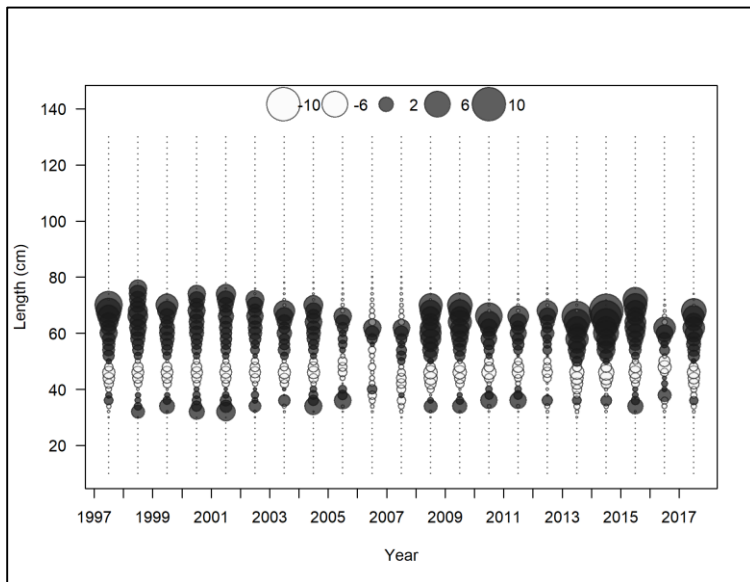
**Figure 3.29.** Pearson residuals (red: female; blue: male) from the fit of the base model run to the ARcomm landings length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



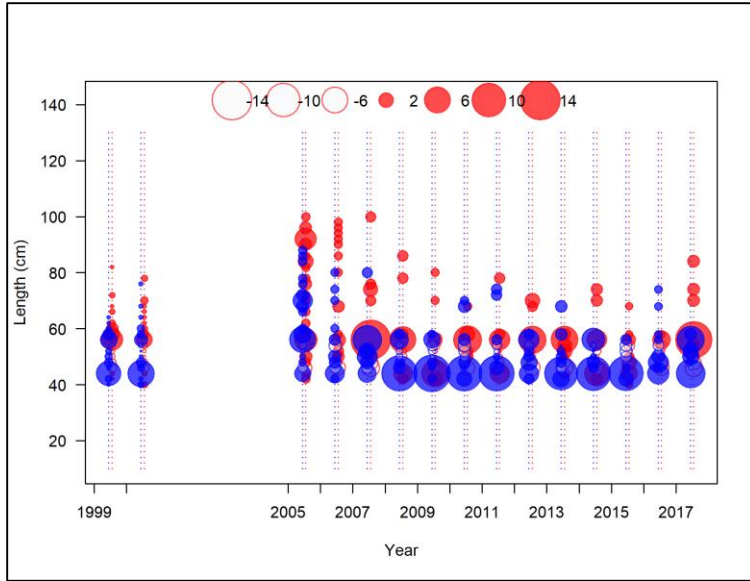
**Figure 3.30.** Pearson residuals from the fit of the base model run to the ARcomm discards length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



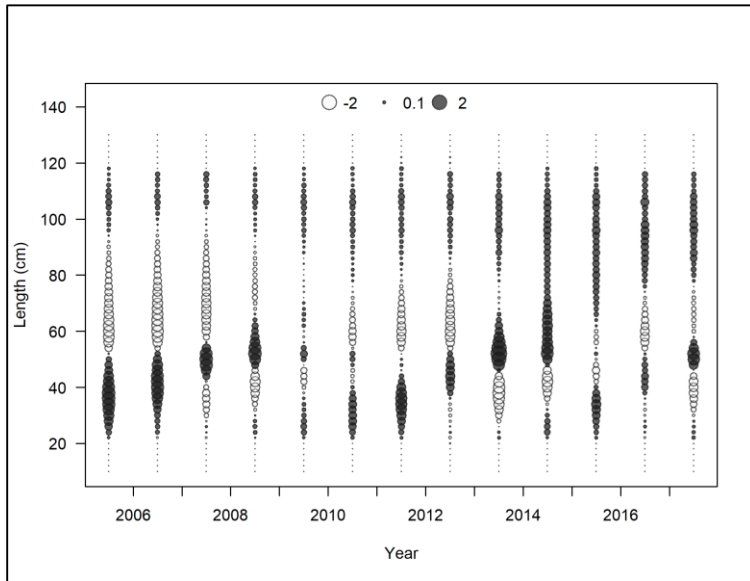
**Figure 3.31.** Pearson residuals from the fit of the base model run to the ASrec harvest length composition data, 1996–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



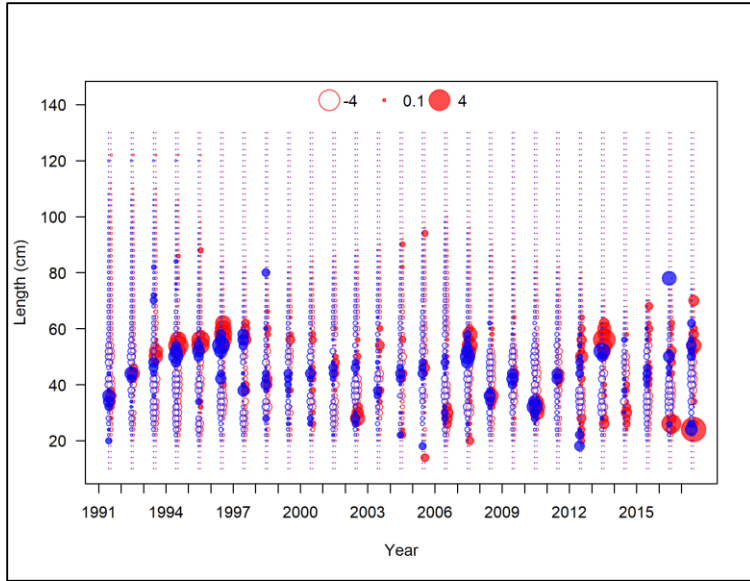
**Figure 3.32.** Pearson residuals from the fit of the base model run to the ASrec discard length composition data, 1997–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



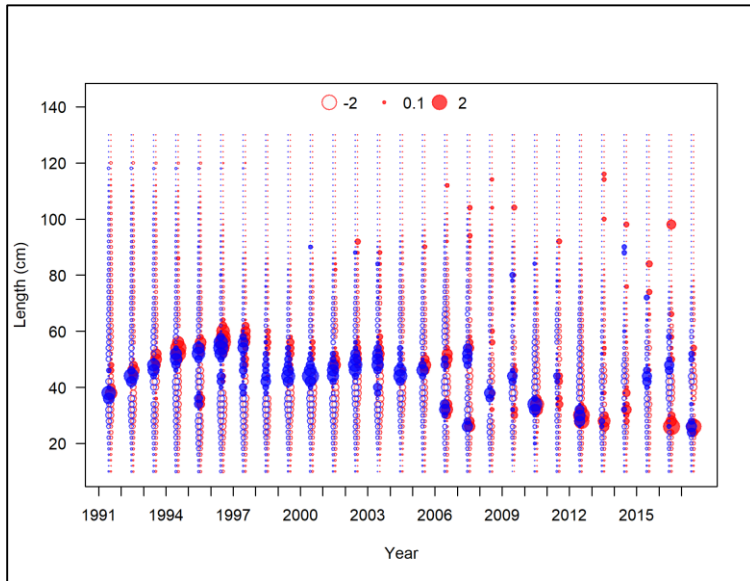
**Figure 3.33.** Pearson residuals (red: female; blue: male) from the fit of the base model run to the RRrec harvest length composition data, 1999–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



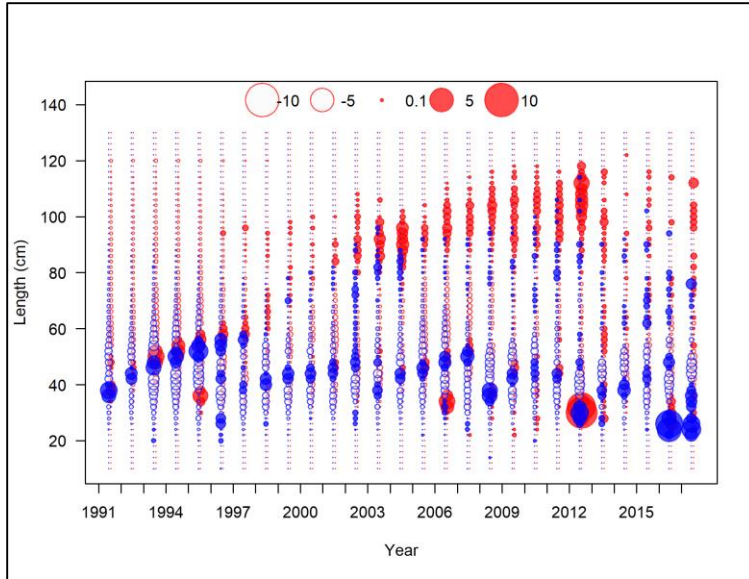
**Figure 3.34.** Pearson residuals from the fit of the base model run to the RRrec discard length composition data, 2005–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).



**Figure 3.35.** Pearson residuals (red: female; blue: male) from the fit of the base model run to the P135fw survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).

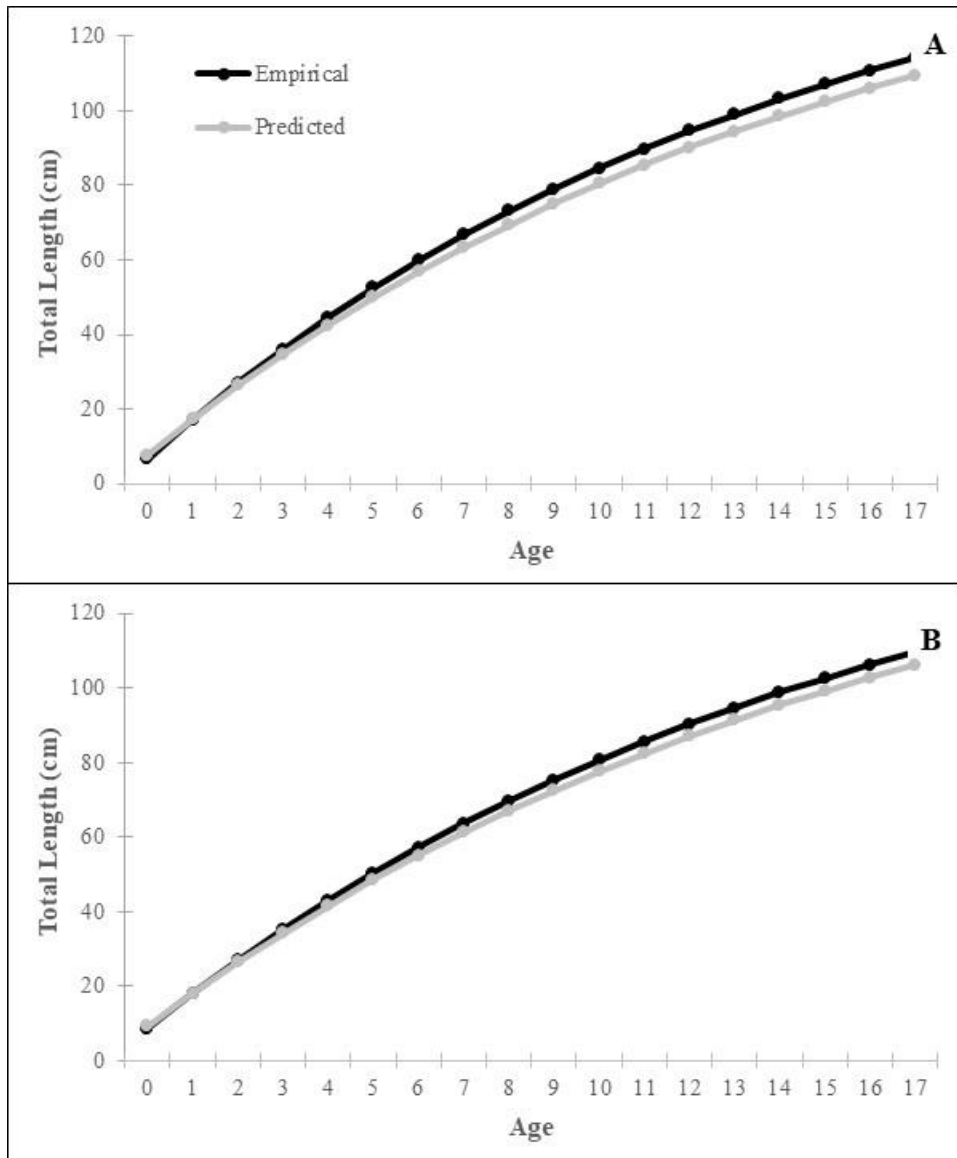


**Figure 3.36.** Pearson residuals (red: female; blue: male) from the fit of the base model run to the P135spr survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).

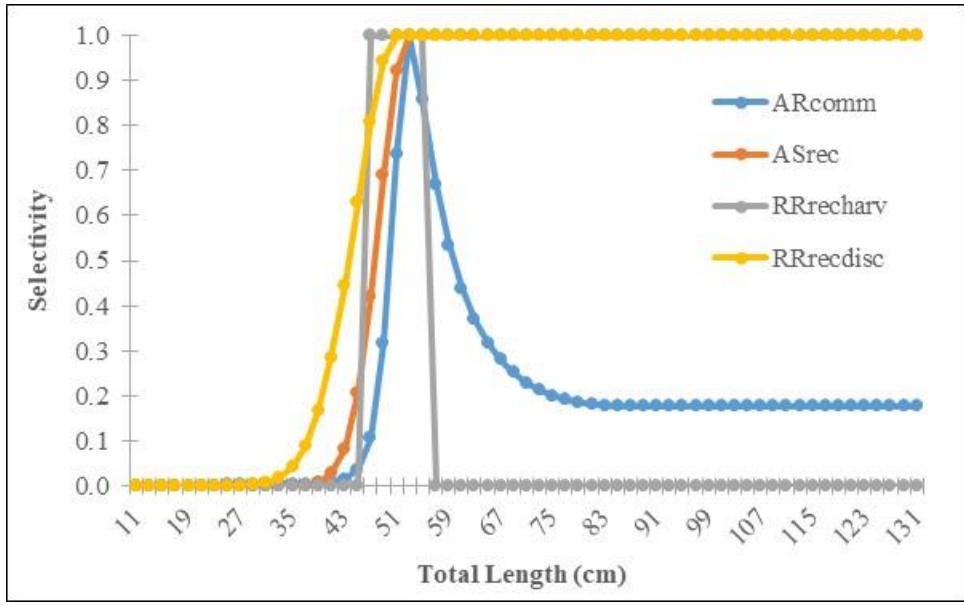


**Figure 3.37.** Pearson residuals (red: female; blue: male) from the fit of the base model run to the RRef survey length composition data, 1991–2017. Closed bubbles represent positive residuals (observed > expected) and open bubbles represent negative residuals (observed < expected).

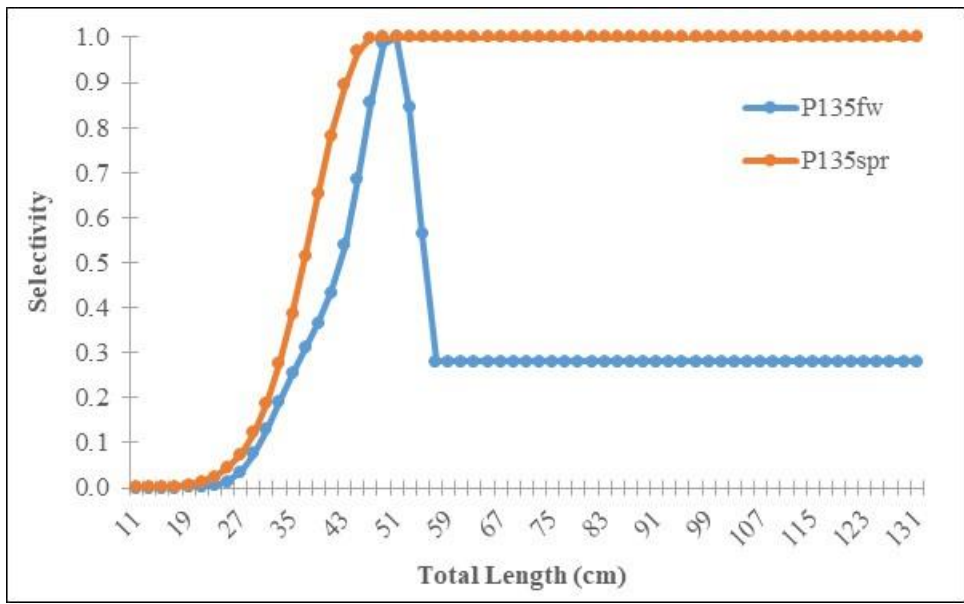




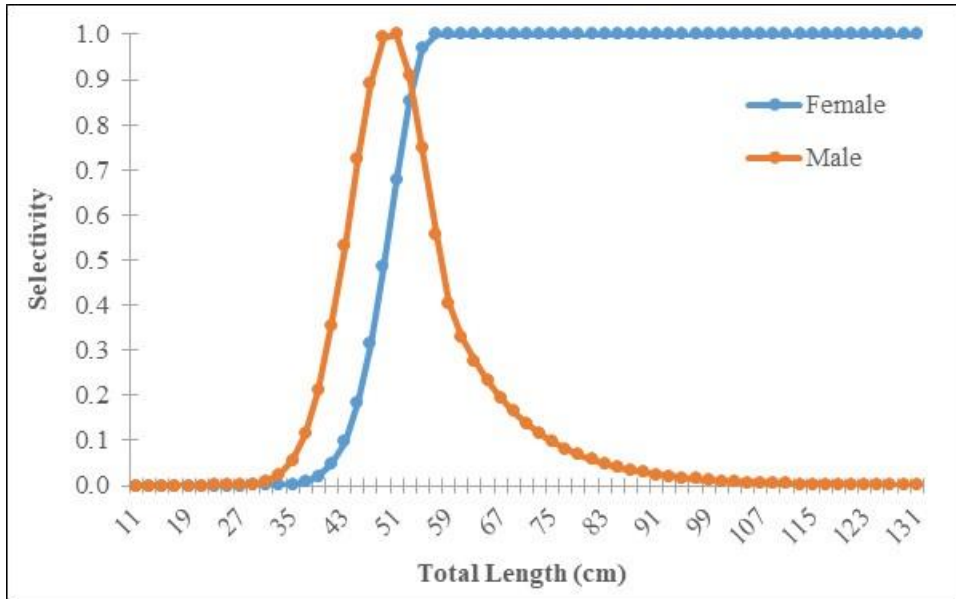
**Figure 3.38.** Comparison of empirical and model-predicted age-length growth curves for (A) female and (B) male striped bass from the base run of the stock assessment model.



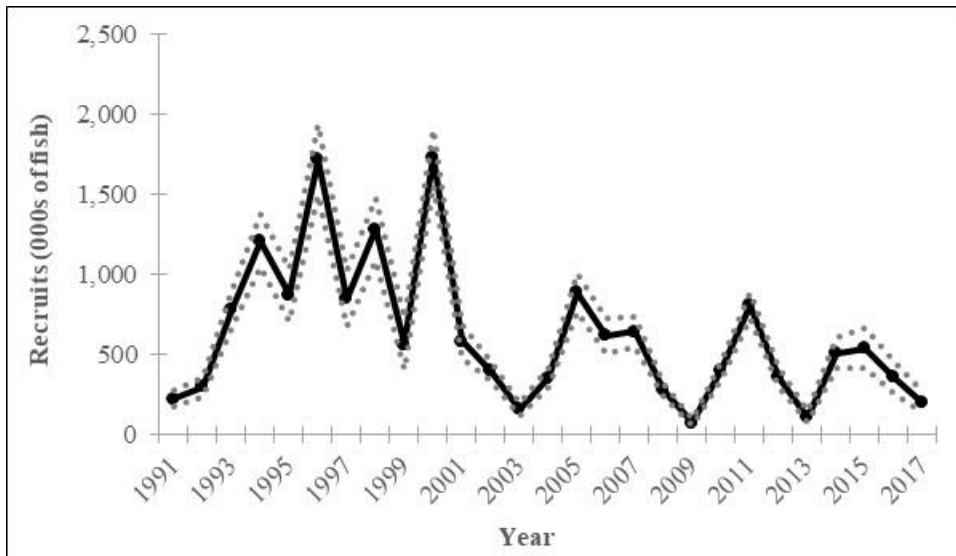
**Figure 3.39.** Predicted length-based selectivity for the fleets from the base run of the stock assessment model.



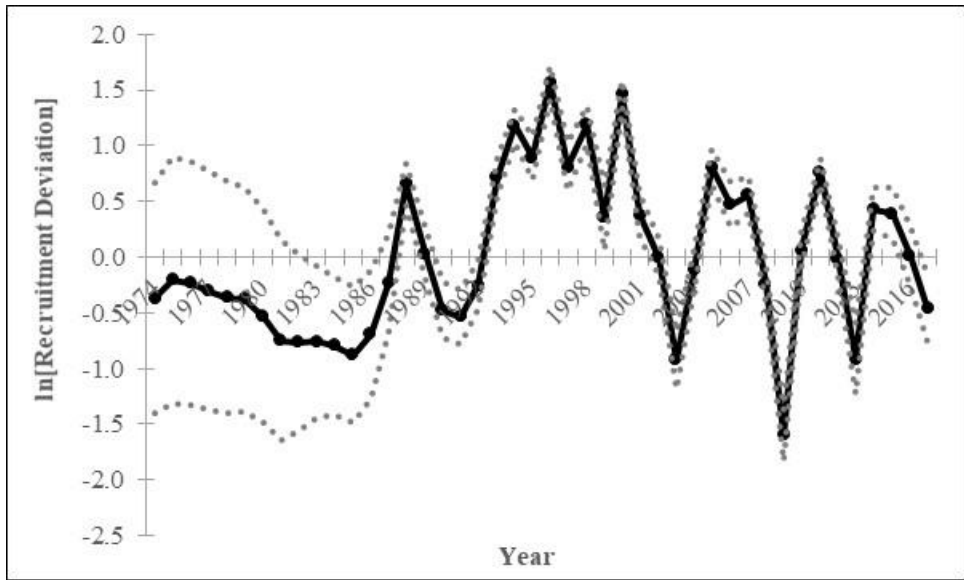
**Figure 3.40.** Predicted length-based selectivity for the P135fw and P135spr surveys from the base run of the stock assessment model.



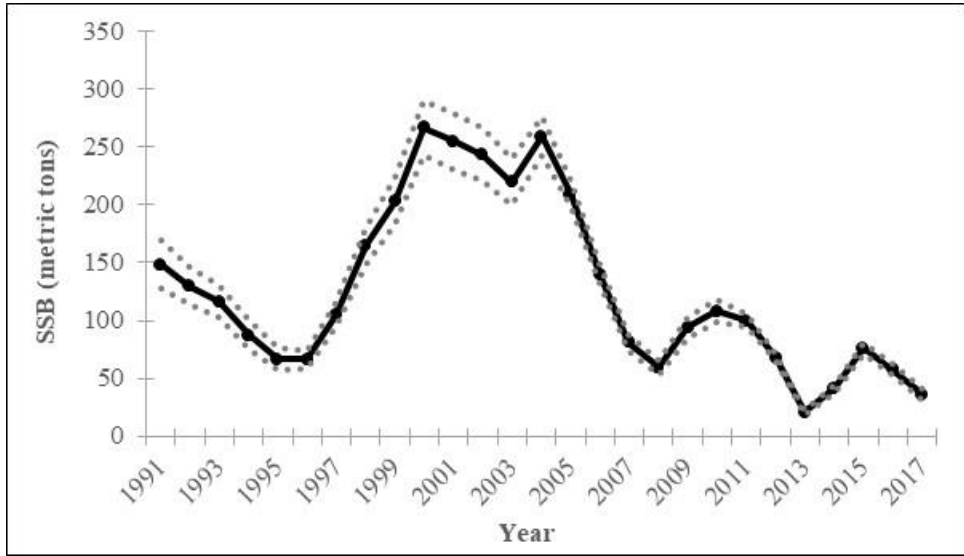
**Figure 3.41.** Predicted length-based selectivity for the RRef survey from the base run of the stock assessment model.



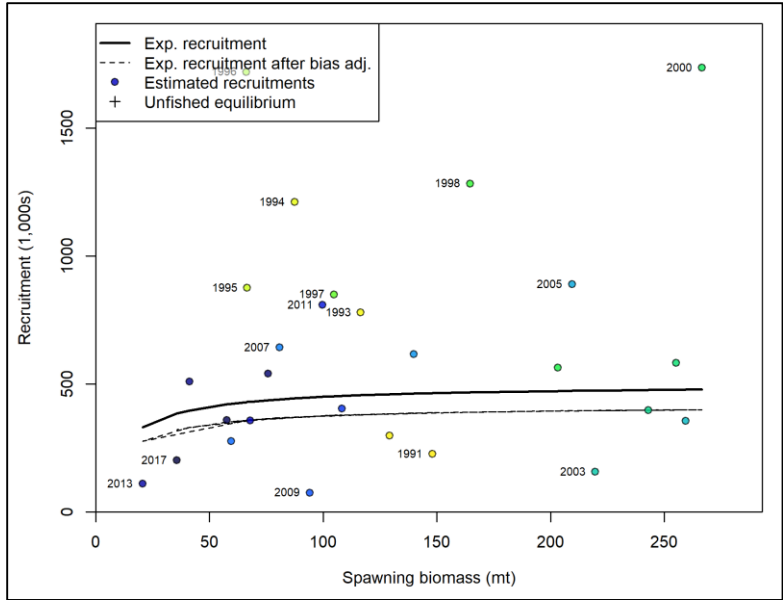
**Figure 3.42.** Predicted recruitment of age-0 fish from the base run of the stock assessment model, 1991–2017. Dotted lines represent  $\pm 2$  standard deviations of the predicted values.



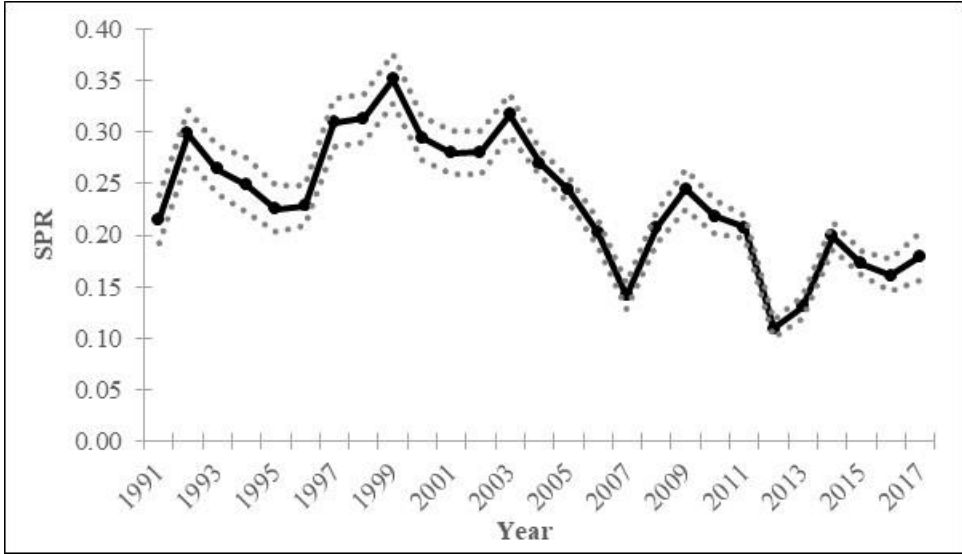
**Figure 3.43.** Predicted recruitment deviations from the base run of the stock assessment model, 1991–2017. Dotted lines represent  $\pm 2$  standard deviations of the predicted values.



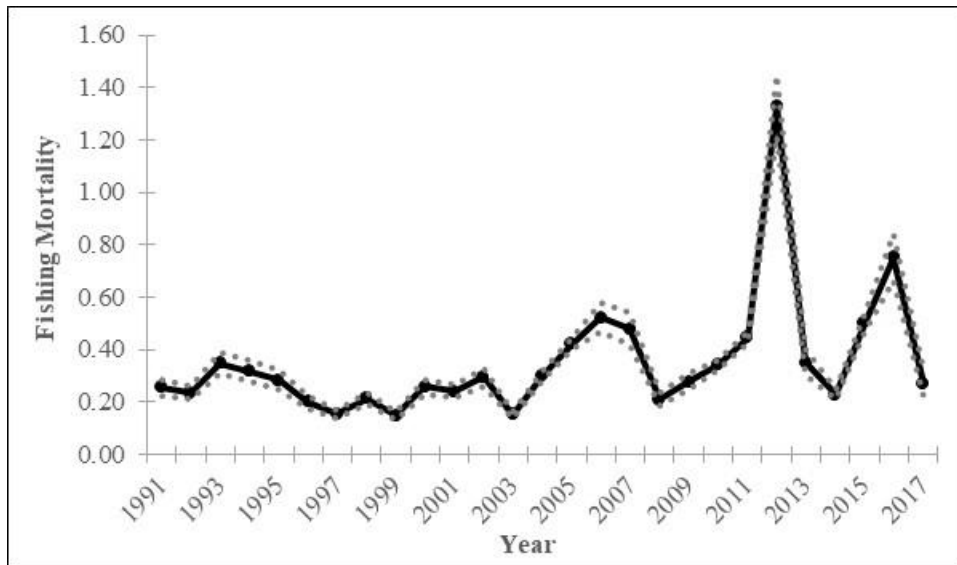
**Figure 3.44.** Predicted female spawning stock biomass from the base run of the stock assessment model, 1991–2017. Dotted lines represent  $\pm 2$  standard deviations of the predicted values.



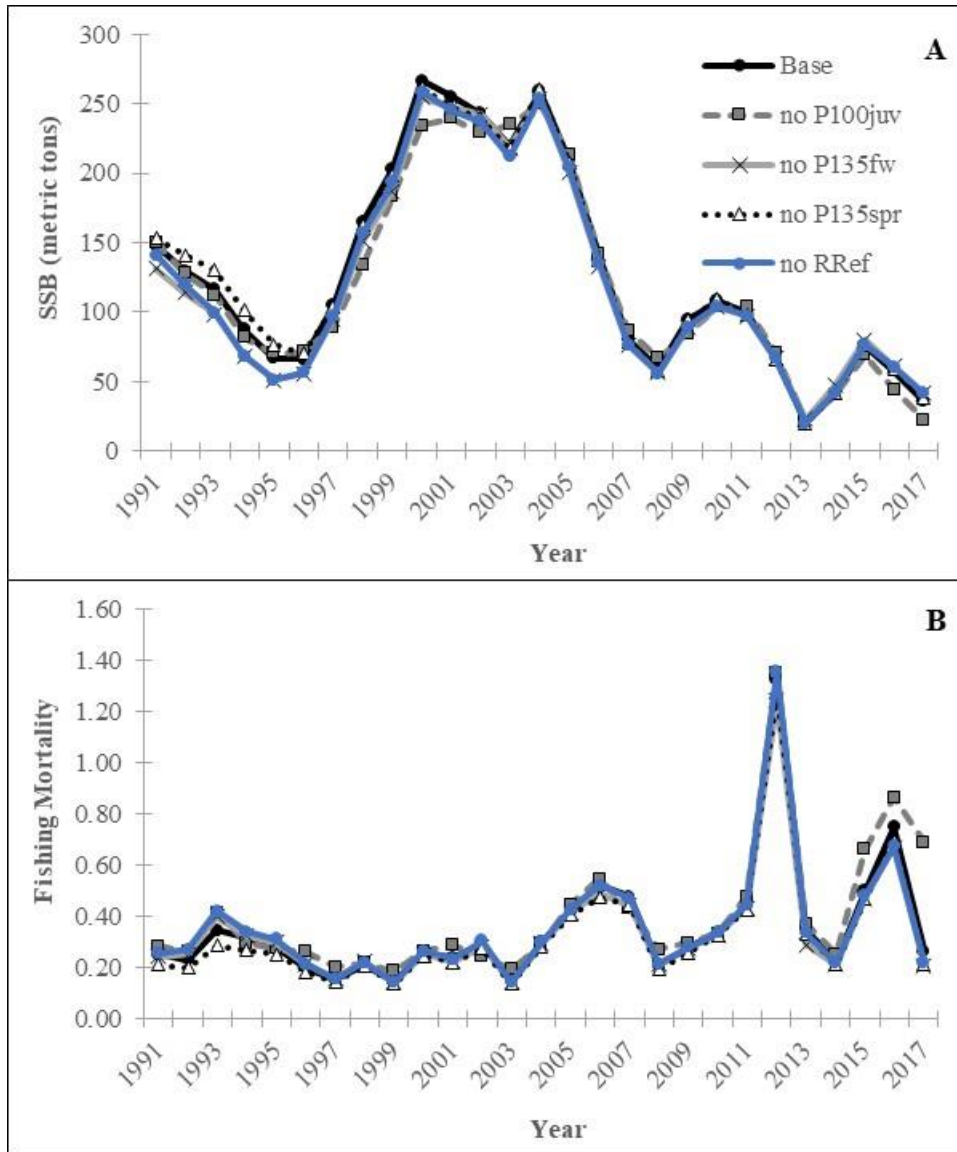
**Figure 3.45.** Predicted Beverton-Holt stock-recruitment relationship from the base run of the stock assessment model with labels on first (1991), last (2017), and years with (log) deviations > 0.5.



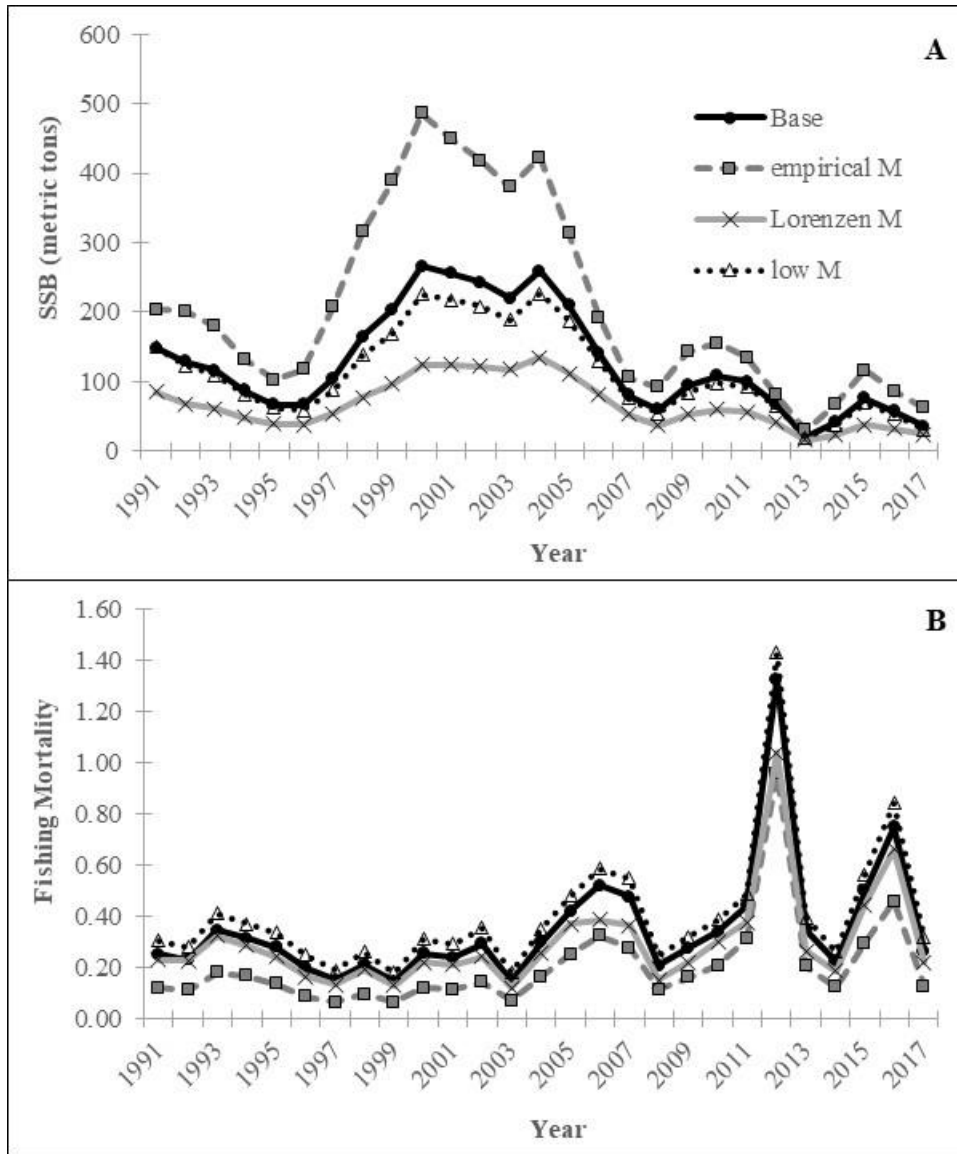
**Figure 3.46.** Predicted spawner potential ratio (SPR) from the base run of the stock assessment model, 1991–2017. Dotted lines represent  $\pm 2$  standard deviations of the predicted values.



**Figure 3.47.** Predicted fishing mortality (numbers-weighted, ages 3–5) from the base run of the stock assessment model, 1991–2017. Dotted lines represent  $\pm 2$  standard deviations of the predicted values.

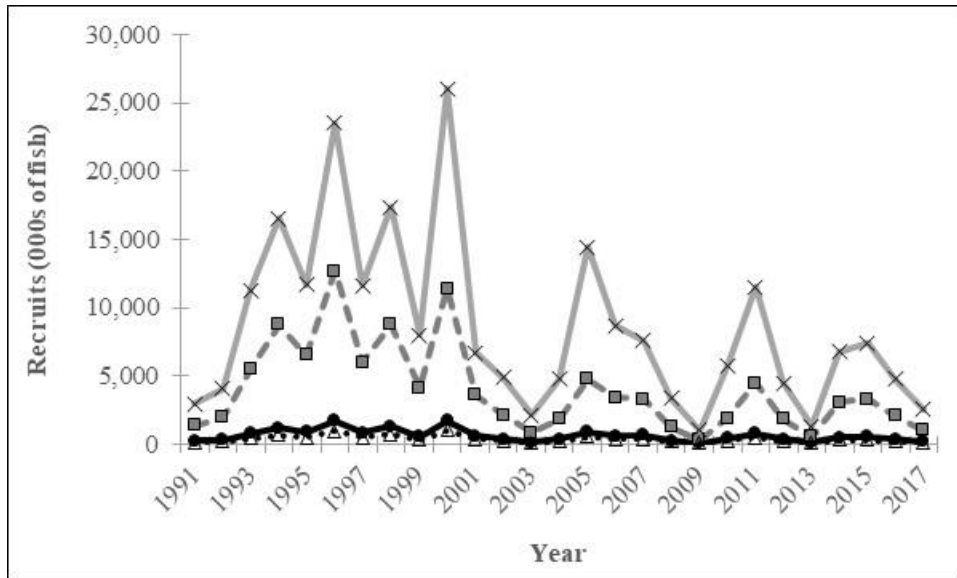


**Figure 3.48.** Sensitivity of model-predicted (A) female spawning stock biomass (SSB) and (B) fishing mortality rates (numbers-weighted, ages 3–5) to removal of different fisheries-independent survey indices from the base run of the stock assessment model, 1991–2017.

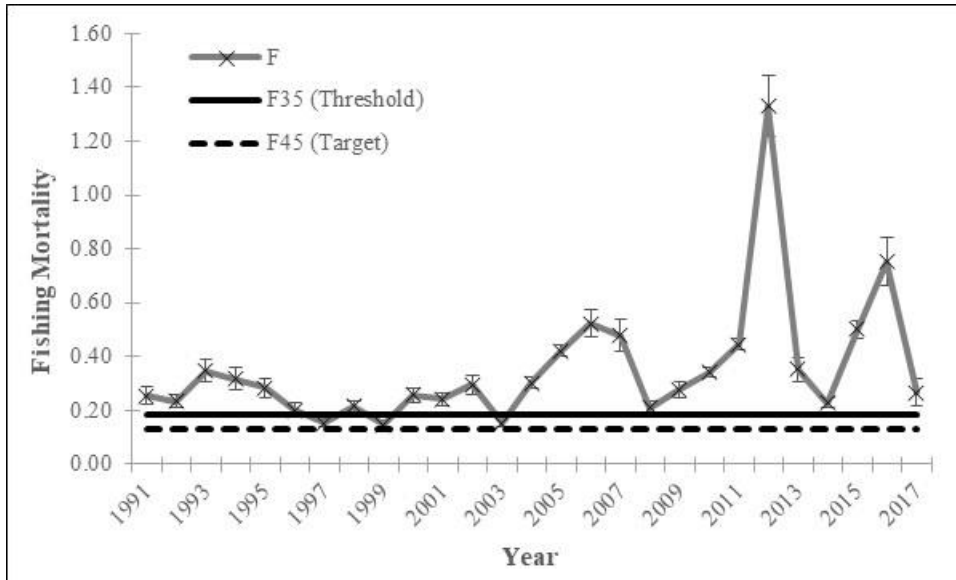


**Figure 3.49.** Sensitivity of model-predicted (A) female spawning stock biomass (SSB) and (B) fishing mortality rates (numbers-weighted, ages 3–5) to the assumption about natural mortality, 1991–2017.

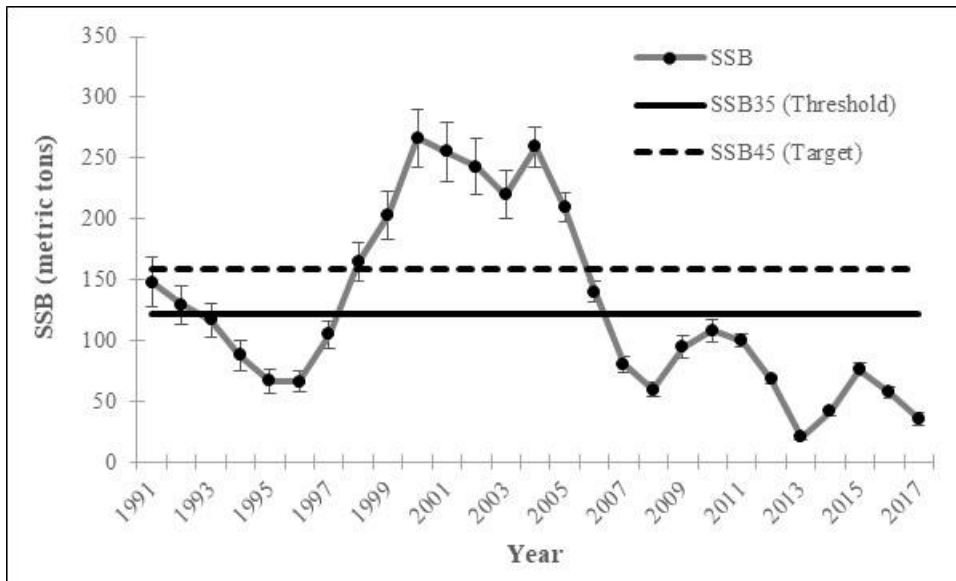




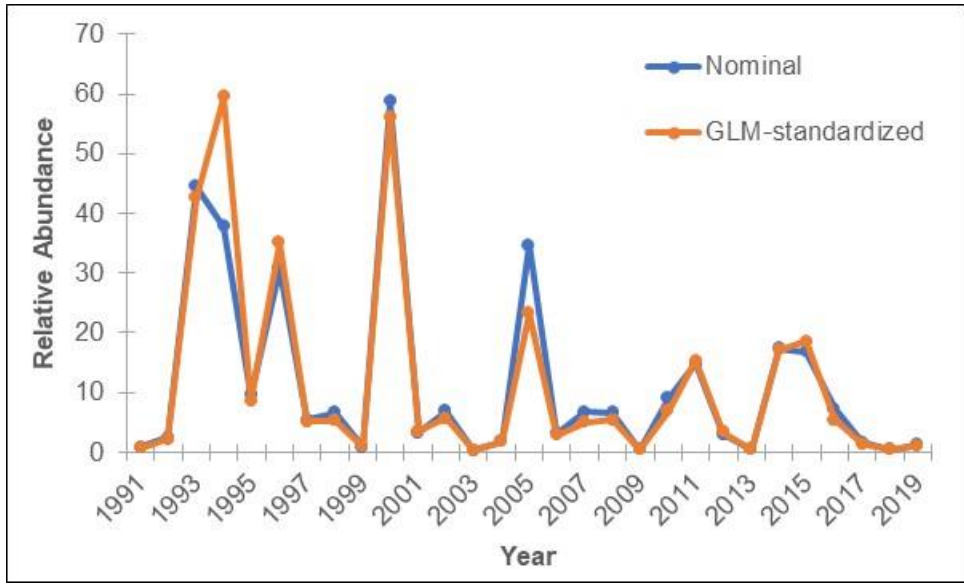
**Figure 3.50.** Predicted recruitment from the sensitivity runs in which the assumption about natural mortality was changed, 1991–2017.



**Figure 4.1.** Estimated fishing mortality (numbers-weighted, ages 3–5) compared to fishing mortality target ( $F_{45\%}=0.18$ ) and threshold ( $F_{35\%}=0.13$ ). Error bars represent  $\pm$  two standard errors.



**Figure 4.2.** Estimated female spawning stock biomass compared to spawning stock biomass target ( $SSB_{45\%}=159$  mt) and threshold ( $SSB_{35\%}=121$  mt). Error bars represent  $\pm$  two standard errors.



**Figure 5.1.** Update of the nominal and GLM-standardized indices of relative age-0 abundance derived from the Juvenile Abundance Survey (P100), 1991–2019.

## 10 APPENDIX

### **Addendum to the External Peer Review Report for the 2019 Stock Assessment of the Albemarle Sound-Roanoke River Striped Bass in North Carolina**

The SAT was able to satisfactorily resolve several of the RP's concerns in the original base model reviewed during the December 2019 workshop. The growth functions fit to observed length-at-age data external to the assessment model to generate starting values for the assessment model (i.e., empirical growth estimates) showed improved fits to the data and the growth functions predicted by the revised assessment model were more consistent with the empirical growth estimates, particularly for males. Residual patterning from fits to the length composition data in the revised assessment model are still present indicating some model misspecification, but were generally reduced. The corrected P135 indices were more consistent with the decline in recent years observed during the RRef survey, reducing some conflict the original base model was forced to reconcile. It's important to note that the revised model overestimated the index values for both P135 indices and the RRef index during the last three years of the time series, indicating the abundance estimates may still be biased high in these recent years. However, the consistent overfished status determination estimated across the revised model and natural mortality sensitivity runs (see below) lessen this concern.

The revised base model specified an age- and sex-constant natural mortality of 0.72 based on Harris and Hightower (2017). The RP still believes the empirical natural mortality estimates from Harris and Hightower (2017) are higher than reality and suggested sensitivity runs exploring the effects of lower natural mortality rates. The RP was less concerned with variation in natural mortality-at-age, as this can be less influential on parameter bias (Deroba and Schueller 2013) and because model insensitivity to age-specific natural mortality was demonstrated by the SAT in the revised report, and more interested in effects of lower natural mortality for all ages. Therefore, various age-constant life history-based natural mortality estimators were applied to the striped bass data. Ultimately, the Alverson and Carney (1975), Hoenig (1983), and Cubillios et al. (1999) estimators were included in sensitivity runs because they estimated high (relative to the other life history-based estimators, but lower than Harris and Hightower 2017 estimates), moderate, and low natural mortality rates, respectively. Additionally, an average across the estimators, which was slightly lower than the Hoenig (1983) rate, was included in the sensitivity analysis. The SAT conducted a thorough sensitivity analysis of natural mortality with model configurations that included sex-specific and sex-aggregate natural mortality rates with growth fixed or estimated. The sensitivity runs that converged on a solution produced some differences in the scale of estimates, but similar stock trajectories, particularly since the decline in SSB in the mid-2000s (Figures 1-3). The various natural mortality rates had the greatest effect on age-0 recruitment as the model needs to estimate higher recruitment under high mortality scenarios to match the data on subsequent ages that are vulnerable to the fisheries. All sensitivity runs indicated the stock was overfished and experiencing overfishing in the terminal year (Table 1).

The SAT recommended the model with a high, sex-aggregate natural mortality ( $M=0.40$ ) as the most appropriate to acknowledge estimates from established life history-based methods, but also the higher empirical rates estimated directly from the striped bass population by Harris and Hightower (2017). A sex-aggregate natural mortality rate is consistent with the similar growth

estimated between sexes from the available data. Further, a subsequent sensitivity run requested by the RP showed this model configuration is not sensitive to excluding the RRef survey data, as was a primary concern with the original base model. The RP agrees with the SAT's recommendation and recommends this model be used for management advice. The population trajectory and overfished and overfishing stock status estimates from this model are consistent with the available data sets that show poor recruitment in recent years, declining abundance to historically low levels, and a truncated age structure.

#### Needs for Future Assessments

The RP along with the SAT were collectively concerned about declining recruitment in the time series. One key uncertainty identified in this review is to incorporate the effects of changes in river flow on recruitment. It appears that substantial data exists, but they have not yet been incorporated into the stock assessment. Future assessments should consider key environmental drivers of recruitment such as river flow, because declining recruitment in the time series does not appear to result solely from reduced abundance due to harvest. The RP suggests that future assessments should incorporate flow-recruitment relationships into the stock assessment formally to understand how spring flow conditions influence recruitment and ultimately stock abundance. Another potential influence on the striped bass stock is the prevalence of non-native catfishes, primarily blue catfish *Ictalurus furcatus* and flathead catfish *Pylodictis olivaris*. Both species occur in North Carolina river systems and it seems the blue catfish population is expanding in the Roanoke River and Albemarle Sound areas. Predation by catfishes could potentially impact recruitment of striped bass directly, or could influence food resources for striped bass through competition for prey (e.g., Pine et al. 2005). The degree to which this occurs is not known, but future assessments should consider this as a factor that may influence abundance and is not tied to striped bass harvest.

Moderate and evident differences in growth (Figures 1.2 and 1.3, main report) are not resolved within the model. The effect on estimation of sex-specific  $M$  are not readily quantifiable at present. Factors potentially contributing to the poor resolution of male and female growth trajectories, as estimated by the von Bertalanffy growth function, include under-representation of older age classes and lack of sex-specific length data for Ages 0 to 2<sup>+</sup> year old fish. The RP accordingly encourages collection of sex-specific length-at-age data from juveniles (ages 0–2) and as well from older fish to better inform growth estimates.

## **References**

- Alverson, D.L., and M.J. Carney. 1975. A graphic review of the growth and decay of population cohorts. *Journal du Conseil international pour l'Exploration de la Mer* 36(2):133–143.
- Cubillos, L.A., R. Alarocan, and A. Brante. 1999. Empirical estimates of natural mortality for Chilean hake (*Merluccius gayi*): evaluation of precision. *Fisheries Research* 42:147–153.
- Deroba, J.J., and A.M. Schueller. 2013. Performance of stock assessments with misspecified age- and time-varying natural mortality. *Fisheries Research* 46:27–40.
- Hoenig, J. 1983. Empirical use of longevity data to estimate mortality rates. *Fishery Bulletin* 81(4): 898–903.
- Pine, W.E., T.J. Kwak, D.S. Waters, and J.A. Rice. 2005. Diet selectivity of introduced Flathead catfish in coastal rivers. *Transactions of the American Fisheries Society* 134:901–909.

## Tables

Table 1. Specified natural mortality, terminal year and threshold model estimates, and stock status across the revised base model (Baseline) and natural mortality sensitivity runs. The RP recommends the “highMsamesex (est growth)” run be used for a management advice.

Scenario	M (yr <sup>-1</sup> )	Current year (2017)		Threshold		Overfished	Overfishing	Reference
		SSB (mt)	F (yr <sup>-1</sup> )	SSB <sub>35%</sub> (mt)	F <sub>35%</sub> (yr <sup>-1</sup> )			
Baseline	0.72	62	0.13	89	0.43	Y	N	Harris and Hightower, 2017
avgM (est growth)	0.23F, 0.25M	30.80	0.35	283.88	0.12	Y	Y	
avgM (fix growth)	0.23F, 0.25M	47.46	0.28	153.20	0.13	Y	Y	
midM (fix growth)	0.25F, 0.28M	42.79	0.29	114.46	0.14	Y	Y	Hoening 1983
highM (fix growth)	0.37F, 0.44M	40.22	0.31	182.06	0.19	Y	Y	Alverson and Camey 1975
highMsamesex (est growth)	0.40	35.64	0.27	121.29	0.18	Y	Y	Alverson and Camey 1975
avgMsamesex (est growth)	0.24	32.91	0.28	150.77	0.11	Y	Y	

**Figures**

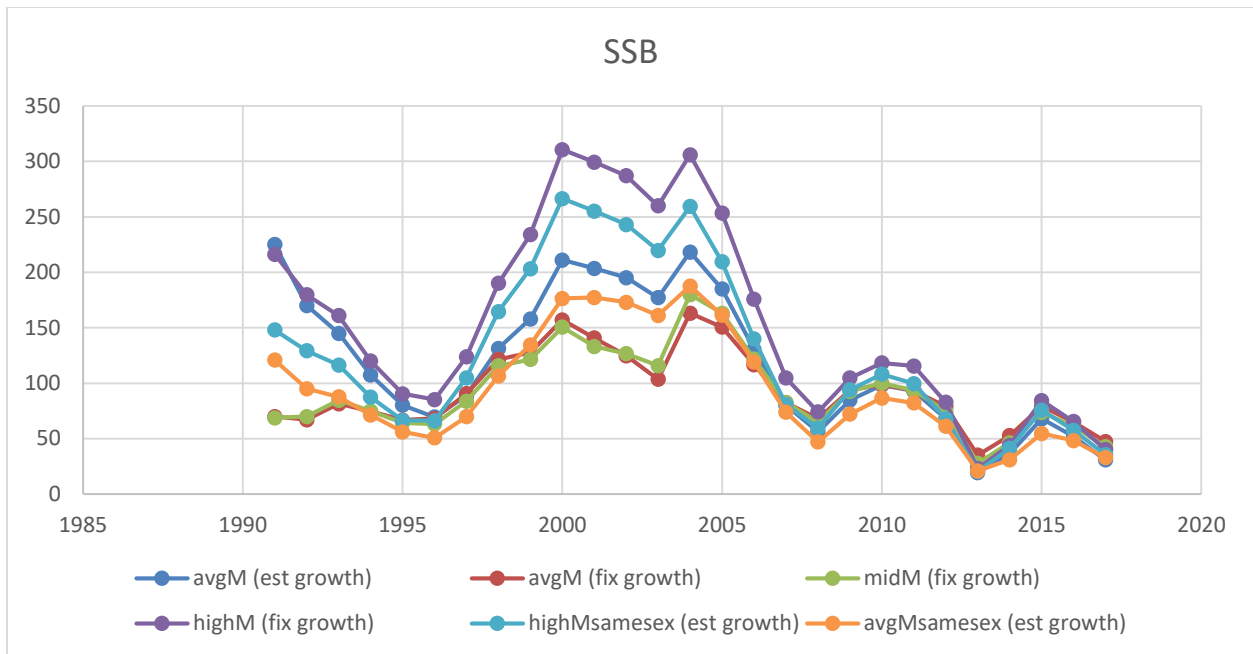


Figure 1. Female spawning stock biomass estimates (metric tons) across natural mortality sensitivity runs.

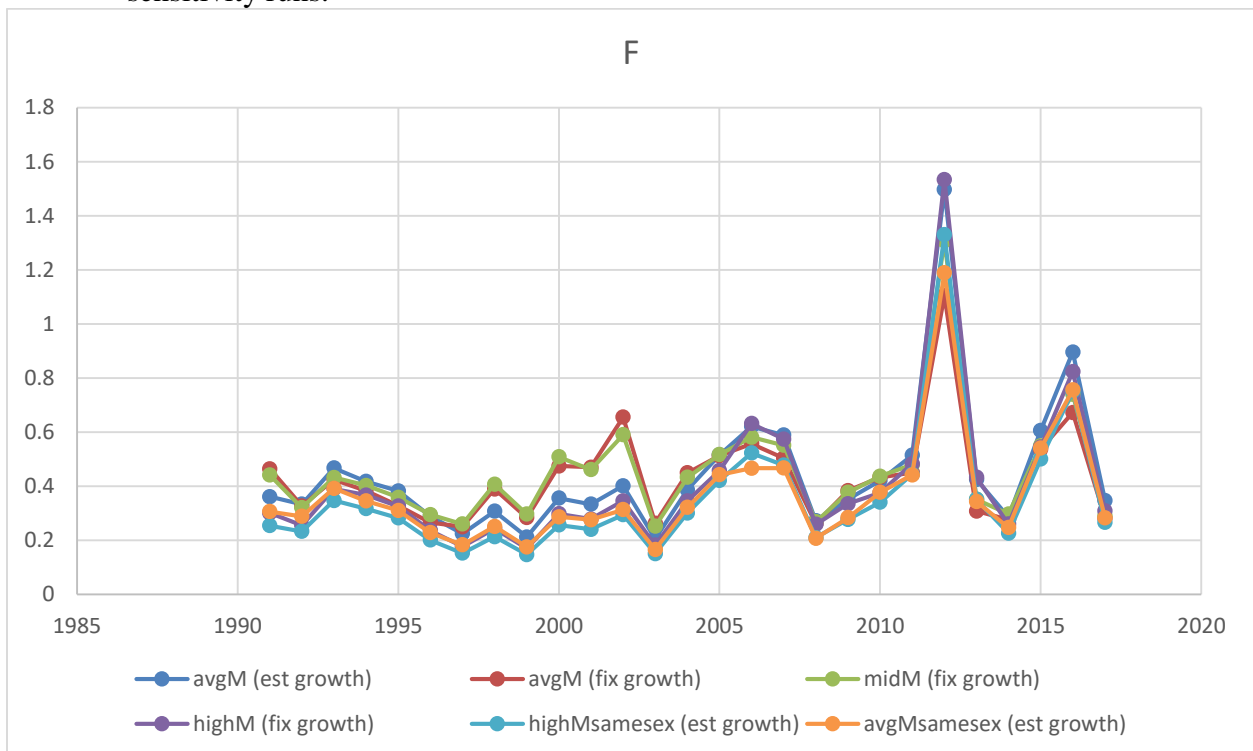


Figure 2. Numbers-weighted ages 3-5 average fishing mortality estimates across natural mortality sensitivity runs.



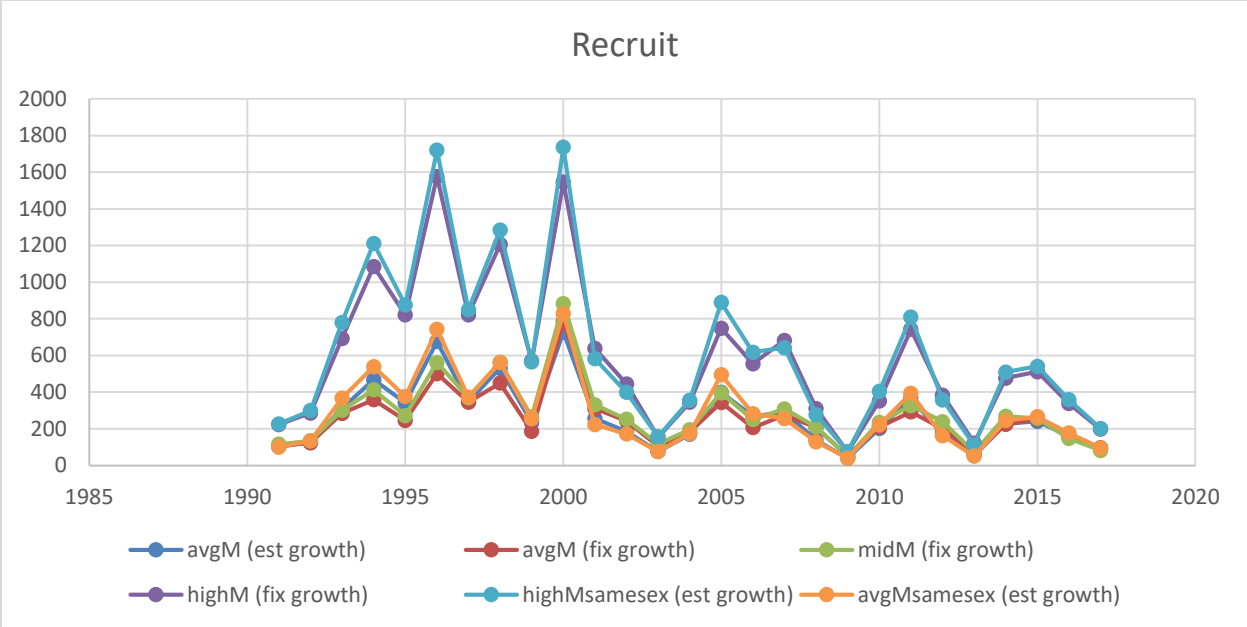


Figure 3. Age-0 recruitment estimates (thousands) across natural mortality sensitivity runs.

**November 2020 Revision**  
to  
**Amendment 1**  
to the  
**North Carolina Estuarine Striped Bass  
Fishery Management Plan**

Prepared By The

North Carolina Department of Environmental Quality  
Division of Marine Fisheries  
3441 Arendell Street  
P.O. Box 769  
Morehead City, NC 28557

and

North Carolina Wildlife Resources Commission  
Inland Fisheries Division  
1751 Varsity Drive  
Raleigh, NC 27606



**November 2020 Revision**  
**to**  
**Amendment 1**  
**to the**  
**North Carolina Estuarine Striped Bass**  
**Fishery Management Plan**

Effective Jan. 1, 2021

**I. ISSUE**

Requirement to reduce the striped bass total allowable landings (TAL) in the Albemarle Sound and Roanoke River Management Areas to remain in compliance with Amendment 1 to the North Carolina Estuarine Striped Bass Fishery Management Plan (FMP) and the Atlantic States Marine Fisheries Commission (ASMFC) Addendum IV to Amendment 6 to the Interstate FMP for Atlantic Striped Bass. The reduction in TAL is required based on results of the 2020 Albemarle-Roanoke (A-R) striped bass benchmark stock assessment that indicates the stock is overfished and overfishing is occurring in the terminal year (2017) of the assessment (Lee et al. 2020).

**II. ORIGINATION**

North Carolina Division of Marine Fisheries (NCDMF) staff and North Carolina Wildlife Resources Commission (NCWRC), Inland Fisheries Division staff.

**III. BACKGROUND**

Atlantic striped bass from Maine through North Carolina are managed under the jurisdiction of the ASMFC since Congress passed the Atlantic Striped Bass Conservation Act in 1984. The A-R striped bass stock is migratory at older ages but contributes minimally to the overall Atlantic striped bass stock complex compared to the Chesapeake Bay, Delaware, and Hudson stocks (ASMFC 2003; Berggren and Lieberman 1978; Callihan et al. 2014). Due to the non-migratory behavior of striped bass stocks south of the Albemarle Sound Management Area (ASMA), the striped bass stocks within the Central Southern Management Area (CSMA) are not included in ASMFC's Interstate FMP for Atlantic striped bass.

The ASMFC Atlantic Striped Bass Management Board approved Addendum IV to Amendment 6 to the Interstate FMP for Atlantic Striped Bass in October 2014 (ASMFC 2014). Through this addendum the ASMFC Atlantic Striped Bass Technical Committee determined it was most biologically appropriate to use NCDMF's A-R stock assessment to determine appropriate fishing mortality ( $F$ ) and spawning stock biomass (SSB) biological reference points (BRPs) specifically for the A-R stock rather than using the same BRPs as the Chesapeake Bay.

Future A-R benchmark stock assessments and updates will recalculate BRPs accordingly based on additional years of harvest, discard data, and indices of relative abundance added to the model. The ASMFC Atlantic Striped Bass Technical Committee and Management Board will continue to review each NCDMF A-R striped bass benchmark stock assessment for approval for management use as a point of compliance.

The 2020 A-R striped bass benchmark stock assessment was conducted to inform development of Amendment 2 to the North Carolina Estuarine Striped Bass FMP, which is currently underway. The A-R stock assessment is periodically undertaken for management purposes to reassess the stock status relative to the BRPs. This is generally undertaken when the ASMFC Striped Bass Technical Committee assesses the coast-wide stock or when the NCDMF initiates an amendment to the North Carolina Estuarine Striped Bass FMP.

The 2020 A-R striped bass benchmark stock assessment was completed in August 2020 (Lee et al. 2020). The assessment went through a multi-day peer review process in which NCDMF staff presented the assessment to three external experts on striped bass and marine fisheries modeling techniques. The external peer review is the standard process to review marine fisheries stock assessments throughout the world. The 2020 benchmark assessment was approved for management use by the peer reviewers for at least the next five years. The NCDMF also approved it for management use.

Results from the 2020 benchmark assessment indicate the A-R striped bass stock is overfished and overfishing is occurring relative to the updated BRPs, which are based on spawning potential ratio (SPR) thresholds of  $F_{35\%SPR}$  and  $SSB_{35\%SPR}$  and targets of  $F_{45\%SPR}$  and  $SSB_{45\%SPR}$  (Table 1) (Lee et al. 2020). The  $F$  estimate in the terminal year (2017) of the assessment was 0.27, above the  $F_{35\%SPR}$  Threshold of 0.18, meaning overfishing is occurring. Female SSB was estimated at 78,576 lb, below the  $SSB_{35\%SPR}$  Threshold of 267,390 lb, indicating the stock is overfished (Table 1). Adaptive management measures in Amendment 1 to the North Carolina Estuarine Striped Bass FMP (NCDMF 2013) are a mechanism to maintain a sustainable harvest. Sustainable harvest is defined in North Carolina General Statute 113-129(14a) as “the amount of fish that can be taken from a fishery on a continuing basis without reducing the stock biomass of the fishery or causing the fishery to become overfished.” With overfishing occurring in the terminal year of the assessment (2017), adaptive management measures contained in Amendment 1 are required to be implemented to reduce the TAL to a level that is projected to lower  $F$  to the  $F_{45\%SPR}$  Target, a 47.6 % reduction in  $F$  (Table 1) (NCDMF 2013). This action maintains compliance with Amendment 1 to the North Carolina Estuarine Striped Bass FMP and ASMFC’s Addendum IV to Amendment 6 to the Interstate FMP for Atlantic Striped Bass.

Until adoption of Amendment 2 or another revision, the A-R striped bass stock is managed through Amendment 1 to the North Carolina Estuarine Striped Bass FMP and the November 2014 Revision to Amendment 1. The following management strategies are in place for the ASMA and RRMA by these documents:

**Strategies currently in place under the November 2014 Revision to Amendment 1 and Amendment 1 to the North Carolina Estuarine Striped Bass FMP:**

### **A-R stock has been managed with a TAL since 1991**

- Maintain current TAL of 275,000 lb
- The TAL will continue to be split evenly between commercial and recreational sectors
- ASMA commercial TAL = 137,500 lb
- ASMA recreational TAL = 68,750 lb
- RRMA recreational TAL = 68,750 lb

### **ASMA Commercial Harvest (TAL = 137,500 lb)**

- 18-inch total length (TL) minimum size limit (ASMFC compliance requirement)
- Continue to operate as a bycatch fishery
- Spring season, anytime between Jan. 1–April 30
- Fall season, anytime between Oct. 1–Dec. 31
- Daily trip limits for striped bass
- Maintain gill-net mesh size and yardage restrictions
- Maintain seasonal and area closures
- Maintain attendance requirements for small mesh nets (mid-May through late November)

### **ASMA Recreational Harvest (TAL = 68,750 lb)**

- 18-inch TL minimum size limit
- Daily creel limit (can be adjusted as necessary to keep harvest below the TAL)
- Open 7 days a week all season (can be adjusted as necessary to keep harvest below the TAL)
- Spring season, anytime between Jan. 1–April 30
- Fall season, anytime between Oct. 1–Dec. 31

### **RRMA Recreational Harvest (TAL = 68,750 lb)**

- 18-inch TL minimum size limit
- Protective slot (no harvest): 22–27 inches TL
- 2 fish daily creel, only one of which can be greater than 27 inches TL
- Harvest season in entire river opens on March 1 and closes on April 30 by rule since 2008
- Single barbless hook regulation from April 1–June 30 in Inland waters above the US 258 Bridge

### **Management of TALs for ASMA and RRMA**

- BRPs (*F* and SSB) for the A-R stock will be determined through North Carolina A-R striped bass benchmark stock assessments, which must be approved by the ASMFC Atlantic Striped Bass Management Board
- Short-term Overages: if the harvest point estimate exceeds the total TAL by 10% in a single year, overage is deducted from the next year and restrictive measures implemented in the responsible fishery(ies)
- Long-term Overages: five-year running average of harvest point estimate exceeds the five-year running average of the total TAL harvest by 2%, the responsible fishery exceeding the harvest limit will be reduced by the amount of the overage for the next five years.

**Should the target *F* be exceeded, then restrictive measures will be imposed to reduce *F* to the target level**

#### IV. AUTHORITY

North Carolina’s existing fisheries management system is powerful and flexible, with rule-making authority granted to the North Carolina Marine Fisheries Commission (NCMFC) and the NCWRC within their respective jurisdictions. Further, the NCMFC has delegated specified proclamation authority to the NCDMF Director in its rules. The NCWRC has authority to issue limited proclamations and may delegate this authority to the NCWRC Executive Director.

#### **Proclamation Authority for the ASMA, RRMA, and CSMA striped bass stocks:**

The NCMFC can regulate fishing times, areas, fishing gear, seasons, size limits, and quantities of fish harvested and possessed in joint and coastal waters (G.S. 113-182 and 143B-289.52). The NCMFC can delegate the authority to implement its regulations for fisheries as set forth in NCMFC rules “which may be affected by variable conditions” to the Director of the NCDMF who may then issue public notices called “proclamations” (G.S. 113-221.1 and 143B-289.52). The NCWRC has authority to license and regulate all fishing activities in inland waters, and the NCWRC also has proclamation authority, which may be delegated to the Executive Director, to suspend or extend seasons for taking of striped bass in inland and joint waters of coastal rivers and their tributaries (G.S. 113-292). Thus, all necessary authority needed for management of the striped bass fisheries is available through the existing state fishery management process.

It should also be noted that under the provisions of the North Carolina Estuarine Striped Bass FMP Amendment 1 the NCDMF Director maintains proclamation authority to establish seasons, authorize or restrict fishing methods and gear, limit quantities taken or possessed, and restrict fishing areas as deemed necessary to maintain a sustainable harvest. The NCWRC Executive Director maintains proclamation authority to establish seasons.

#### N.C. General Statutes

G.S. 113-134.	RULES
G.S. 113-182.	REGULATION OF FISHING AND FISHERIES
G.S. 113-182.1.	FISHERY MANAGEMENT PLANS
G.S. 113-221.1.	PROCLAMATIONS; EMERGENCY REVIEW
G.S. 113-292.	AUTHORITY OF THE WILDLIFE RESOURCES COMMISSION IN REGULATION OF INLAND FISHING AND THE INTRODUCTION OF EXOTIC SPECIES.
G.S. 143B-289.52.	MARINE FISHERIES COMMISSION—POWERS AND DUTIES

#### N.C. Marine Fisheries Commission Rules 2020 and N.C. Wildlife Resources Commission Rules 2020 (15A NCAC)

15A NCAC 03M .0201	GENERAL
15A NCAC 03M .0202	SEASON, SIZE AND HARVEST LIMIT: INTERNAL COASTAL WATERS
15A NCAC 03M .0512	COMPLIANCE WITH FISHERY MANAGEMENT PLANS
15A NCAC 03Q .0107	SPECIAL REGULATIONS: JOINT WATERS

15A NCAC 03Q .0108	MANAGEMENT RESPONSIBILITY FOR ESTUARINE STRIPED BASS IN JOINT WATERS
15A NCAC 03Q .0109	IMPLEMENTATION OF ESTUARINE STRIPED BASS MANAGEMENT PLANS: RECREATIONAL FISHING
15A NCAC 03R .0201	STRIPED BASS MANAGEMENT AREAS
15A NCAC 10C .0110	MANAGEMENT RESPONSIBILITY FOR ESTUARINE STRIPED BASS IN JOINT WATERS
15A NCAC 10C .0111	IMPLEMENTATION OF ESTUARINE STRIPED BASS MANAGEMENT PLANS: RECREATIONAL FISHING
15A NCAC 10C .0301	INLAND GAME FISHES DESIGNATED
15A NCAC 10C .0314	STRIPED BASS

## V. DISCUSSION

Results from the 2020 A-R striped bass benchmark stock assessment indicate the stock is overfished and overfishing is occurring (Lee et. al 2020). The estimate of  $F$  in the terminal year of the assessment (2017) was 0.27, above the  $F_{35\%SPR}$  Threshold of 0.18 (Figure 1) and the estimate of SSB was 78,576 lb, below the  $SSB_{35\%SPR}$  Threshold of 267,390 lb (Figure 2). Estimates of  $F$  have been above the  $F_{35\%SPR}$  Threshold in 24 out of the 27 years of the time period of the assessment (Figure 1). Female SSB has declined steadily from a high of 587,516 lb in 2000 to a low of 45,418 lb in 2013. Female SSB increased through 2015 to 167,053 lb and has declined since (Figure 2). Results of the assessment also show a period of strong recruitment (as measured by the number of age-0 fish coming into the stock each year) from 1993 to 2000, then a period of much lower recruitment from 2001 to 2017, which has contributed to the decline in SSB since 2003. Average recruitment from 1993-2000 was 1,127,646 age-0 fish per year while average recruitment for years 2001-2017 was 428,796 age-0 fish per year (Figure 2).

Several years of poor recruitment occurred from 2001–2004 at a time when SSB was at high levels, indicating factors other than abundance of SSB may be contributing to poor spawning success in some years. Appropriate river flow during the spawning period has long been recognized as an important factor in spawning success for A-R striped bass (Hassler et. al 1981; Rulifson and Manooch 1990). Low to moderate flows have been identified as favorable to strong year-class production while high flows (10,000 cubic feet per second or greater) are unfavorable to the formation of strong year classes. The peer reviewers of the 2020 benchmark assessment recognized the importance of river flow on recruitment and noted declining recruitment in the time series does not appear to result solely from reduced abundance due to harvest (Lee et. al 2020).

Concerning trends are also evident in all the juvenile and adult fishery-independent surveys of relative abundance conducted by the NCDMF and NCWRC to monitor the A-R striped bass stock. Both NCDMF gill-net surveys and the NCWRC electrofishing survey show declining trends, especially in the number of older fish, in recent years below levels of abundance observed when the stock was severely depressed in the early 1990s. Harvest from all sectors since about 2005 have shown similar declining trends as total abundance estimates from the stock assessment, which indicate a declining trend in total abundance since the early 2000s (Figures 1 and 3).

Since the TAL increase to 550,000 lb in 2003 (Table 2, Figure 3), total combined landings from all fisheries in the ASMA and RRMA have not exceeded 460,853 lb and have averaged 235,278 lb per year with a low of 108,432 lb in 2013 (Figure 3). For the years 2005–2013, the commercial sector did not reach their TAL. Estimates of total abundance from the stock assessment (Figure 1), suggest the reason for the decline in harvest was likely a decline in overall stock abundance due to poor recruitment (Figure 2). Even since the 2014 reduction in the TAL to 275,000 lb the commercial and recreational sectors in the ASMA did not reach the TAL from 2014–2017. Harvest in all sectors has increased since 2017, with the commercial sector reaching the TAL in 2019 causing the NCDMF to close the fall commercial harvest season before Dec. 31 for the first time since 2010. This increase in harvest is likely due to the above-average year classes produced in 2014 and 2015 (Figure 2). The fisheries are primarily composed of fish age 3–6 so the indication of good recruitment in the fishery as seen in landings is offset by 2–4 years as the new recruits grow and begin to enter the fisheries.

Since the early 2000s the recreational sectors have only approached their TAL in 2015 and 2016 (Figure 3). Harvest in the recreational sectors consists primarily of fish age 3–5. Even with an increase in the daily creel limit in the ASMA from two fish per person per day to three fish per person per day in the fall of 2006 through the fall of 2015, harvest was still below the TAL in all years except 2015. The daily creel limit was reduced back to two fish per person per day in the spring of 2016.

Recreational harvest in the RRMA is more controlled by the daily creel limit than in the ASMA. The Roanoke River is a smaller body of water and striped bass congregate in large numbers throughout the river on their way to and while on the spawning grounds. Because the fish are moving through the system for spawning activity in a more compressed area, recreational anglers tend to release more legal sized fish than anglers in the ASMA. An increase in the daily creel limit in the RRMA to more than two fish per person per day would likely result in the TAL being exceeded in most years in the RRMA.

### **Reductions in the TAL to lower $F$ to the target reference point value**

Adaptive management in Amendment 1 to the North Carolina Estuarine Striped Bass FMP states “should the target  $F$  be exceeded, then restrictive measures will be imposed to reduce  $F$  to the target level”. Amendment 1 does not specify a time frame to bring  $F$  back to the target. Total removals in 2017 included 119,244 lb of harvest and 23,795 lb of dead discards. Assuming the same level of discards, landings will need to be reduced by 57% compared to 2017 landings to lower  $F$  to the target of  $F_{45\%SPR}$  of 0.13. This 57% reduction from 2017 landings equates to a new overall TAL of 51,216 lb. for the ASMA and RRMA. As with all fisheries, the A-R stock recovery under the new TAL is subject to other factors. Future spawning success and subsequent recruitment levels are the main area of uncertainty. If the stock experiences even a few good years of recruitment, stock abundance can increase quickly under low levels of harvest. Given the new TAL reflects the target  $F$  reference point and not the threshold  $F$ , it does provide some amount of buffer for changing circumstances and provides a constant level of constrained harvest while Amendment 2 is developed to address long-term management needs.



There are several management measures available through proclamations or rules that allow the NCDMF and NCWRC to keep harvest levels below the proposed TAL in the ASMA and RRMA. For the commercial fishery these include daily reporting of landings by striped bass dealers for daily monitoring of harvest, mandatory tagging of all striped bass sold, adjusting the daily possession limit, adjusting the opening and closing of the season, area closures, and gill-net yardage restrictions. For the ASMA and RRMA recreational fisheries, measures include a creel survey that allows for weekly estimates of harvest, adjusting the daily possession limit, adjusting the allowable harvest days during the open season, adjusting the opening and closing of the season, and area closures.

Starting in January 2021 the above-mentioned management measures will be used to keep harvest below the newly reduced TAL.

The NCDMF and NCWRC members of the FMP Plan Development Team met several times to discuss the issues outlined in this document, and based on those discussions, agreed to set the new TAL for the A-R striped bass stock at 51,216 lb. The following section serves to revise Amendment 1 to the North Carolina Estuarine Striped Bass FMP to reflect the new TAL that will lower  $F$  to the target level.

## **VI. TOTAL ALLOWABLE LANDINGS MANAGEMENT REVISION TO AMENDMENT 1 TO THE NORTH CAROLINA ESTUARINE STRIPED BASS FMP**

Amendment 1 to the North Carolina Estuarine Striped Bass FMP, in conjunction with the North Carolina FMP for Interjurisdictional Fisheries, provides the framework for the changes in management proposed herein. This document will be incorporated as the November 2020 Revision to Amendment 1 to the North Carolina Estuarine Striped Bass FMP, and replaces the November 2014 Revision to Amendment 1 to the North Carolina Estuarine Striped Bass FMP. It will serve to document the rationale agreed to by the NCDMF and NCWRC for the following management strategy to begin Jan. 1, 2021 and continue until the adoption of Amendment 2.

- Biological Reference Points ( $F$  and SSB) for the A-R stock will be determined through North Carolina A-R striped bass benchmark stock assessments and updates
- Benchmark assessments will be reviewed by the ASMFC Striped Bass Management Board for approval
- Set the TAL for the A-R stock at 51,216 lb, to be split evenly between the commercial and recreational sectors as follows:
  - ASMA commercial TAL = 25,608 lb
  - ASMA recreational TAL = 12,804 lb
  - RRMA recreational TAL = 12,804 lb

All other management strategies contained in Amendment 1 will remain in force until another North Carolina Estuarine Striped Bass FMP revision is implemented or amendment is adopted.

Table 1. Biological reference points for the Albemarle-Roanoke striped bass stock and the point estimate from the terminal year (2017) of the assessment. Source: Lee et al. 2020

<b>Biological Reference Points</b>		<b>Terminal Year (2017) Estimate</b>
$F_{45\%SPR}$ Target	0.13	$F = 0.27$
$F_{35\%SPR}$ Threshold	0.18	
SSB $_{45\%SPR}$ Target	350,371 lb	SSB = 78,576 lb
SSB $_{35\%SPR}$ Threshold	267,390 lb	

Table 2. Total allowable landings (lb) for the Albemarle-Roanoke striped bass stock, 1991–2019.

<b>Years</b>	<b>Total Allowable Landings</b>	<b>ASMA Commercial</b>	<b>ASMA Recreational</b>	<b>RRMA Recreational</b>
1991–1997	156,800	98,000	29,400	29,400
1998	250,800	125,400	62,700	62,700
1999	275,880	137,940	68,970	68,970
2000–2002	450,000	225,000	112,500	112,500
2003–2014	550,000	275,000	137,500	137,500
2015–2019	275,000	137,500	68,750	68,750

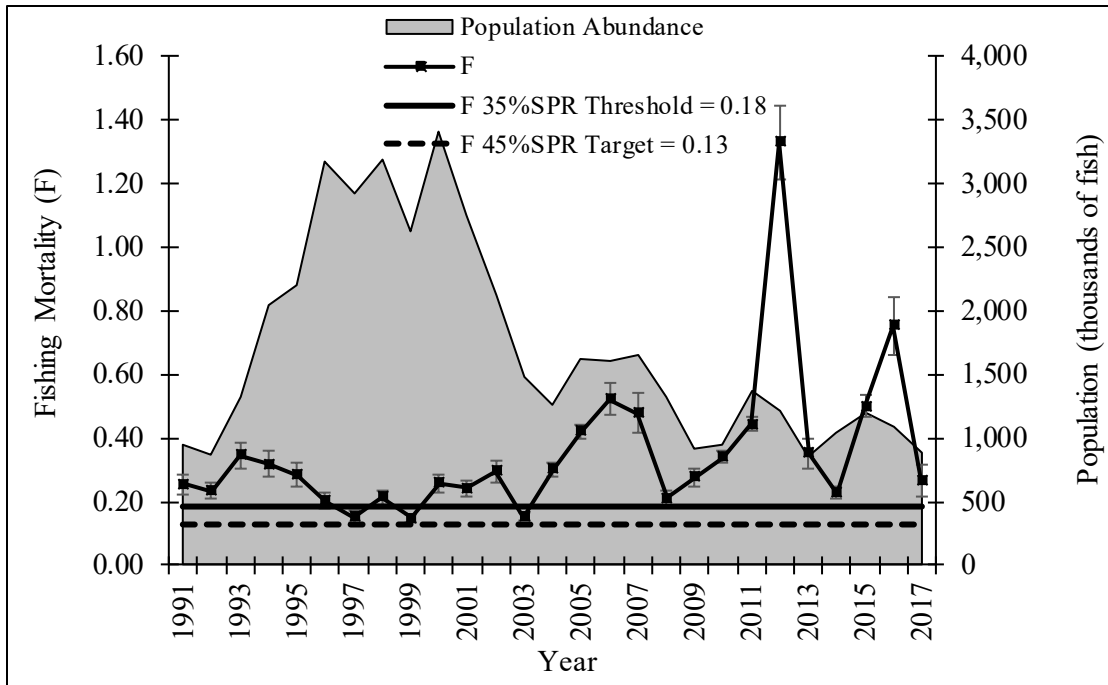


Figure 1. Estimates of fishing mortality ( $F$ ) and population abundance for the Albemarle-Roanoke striped bass stock, 1991–2017. Source: Lee et al. 2020

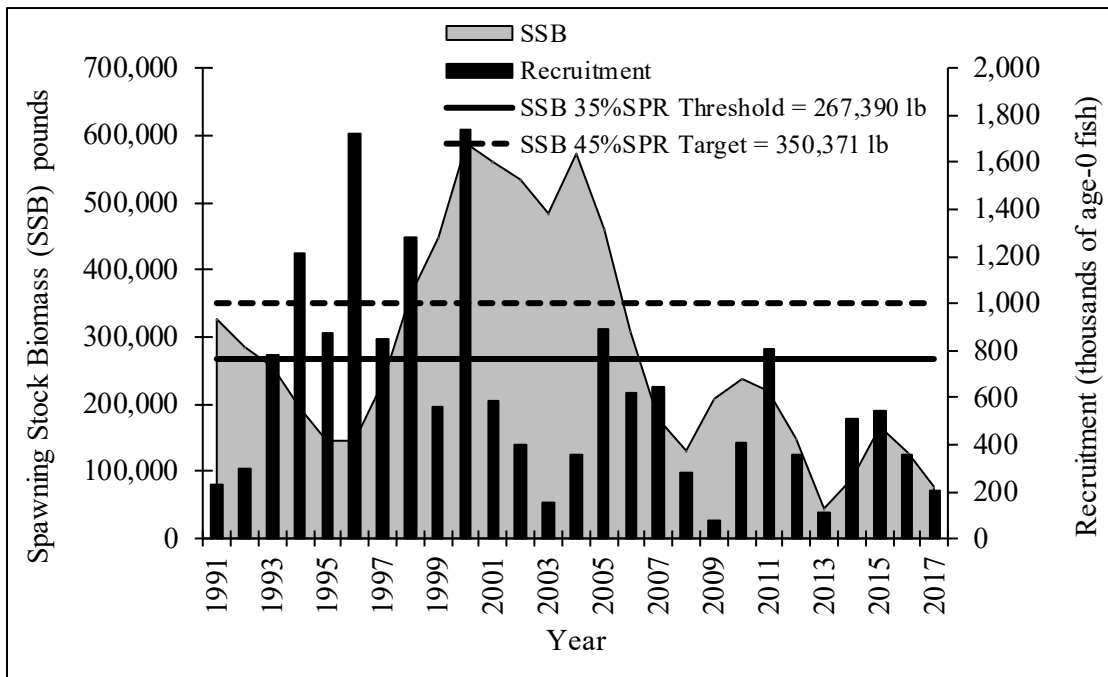


Figure 2. Estimates of spawning stock biomass (SSB) and recruitment of age-0 fish coming into the population each year for the Albemarle-Roanoke striped bass stock, 1991–2017. Source: Lee et al. 2020

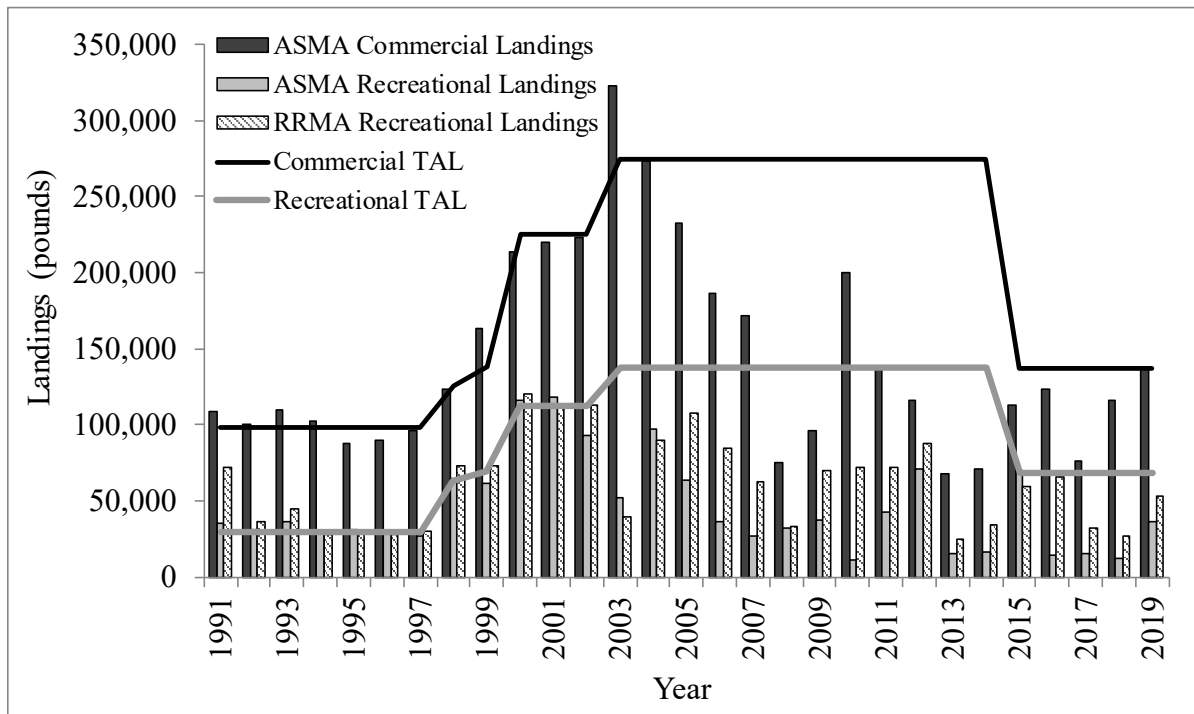


Figure 3. Striped bass landings from the Albemarle Sound Management Area commercial and recreational sectors and Roanoke River Management Area recreational sector and the total allowable landings, 1991–2019.

## VII. REFERENCES CITED

- Atlantic States Marine Fisheries Commission (ASMFC). 2003. Amendment # 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 41, Washington, DC. 63 p.
- ASMFC. 2014. Addendum IV to Amendment # 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Washington, DC. 20 p.
- Berggren, T.J., and J.T. Lieberman. 1978. Relative contribution of Hudson, Chesapeake and Roanoke striped bass, *Morone saxatilis*, stocks to the Atlantic coast fishery. Fishery Bulletin 76(2): 335–345.
- Callihan, J.L., C.H. Godwin, and J.A. Buckel. 2014. Effects of demography on spatial distribution: Movement patterns of Albemarle-Roanoke striped bass *Morone saxatilis* in relation to their stock recovery. Fishery Bulletin 112:131–143.
- NCDMF. 2013. Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environmental and Natural Resources, Division of Marine Fisheries. Morehead City, NC. 826 p.
- Lee, L.M., T.D. Tears, Y. Li, S. Darsee, and C. Godwin (editors). 2020. Assessment of the Albemarle Sound-Roanoke River striped bass (*Morone saxatilis*) in North Carolina, 1991–2017. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2020-01, Morehead City, North Carolina. 171 p.

# Scoping Document

Photo By: Buzz Bryson  
Striped bass spawning in the  
Roanoke River, Weldon, NC



## Management Strategies for Amendment 2 to the North Carolina Estuarine Striped Bass Fishery Management Plan



October 2020



Photo By: Jesse Bissette

## The N.C. Division of Marine Fisheries seeks your input on management strategies for the Estuarine Striped Bass Fishery Management Plan.

A scoping period for public comment begins  
Nov. 2, 2020 and ends Nov. 15, 2020.

Comments must be received by  
5 p.m. (EST) on Nov. 15, 2020.

### Scoping Meetings

DMF staff will provide information about Amendment 2 to the N.C. Estuarine Striped Bass FMP. A public comment period will follow.

The public may participate in the meeting online or by telephone. To facilitate comments, the division is asking those who wish to speak during the meeting to pre-register.

Links to scoping information, including registration to speak, webinar instructions, the call-in telephone number, and other references, can be found through the N.C. Estuarine Striped Bass Amendment 2 Information Page (<http://portal.ncdenr.org/web/mf/striped-bass-amendment-topic>).

#### **Thursday, Nov. 5, 2020: 6 p.m. to 8 p.m.**

[https://ncdenrits.webex.com/ncdenrits/onstage/g.php?  
MTID=e4fc435aebfcdedafed56b82e7def8173](https://ncdenrits.webex.com/ncdenrits/onstage/g.php?MTID=e4fc435aebfcdedafed56b82e7def8173)

Event number 171 493 2224

Event password 1234

Join by audio only +1-415-655-0003 US TOLL

#### **Monday, Nov. 9, 2020: 6 p.m. to 8 p.m.**

[https://ncdenrits.webex.com/ncdenrits/onstage/g.php?  
MTID=ebedeb5306d80ed62d46c9b0db81f9783](https://ncdenrits.webex.com/ncdenrits/onstage/g.php?MTID=ebedeb5306d80ed62d46c9b0db81f9783)

Event number 171 937 9432

Event password 1234

Join by audio only +1-415-655-0003 US TOLL

### Can't attend but want to submit comments? Here's how!

Written comments can be submitted by online form or by U.S. mail. Comments sent by U.S. mail must be received by Nov. 15, 2020 to be accepted. The division will not accept public comment through email.

#### **To comment by online form:**

The online form can be accessed through the N.C. Estuarine Striped Bass Amendment 2 Information Page (<http://portal.ncdenr.org/web/mf/striped-bass-amendment-topic>). Please use the link at the bottom of the information page.

#### **To comment by U.S. mail, please submit written comments to:**

N.C. Division of Marine Fisheries  
N.C. Estuarine Striped Bass  
FMP Amendment 2  
Scoping Comments  
P.O. Box 769  
Morehead City, NC 28557

# Questions about the estuarine striped bass stocks, fisheries, or Amendment 2 to the North Carolina Estuarine Striped Bass Fishery Management Plan?

Pictured: Brent Griffin



## Contact the leads:

### Charlton Godwin

Fisheries Biologist DMF, Elizabeth City

252-264-3911

Co-lead

### Todd Mathes

Fisheries Biologist DMF, Washington

252-948-3872

Co-lead

### Jeremy McCargo

Fisheries Biologist WRC, Raleigh

919-707-4081

## Questions about the FMP Process?

### Kathy Rawls

Fisheries Management Section Chief, Morehead City

252-808-8074

### Corrin Flora

Fisheries Management Plan Coordinator, Morehead City

252-726-7021



## Purpose of the Scoping Document

The purpose of this document is to inform the public the review of the N.C. Estuarine Striped Bass Fishery Management Plan (FMP) is underway and to provide an opportunity for the public to comment on identified management strategies or identify other relevant strategies in the management of the estuarine striped bass fishery. Striped bass in North Carolina are jointly-managed by the N.C. Marine Fisheries Commission (MFC) and N.C. Wildlife Resources Commission (WRC). Input received at the start of the FMP review process may shape the final amendment and its management measures (solutions). To help focus the input received from the public, this document provides an overview of initially identified strategies, as well as background information on the fisheries and the stocks. A series of questions about each strategy is also provided for the public to consider when thinking about the strategies; in general: What should estuarine striped bass management be? Are changes needed and, if so, what changes are needed?

Additional management strategies may be considered in Amendment 2 dependent on statutory requirements, available data, research needs, and the degree of impact the management strategy would have and how effective the solution would be. If the division determines a management strategy raised during the scoping period might have positive impacts on the stocks, additional examination of the strategy may be undertaken in the development of the FMP.

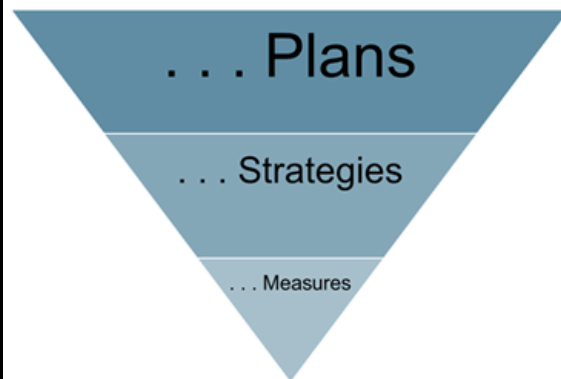
Scoping provides an opportunity for the public to comment on strategies identified by the division as well as any additional relevant strategies for possible consideration for the development of the FMP.

## What is Scoping?

Scoping is the first stage of the process to determine the appropriate contents of an FMP. Scoping serves many purposes including: (1) to provide notice to the public that a formal review of the FMP is underway by the N.C. Division of Marine Fisheries (DMF or division), (2) inform the public of the stock status of the species (3) solicit stakeholder input on a list of strategies identified by the DMF and identify other relevant strategies that may need to be addressed, and (4) recruit potential advisors to serve on the advisory committee (AC) for the FMP that is appointed by the MFC. The public will have more opportunity to provide comments as the amendment is developed; however, scoping is the first and best opportunity to provide input on potential strategies for DMF to consider before an amendment is developed.

## FISHERY MANAGEMENT PLANS - A TIERED APPROACH

### Fishery Management



Management PLANS are implemented to achieve specified management goals for a fishery, such as sustainable harvest, and include background information, data analyses, fishery habitat and water quality considerations consistent with Coastal Habitat Protection Plans, research recommendations, and management strategies.

Management STRATEGIES are adopted to help reach the goal and objectives of the plan. They are the sum of all the management measures selected to achieve the biological, ecological, economic, and social objectives of the fishery.

Management MEASURES are the actions implemented to help control the fishery as stipulated in the management strategies.

## Developing an amendment

Annually, the DMF reviews all species for which there are FMPs for North Carolina and provides an update to the MFC. This review includes any recommended changes to the schedule for FMP review and amendment development. Per N.C. law, any changes to the schedule must be approved by the N.C. Department of Environmental Quality (N.C. DEQ) Secretary.

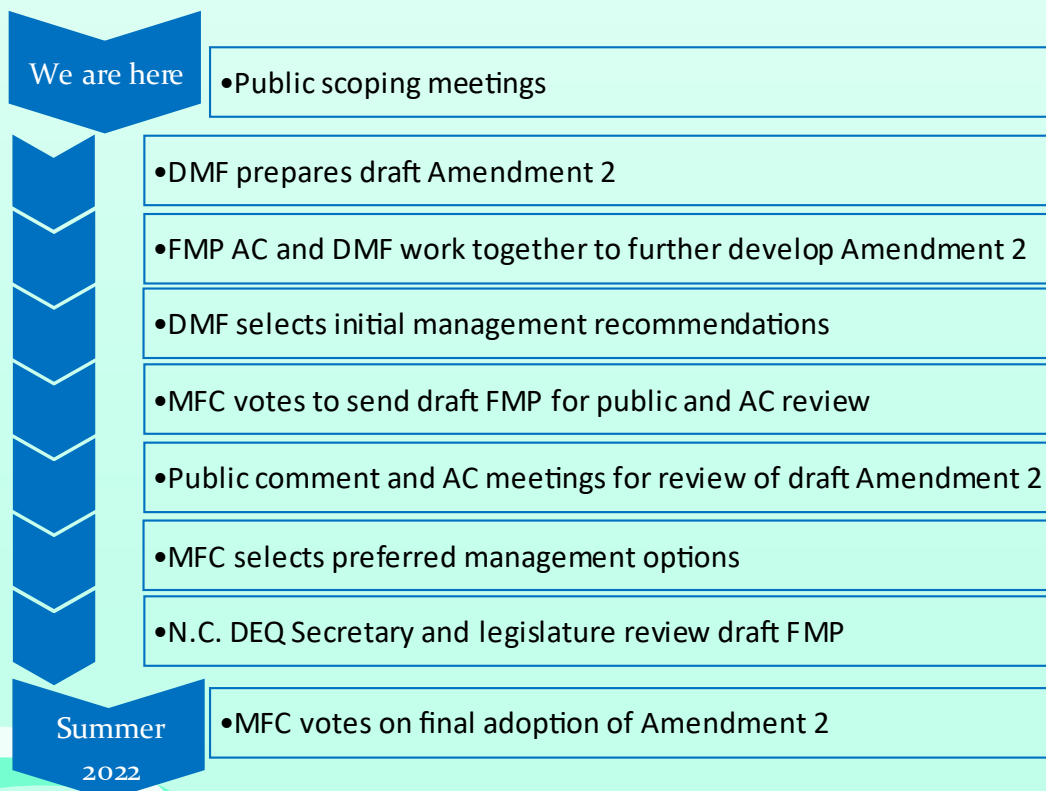
When a plan is opened for review, the first step of the formal amendment process begins with a stock assessment of the species when applicable, followed by the scoping period. After relevant strategies have been identified by the DMF, the public (during the scoping period), and by the MFC, the division's plan development team (PDT) develops a preliminary draft amendment. The first draft will be completed before the FMP AC is appointed. Once appointed, the AC will meet with the PDT at a series of workshops to assist in developing the FMP by further refining the draft amendment. Upon completion of this draft, the amendment is taken to the MFC for approval to go out for public comment and review by the MFC's standing and regional ACs. Following consideration of public and AC comment, the MFC selects its preferred management measures for Amendment 2. Next, draft Amendment 2 goes to the N.C. DEQ Secretary and the legislature for review before the MFC votes on final approval of the amendment.

In the case of a jointly managed species such as striped bass, the WRC consults throughout the FMP amendment process. WRC staff participate in the development of the stock assessment and serve on the PDT. Concurrent with MFC actions, the WRC board reviews the draft FMP, selects preferred management measures, considers its support of the final FMP recommendations, and initiates rulemaking as required.



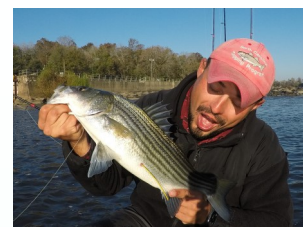
WRC electrofishing spawning stock survey index of abundance  
Roanoke River, Weldon, NC.

## FMP Timeline



## Why is this happening now?

The 2020 N.C. FMP Review Schedule shows the review of the N.C. Estuarine Striped Bass FMP is underway. To begin the development of Amendment 2 to the N.C. Estuarine Striped Bass FMP, the division conducted assessments of the Albemarle-Roanoke striped bass stock, and the striped bass stocks in the Tar-Pamlico, Neuse, and Cape Fear rivers.



Pictured: Adam B.  
Cape Fear River, N.C.

## Amendment 2 Background

There are two geographic management units and four striped bass stocks included in the North Carolina Estuarine Striped Bass FMP. The northern management unit is comprised of two harvest management areas: the Albemarle Sound Management Area (ASMA) and the Roanoke River Management Area (RRMA). The striped bass stock in these two harvest management areas is referred to as the Albemarle-Roanoke (A-R) stock, and its spawning grounds are in the Roanoke River in the vicinity of Weldon, NC. The southern geographic management unit is the Central Southern Management Area (CSMA) and includes all internal coastal, joint and contiguous inland waters of North Carolina south of the ASMA to the South Carolina state line. There are spawning stocks in each of the major river systems within the CSMA; the Tar-Pamlico, the Neuse, and the Cape Fear. Only the A-R stock is included in the management unit of Amendment 6 to the Atlantic States Marine Fisheries Commission's (ASMFC) Interstate FMP for Atlantic Striped Bass (ASMFC 2003).

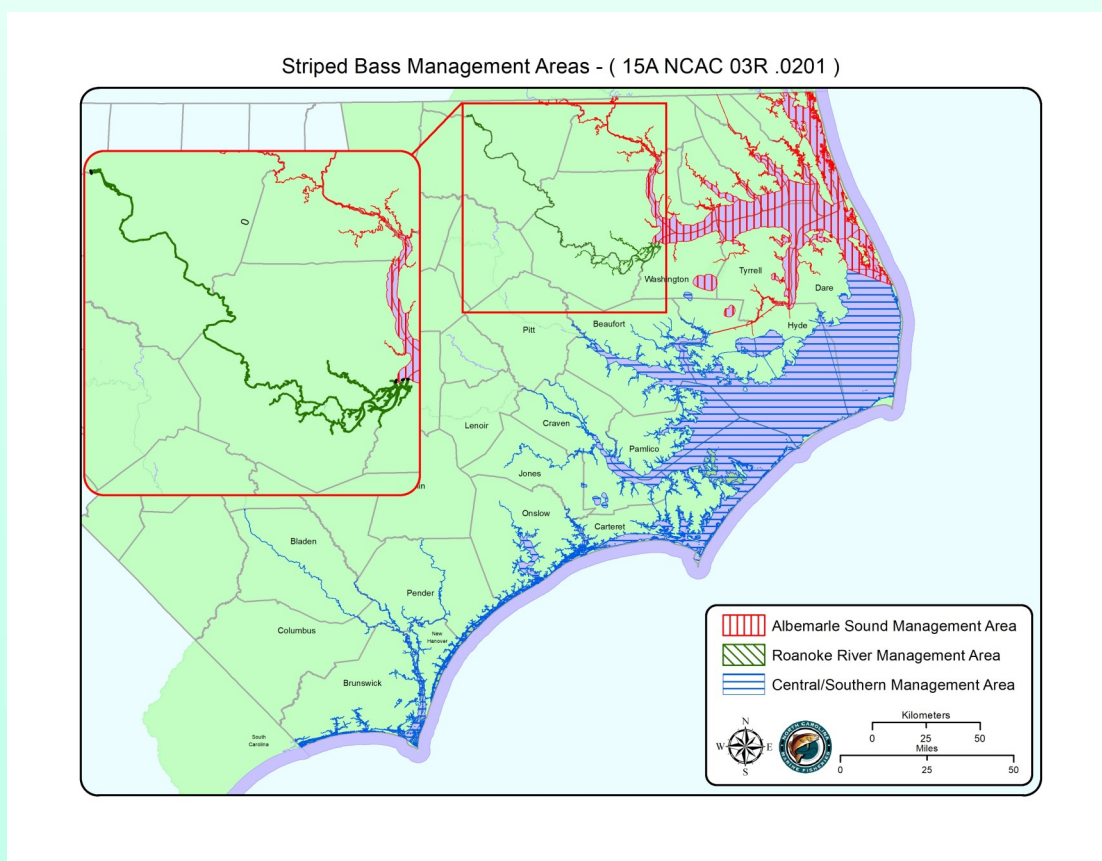


Figure 1. North Carolina's estuarine striped bass management areas.

# Albemarle-Roanoke striped bass stock assessment and stock status

Results from the 2020 benchmark stock assessment indicate the A-R striped bass stock is overfished and overfishing is occurring in the terminal year of the assessment (2017) relative to the updated biological reference points (BRPs). These BRPs are based on spawning stock biomass (SSB) targets and thresholds of  $SSB_{45\%SPR}$  Target = 350,371 lb and  $SSB_{35\%SPR}$  Threshold = 267,390 lb respectively, and fishing mortality (F) targets and thresholds of  $F_{45\%SPR}$  Target = 0.13 and  $F_{35\%SPR}$  Threshold = 0.18 (Figures 2 and 3; Lee et al. 2020).

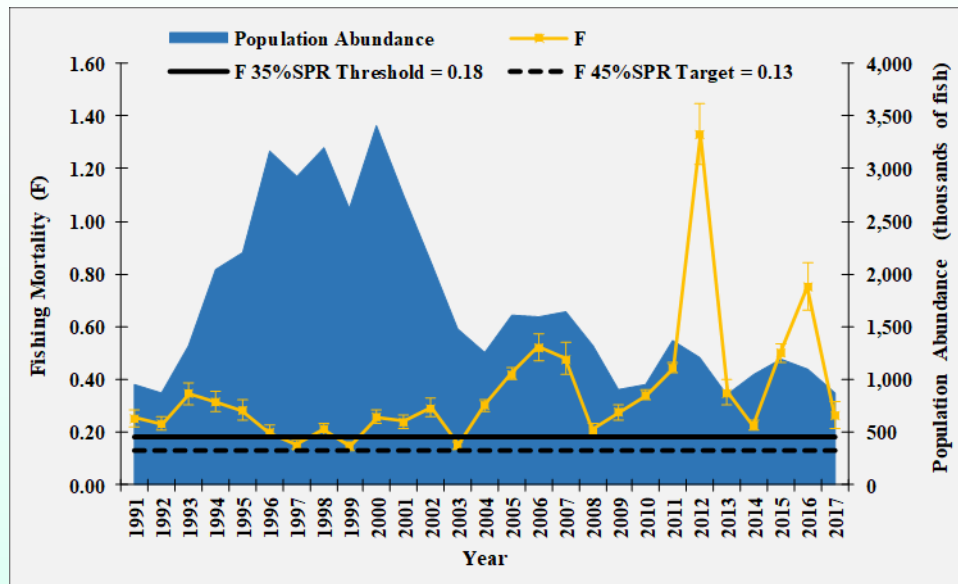


Figure 2. Estimates of fishing mortality (F) and population abundance for the Albemarle-Roanoke striped bass stock, 1991–2017. Source: Lee et al. 2020.

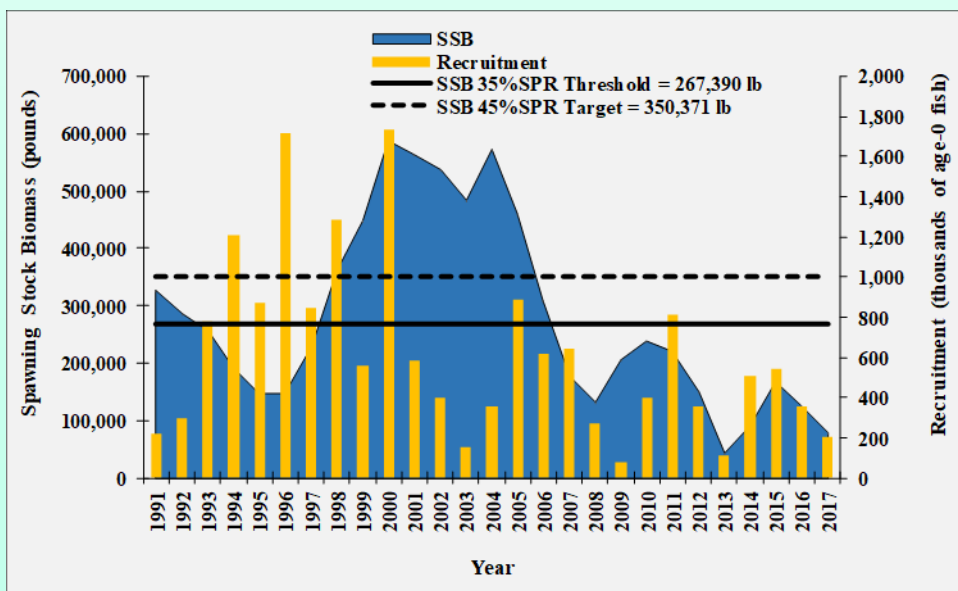


Figure 3. Estimates of spawning stock biomass (SSB) and recruitment of age-0 fish coming into the population each year for the Albemarle-Roanoke striped bass stock, 1991–2017. Source: Lee et al. 2020.

# Albemarle-Roanoke Striped Bass in North Carolina

A-R striped bass have long supported recreational and commercial fisheries in the Albemarle Sound region and its tributaries and the northern Outer Banks. Commercial harvest of striped bass occurs throughout the fall and winter into the early spring. Since 1991 gill-nets are the main commercial harvest gear with minimal harvest also from pound nets. Recreational striped bass fishing occurs throughout the year, with harvest seasons allowed in the fall and winter and through the spring as striped bass migrate to the spawning grounds. During the late spring and summer, catch-and-release fishing is also popular.



Recreational anglers, Albemarle Sound bridge. Photo credit: DMF staff  
Pictured: K.D. and Kenny Hewitt

Harvest has been controlled by a fixed annual poundage amount known as total allowable landings (TAL) since 1991. The TAL is split evenly between commercial and recreational sectors, and the recreational TAL is further divided evenly between the ASMA and RRMA (Figure 4). Since the last TAL increase to 550,000 lb in 2003, combined landings from all fisheries in the ASMA and RRMA have not exceeded 460,853 lb and have averaged 235,278 lb per year with a low of 108,432 lb in 2013. The commercial sector did not reach their TAL in any years from 2005 to 2013. Even with the 2014 reduction in the TAL to 275,000 lb the commercial and recreational sectors in the ASMA did not reach the TAL for years 2014–2017. Harvest in all sectors has increased since 2017, with the commercial sector reaching the TAL in 2019 causing the DMF to close the fall commercial harvest season before December 31 for the first time since 2010. This increase in harvest is likely due to the above-average year classes produced in 2014 and 2015 (Figures 3 and 4).

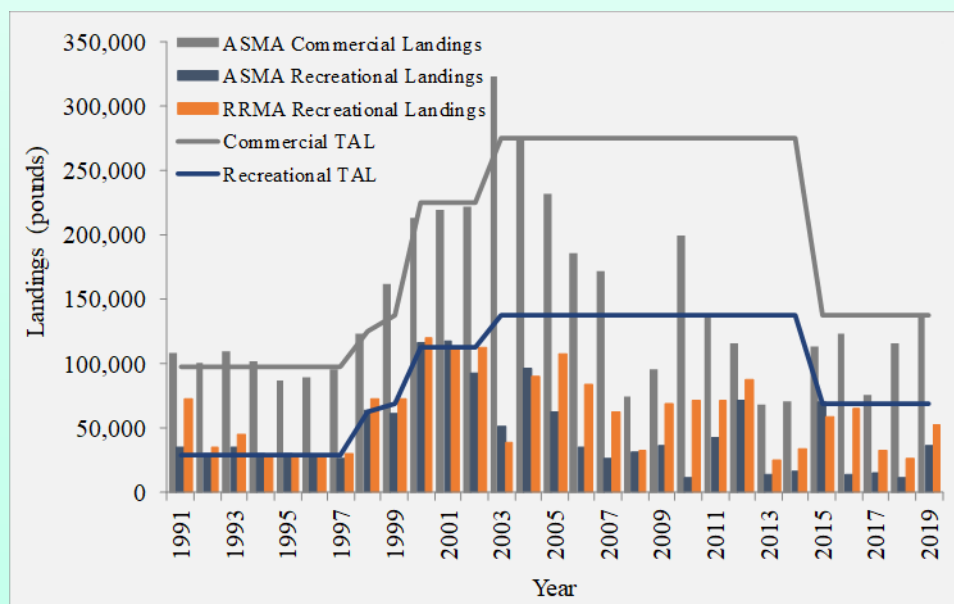


Figure 4. Striped bass landings from the Albemarle Sound Management Area commercial and recreational sectors and Roanoke River Management Area recreational sector, and the commercial and recreational total allowable landings, 1991–2019.



Pictured: Kaden

Based on results from the estimates of total abundance from the stock assessment (Figure 2), the reason for the decline in harvest is likely a decline in overall stock abundance due to poor recruitment starting in 2001 (Figure 3). The assessment noted the importance of river flow on recruitment and noted declining recruitment in the time series does not appear to result solely from reduced abundance due to amount harvested, as recruitment started declining when SSB was at high levels (Figure 3; Lee et. al 2020).

Average total removals in the fisheries (sector combined) during 2012–2017 were composed of 84% landings, with dead discards equaling 16% in numbers of fish (Figure 5). Discards in the ASMA commercial fishery from 2012 to 2017 were estimated using a generalized linear model framework based on on-board observer data combined with data from the DMF Trip Ticket Program. Discards in the recreational fishery are estimated by multiplying the number of fish released by a delayed mortality estimate of 6.4% (Nelson 1998).

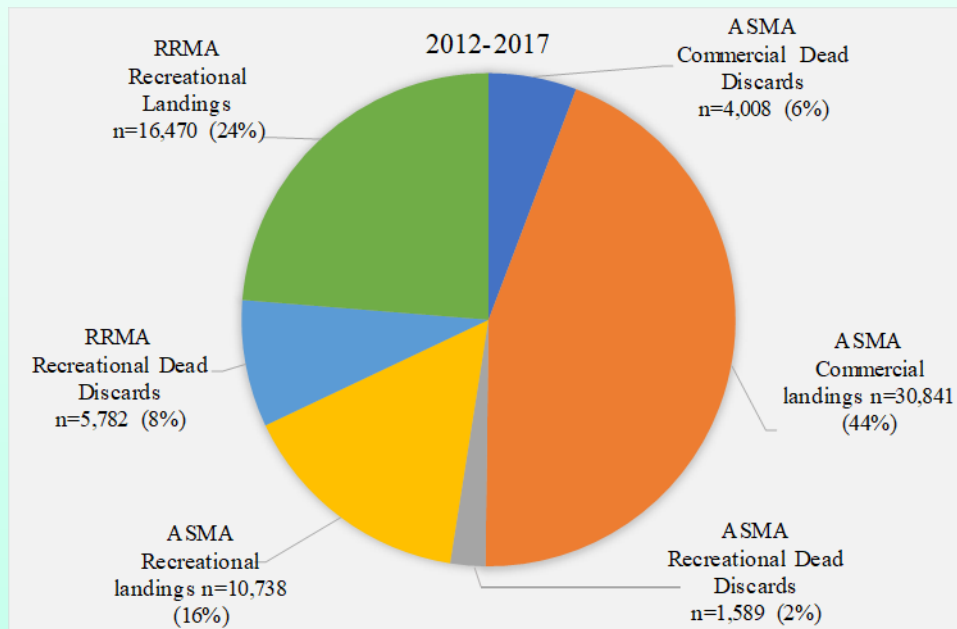


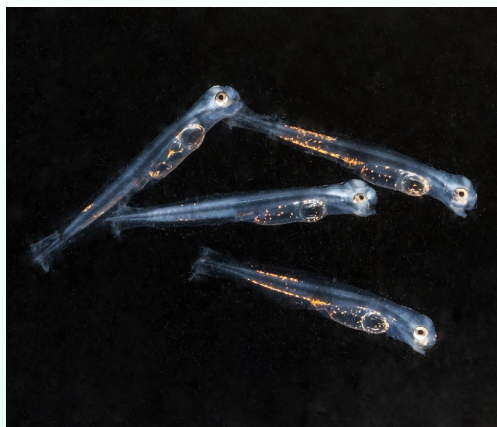
Figure 5. Average number of striped bass landed and discarded from the commercial and recreational fisheries in the Albemarle Sound Management Area (ASMA) and Roanoke River Management Area (RRMA), 2012-2017. Source: Lee et al. 2020.

## Tar-Pamlico, Neuse, and Cape Fear river striped bass stocks review

There is no stock status determination for the CSMA striped bass stocks, comprised of the Tar-Pamlico, Neuse, and Cape Fear rivers. Continuous stocking efforts since 1980 and lack of natural recruitment in these waters prevent the use of traditional stock assessment techniques. The Central Southern Management Area Stock Report (Mathes et al. 2020) is a documentation of all data collected, management efforts, and major analyses completed for these river stocks.



Juvenile striped bass tagged for stocking into the Tar-Pamlico River  
Photo By: Corrin Flora



Striped Bass Larvae  
Photo By: Robert Michelson,  
Coastal Review Online

The report also serves as a record of completed research efforts with implications for fishery management and as a guide for future research based on results and identified data gaps. It evaluates the likelihood of successful population rebuilding under various simulations of stocking and fishery management strategies such as different harvest levels and size limits. Tagging studies in the Cape Fear River showed a consistent decline in striped bass abundance estimates from 2012 to 2018 despite a no-possession regulation since 2008. The need for continued conservation to achieve a sustainable harvest is supported by the lack of recruitment, constrained size and age distributions, low abundance, the absence of older fish in all stocks, and the high percentage of stocked fish in the population (Cushman et al. 2018; Farrae and Darden 2018).

## Tar-Pamlico, Neuse, and Cape Fear river striped bass in North Carolina

Striped bass have long supported recreational and commercial fisheries in the CSMA region and its tributaries. Since 2004 commercial landings in the CSMA have only been allowed in the spring of the year and have been constrained by an annual TAL of 25,000 pounds established in 1994. Over the past 10 years, landings have closely followed the annual TAL due to daily quota monitoring that allows the season to be closed each year when the TAL is reached, except for 2008 when less than half of the TAL was landed and the season stayed open through April 30. Since 2004 striped bass commercial landings in the CSMA have averaged 24,179 pounds and ranged from a low of 10,115 pounds in 2008 to a high of 32,479 pounds in 2004 (Figure 6).

Within the CSMA recreational harvest occurs in the fall and spring and there is a significant recreational catch-and-release fishery throughout the year. Since 2004 striped bass recreational landings have averaged 13,511 pounds but in 2016 and 2017 recreational harvest increased to just over 25,000 lb each year (Figure 6).

From 2012 to 2017 total removals in the commercial and recreational fisheries were composed of 73% landings and 27% dead discards (Figure 7). Discards in the CSMA commercial fishery from 2012 to 2017 were estimated using a generalized linear model framework using on-board observer data combined with data from the DMF trip ticket program. Discards in the recreational fishery are estimated by multiplying the number of fish released by a delayed mortality estimate of 6.4% (Nelson 1998).

There has been a commercial and recreational no-possession provision in the Cape Fear River since 2008. At the MFC's February 2019 business meeting, Supplement A to Amendment 1 to the North Carolina Estuarine Striped Bass FMP was approved instituting a recreational and commercial no-possession provision in the CSMA. On March 13, 2019, the MFC held an emergency meeting at which time they passed a motion requiring the Director to issue a proclamation prohibiting the use of all gill-nets upstream of the ferry lines from the Bayview Ferry to Aurora Ferry on the Pamlico River and the Minnesott Beach Ferry to Cherry Branch Ferry on the Neuse River.



Pictured: DMF Staff. Roanoke River, Weldon, NC

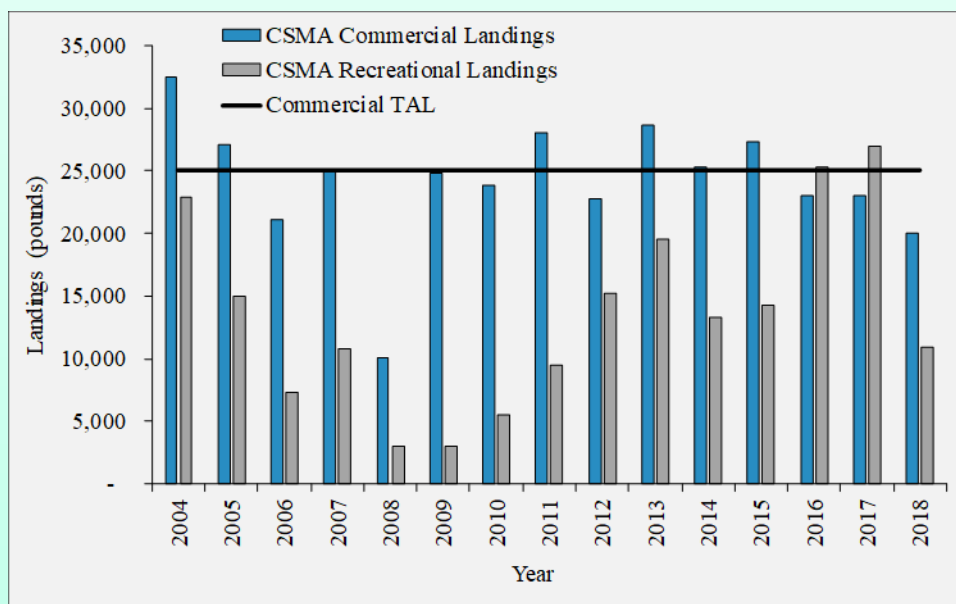


Figure 6. Striped bass landings from the Central Southern Management Area (CSMA) commercial and recreational sectors and the commercial total allowable landings (TAL), 2004–2018. Commercial landings were included for the Cape Fear River for 2004–2008. Recreational landings include the Tar-Pamlico and Neuse rivers only.



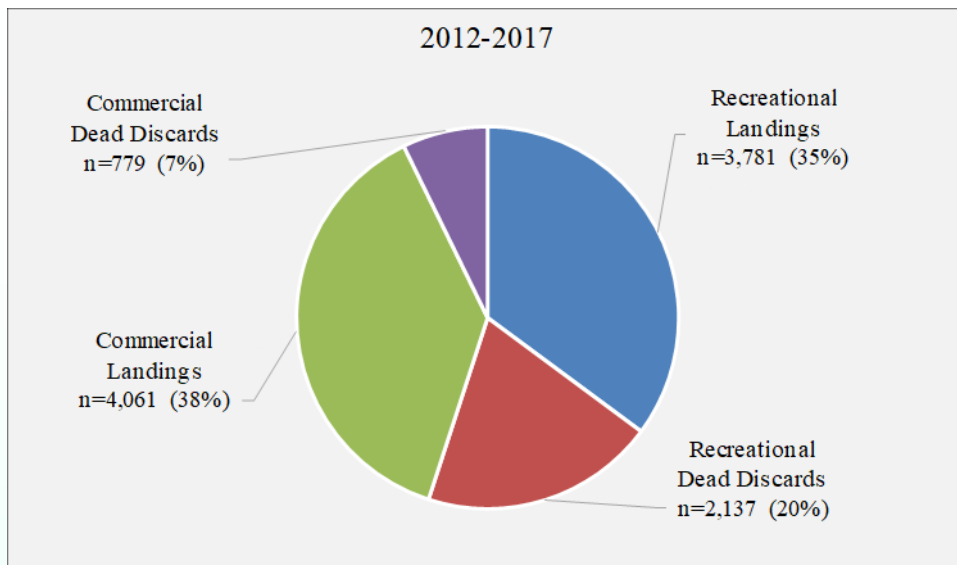


Figure 7. Average number of striped bass landed and discarded from the commercial and recreational fisheries in the Tar-Pamlico and Neuse rivers, 2012–2017.

## Habitat and Fish Stocks

With the important relationship between habitat and fish populations, the goal to protect and enhance habitats supporting coastal fisheries comes from the implementation of the Coastal Habitat Protection Plans (NCDEQ, 2016; CHPP, G.S. 143B-279.8). While much of the concern over declining fish stocks has been directed at overfishing, habitat loss and water quality degradation make a stock more susceptible to decline and may hinder stock recovery efforts. The CHPP is undergoing its mandated five-year review, with adoption planned for summer 2021. One of the priority issues, “Submerged Aquatic Vegetation (SAV) Protection and Restoration, with Focus on Water Quality Improvements” has implications for North Carolina striped bass stocks. SAV is especially sensitive to water quality impairment from nutrient and sediment pollution and has been considered a “coastal canary”, serving as a valuable bio-indicator of the overall health of coastal ecosystems (Stevenson, 1998). The primary mechanism to restore and sustain SAV is by improving water quality. The CHPP strategy for SAV involves modifying water quality criteria, such as chlorophyll a levels and nutrient standards to reduce nutrient loading, allowing increased light penetration that is critical for submerged vegetation. This will not only benefit SAV but address the algal blooms in the Albemarle Sound area and other poor water quality impacts to fish like striped bass. It is imperative the fishing community actively participate in the ongoing CHPP review and add their voice to support the actions outlined in the CHPP.



Algae Bloom, Chowan River, Bertie County.  
Photo By: DMF Staff

# Amendment 2 Management Strategies

## Albemarle-Roanoke Striped Bass Stock Sustainable Harvest:

### Background

Although this document is specific to the ongoing development of Amendment 2 to the N.C. Estuarine Striped Bass FMP, it is important to note under the existing Amendment 1 there is adaptive management language that states, “Should the target F be exceeded, then restrictive measures will be imposed to reduce F to the target level” (NCDMF 2013). Actions authorized in Amendment 1 are being considered to lower F to address sustainable harvest in the interim as Amendment 2 is completed. This action maintains compliance with Amendment 1 to the North Carolina Estuarine Striped Bass FMP and ASMFC’s Addendum IV to Amendment 6 to the Interstate FMP for Atlantic Striped Bass while the Amendment 2 sustainable harvest management strategy is developed.

Amendment 2 will focus on development of management strategies that address both the overfished and overfishing status of the A-R stock relative to the Fisheries Reform Act (FRA) of 1997, which states each plan “shall specify a time period, not to exceed two years from the date of the adoption of the plan, for ending overfishing...” and “specify a time period, not to exceed 10 years from the date of adoption of the plan, for achieving a sustainable harvest”. Projections from the terminal year of the stock assessment that model how SSB responds in the coming years to various levels of harvest are used to calculate a new TAL that will accomplish the dual mandate of the FRA. As shown in Figure 8, the actual level of recruitment occurring in future years is an important factor in the level of expected increase in SSB. Projections use multiple levels of recruitment to inform managers of the uncertainty associated with assumptions about future stock recruitment and the related increases in SSB.



Pictured: Jennifer Lewis



Tagging on the spawning grounds  
Roanoke River, Weldon NC. DMF staff

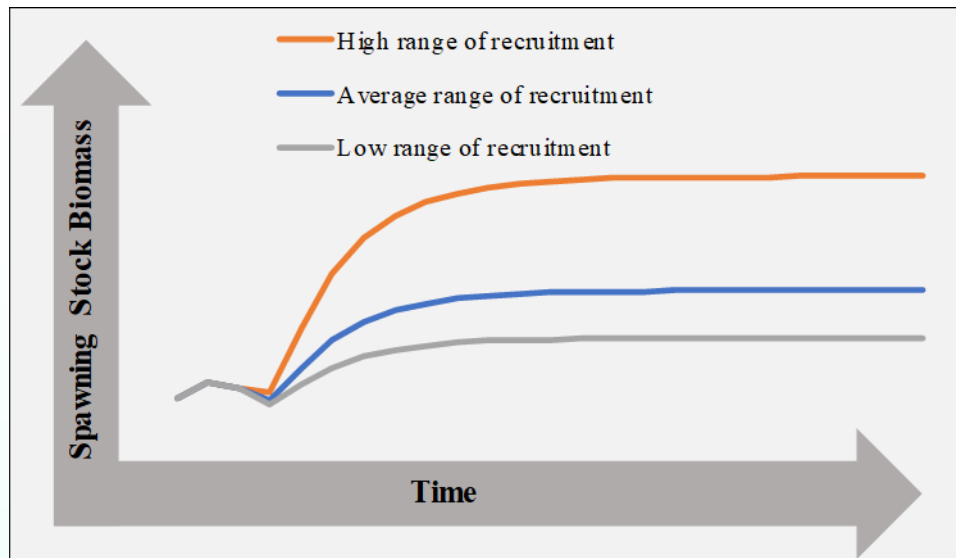


Figure 8. A graphical illustration of how assumptions about the level of future recruitment impacts stock projections of spawning stock biomass (SSB).

The necessary management measures currently in place in Amendment 1 to manage a TAL and prevent harvest from exceeding it each year include:

- adjust the TAL based on benchmark stock assessments and assessment updates
- daily quota monitoring of commercial harvest
- weekly quota monitoring of recreational harvest
- open and/or close harvest seasons to remain below the TAL
- authorize or restrict fishing methods and gear
- limit size, quantities taken or possessed (i.e., daily recreational creel limits and commercial limits)
- restrict fishing areas



Striped bass being tagged with commercial harvest tags  
Frog Island fish house Weeksville, NC  
Photo By: Chris Kelly



Pictured: Shane

### Questions for the Public

- Which of the existing management measures do you support to maintain harvest within limits of the specified TAL?
- In the event of a low TAL that restricts the regular harvest seasons, would you prefer a short season of consecutive harvest days or slightly longer season with only selected harvest days each week? Which harvest days would you prefer?
- Do you support investigating size limit changes for A-R striped bass?
- What recreational and/or commercial gear or area restrictions would you support to reduce discard mortality to rebuild the A-R stock?

# Tar-Pamlico, Neuse, and Cape Fear rivers striped bass stocks:

## Sustainable Harvest:

### Background

There has been a commercial and recreational no-possession provision in the Cape Fear River and its tributaries since 2008. This no-possession measure was implemented to help support specific goals of Amendment 1, which are to achieve sustainable harvest through science-based decision-making processes that conserves the resource. Prior to 2019, harvest in the CSMA was managed by commercial and recreational seasons, harvest and size limits, and gear restrictions, and constrained by an annual commercial TAL of 25,000 lb. Additionally, measures in Supplement A to Amendment 1 of the N.C. Estuarine Striped Bass FMP were implemented in March 2019 that implemented a no-possession provision in the commercial and recreational striped bass fisheries, as well as commercial set gill-net restrictions requiring tie-downs and distance from shore (DFS) measures to apply year-round, in the CSMA (NCDMF 2019). Supplement actions need to be contained within Amendment 2 management strategies in order to stay in effect.

Concurrent in timing but independent of the MFC's adoption of Supplement A is the MFC directed proclamation that prohibits the use of all gill-nets upstream of the ferry lines from the Bayview Ferry to Aurora Ferry on the Pamlico River and the Minnesott Beach Ferry to Cherry Branch Ferry on the Neuse River. As in this case when the commission enacted the provision to direct issuance of a proclamation, the fisheries director has no discretion to choose another management option and is bound by law to follow the commission decision. The MFC may alter this directive at any time or as part of Amendment 2, and if they choose not to do so, the proclamation actions remain in effect.

Harvest will be allowed if the no-possession measure in Supplement A is not continued in Amendment 2, and other management strategies should be considered to rebuild the stock. Possible stocking and fishery management strategies for CSMA striped bass were evaluated using a demographic matrix model (Mathes et al. 2020). Model results indicated CSMA striped bass populations are depressed to an extent that sustainability is unlikely at any level of fishing mortality. Lack of natural reproduction in CSMA systems requires continuous stocking to maintain the populations unless environmental and biological characteristics are improved.



NCSU graduate student surgically implanting a acoustic tracking tag in a striped bass to be stocked in the Neuse river. Photo By: USFWS.

Management strategies could be implemented to expand the age structure of the population and increase abundance of older fish which, given appropriate environmental conditions, may promote natural reproduction. Some environmental conditions can be addressed through the CHPP while biological characteristics can be addressed by altering stocking strategies including consideration of stocking fish better suited to environmental conditions in the CSMA. However, if management strategies implemented through Amendment 2 are unsuccessful at achieving sustainable harvest and external factors are deemed to make establishment of sustainable striped bass populations in CSMA systems impossible, other management strategies, including returning to a hatchery-supported fishery, could be considered in future Amendments.

If the no-harvest provision in the CSMA remains in place, adaptive management could be used to determine under what conditions the fishery could re-open. For example, collecting young-of-year striped bass in juvenile sampling would indicate successful natural reproduction, decreased contribution of stocked fish could potentially indicate successful recruitment, an increase in the number of older fish would indicate expansion of the age structure of the stock, and increased abundance in the independent surveys could indicate population growth. Conversely, adaptive management could also be used as a means to reconsider management strategies if establishment of self-sustaining populations in CSMA systems is determined to be unattainable.



DMF staff conducting Independent Gill Net Index of Abundance Survey  
Western Albemarle Sound

### *Questions for the Public*

#### No-Possession Provision – Amendment 1 (applicable to Cape Fear River) and Supplement A Management Measures

##### If the No-Possession Provision is Continued

- Do you support continuing the no-possession provision in the CSMA? For how long?
- If the no-possession provision remains, what gear modifications or restrictions should be considered to reduce bycatch and discards?
- Do you support continued stocking in the CSMA?

##### If the No-Possession Provision is Not Continued

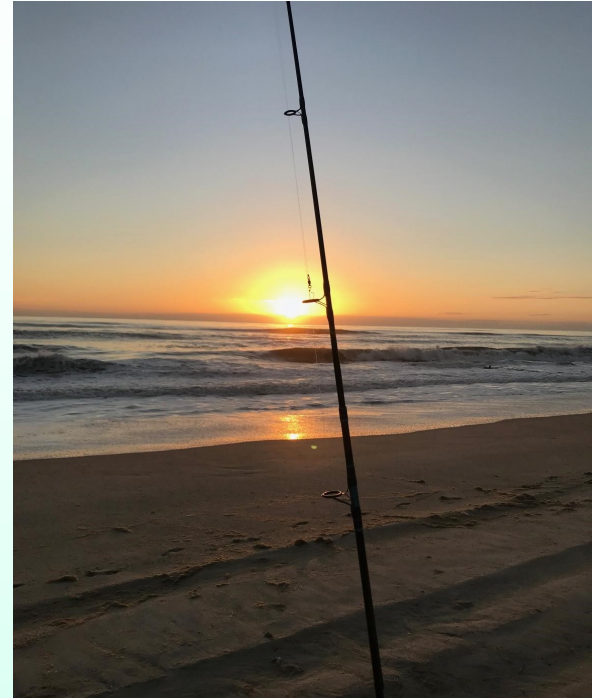
- What management measures should be considered to allow for sustainable harvest (i.e., TAL, closed and open harvest seasons, daily trip limits)?
- Do you support investigating size limit changes for CSMA striped bass?
- What gear modifications or restrictions should be considered to reduce bycatch and discards?
- Do you support continued stocking in the CSMA?

# Applicable to all North Carolina's Striped Bass stocks:

## Hook-and-line allowed as legal commercial gear in North Carolina's striped bass fisheries:

### Background

Amendment 1 to the N.C. Estuarine Striped Bass FMP included an issue paper discussing hook-and-line as a legal commercial gear in the ASMA and CSMA commercial striped bass fisheries. The result was a recommendation by the DMF and MFC to maintain status quo with adaptive management – (Do not allow hook-and-line as commercial gear in the estuarine striped bass fishery unless the use of traditional gears is prohibited). However, through development of the Amendment 1 and discussing the issue paper, the ACs and the DMF recognized that while allowing hook-and-line as a commercial gear could potentially have some positive impacts to the striped bass resource and stakeholders, there would need to be additional discussion of how to best implement the measure. Therefore, the rule that specifically prohibited the use of hook-and-line as a commercial gear was repealed and now that gear is prohibited as a commercial gear in the striped bass fishery through proclamation. If through development of Amendment 2 the MFC votes to allow hook-and-line as a commercial gear, the tools are already in place to implement the measure.



Recreational angling, Outer Banks N.C.  
Photo By: Rick Denton

### Questions for the Public

- Do you support hook-and-line as a legal commercial gear in the striped bass commercial fishery?



Photo By: Mitchell Blake



Pictured: DMF Staff



Pictured: Adam B. Cape Fear River, N.C.

## Questions for the Public about Potential Management Strategies



1. What management strategies already under consideration do you support for Amendment 2?
2. Are there other relevant strategies not included herein that should be consider for Amendment 2?

*Additional management strategies may be considered in Amendment 2 dependent on statutory requirements, available data, research needs, and the degree of impact the management strategy would have and how effective the solution would be. If the division determines a management strategy raised during the scoping period might have positive impacts on the stocks, additional examination of the strategy may be undertaken in the development of the FMP Amendment 2.*



Photo By: Adam B. Cape Fear River, N.C.

## Literature Cited

ASMFC. 2003. Amendment # 6 to the interstate fishery management plan for Atlantic striped bass. ASMFC, Fisheries Management Report No. 41, Washington, DC.

NCDEQ (North Carolina Department of Environmental Quality) 2016. North Carolina Coastal habitat Protection Plan. Morehead City, NC. Division of Marine Fisheries. 33 p.

Cushman, B., T. O'Donnell, and D. Farrae. 2018. South Carolina Department of Natural Resources. 2017 Striped Bass Genotyping and Parentage Analysis Final Report for the North Carolina Wildlife Resources Commission. 39 pp.

Farrae, D., and T. Darden. 2018. South Carolina Department of Natural Resources. 2017 Striped Bass Genotyping Report for the North Carolina Division of Marine Fisheries. 9 pp.

Lee, L.M., T.D. Tears, Y. Li, S. Darsee, and C. Godwin (editors). 2020. Assessment of the Albemarle Sound -Roanoke River striped bass (*Morone saxatilis*) in North Carolina, 1991–2017. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2020-01, Morehead City, North Carolina. 171 p.

Mathes, T., Y. Li, T. Tears, and L.M. Lee (editors). 2020. Central Southern Management Area striped bass stocks in North Carolina, 2020. North Carolina Division of Marine Fisheries, NCDMF SAP-SAR-2020-02, Morehead City, North Carolina. 161 p. + appendices.

Nelson, K.L. 1998. Catch-and-release mortality of striped bass in the Roanoke River, North Carolina. *North American Journal of Fisheries Management* 18:25–30.

NCDMF. 2013. Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environment and Natural Resources. North Carolina Division of Marine Fisheries. Morehead City, NC. 826 pp.

NCDMF. 2019. Supplement A to Amendment 1 to the North Carolina estuarine striped bass fishery management plan. North Carolina Department of Environment and Natural Resources, North Carolina Division of Marine Fisheries, Morehead City, North Carolina. 40 p.

Stevenson, J. C. 1988. Comparative Ecology of Submersed Grass Beds in Freshwater, Estuarine and Marine Environments. *Limnology and Oceanography* 33: 867–893.



Photo By: K. D. Hewitt



Wild young-of-year striped bass,  
DMF juvenile striped bass index  
of abundance survey





Recreational anglers, Roanoke River, Weldon, N.C.

## Scoping Document

Management Strategies for Amendment 2  
to the N. C. Estuarine Striped Bass  
Fishery Management Plan

### **NORTH CAROLINA DIVISION OF MARINE FISHERIES**

DMF Headquarters  
3441 Arendell Street  
PO Box 769  
Morehead City, NC 28557

800-682-2632  
252-726-7021

<http://portal.ncdenr.org/web/mf/home>

